

Enzyme immobilised novel core-shell superparamagnetic nanocomposites for enantioselective formation of 4-(*R*)-hydroxycyclopent-2-en-1-(*S*)-acetate†

Cite this: DOI: 10.1039/c4cc05358c

Received 11th July 2014,
Accepted 6th August 2014

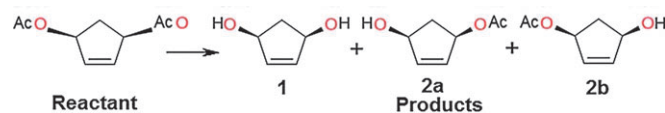
DOI: 10.1039/c4cc05358c

www.rsc.org/chemcomm

Maneea Eizadi Sharifabad,^{ab} Ben Hodgson,^{ab} Mourad Jellite,^{ab} Tim Mercer^{ac} and Tapas Sen^{ab}

Lipase immobilized novel high surface area core-shell superparamagnetic nanoparticles have been fabricated and used as efficient reusable catalysts for the selective production of pharmaceutically important chiral isomers from *meso*-cyclopent-2-en-1,4-diacetate.

Enzymes are well known as catalysts for various bio-transformations however, it is difficult to separate the expensive enzymes from the reaction mixtures. Hence immobilization of enzymes on solid supports such as zeolites,¹ agarose beads,² epoxy resins³ and porous glass⁴ is an important step for the recovery and recycling of the enzymes. Immobilization of lipase in magnetisable solid supports has recently been exploited^{5,6} for the ester hydrolysis due to an additional advantage of easy recovery of expensive enzymes from the reaction mixture by a single step separation under the influence of an external magnetic field. The use of immobilised enzymes is particularly important as they exhibit increased stability and catalytic activity compared to free enzymes.⁷ Immobilization of an enzyme is important as that leads to the enzyme being dispersed over a large surface area, enhancing mass transfer and preventing aggregation.⁸ Catalytic hydrolysis of *meso*-cyclopent-2-en-1,4 diacetate has been reported previously using *Mucor* sp.,⁹ pancreatin¹⁰ and *Mucor miehei* lipase (Chirazyme),¹¹ however recovery of enzymes from the reaction mixture was a great concern and drawback to the technology. Herein, we report (a) fabrication of a high surface area novel core-shell superparamagnetic nanoparticles (b) surface modification of core-shell nanoparticles with aminopropyl tri-ethoxy silane by recently



Scheme 1 Enzymatic hydrolysis of *meso*-cyclopent-2-en-1,4-diacetate to chiral products (**2a** and **2b**).

developed Tri-phasic Reverse Emulsion (TPRE)¹² method for the immobilisation of *Pseudomonas Fluorescens Lipase* (PFL) and *Candida Rugosa Lipase* (CRL) by covalent coupling (c) one step catalytic conversion (see reaction Scheme 1) of *meso*-cyclopent-2-en-1,4 diacetate to important optical isomers such as 4-(*R*)-hydroxycyclopent-2-en-1-(*S*)-acetate and 4-(*S*)-hydroxycyclopent-2-en-1-(*R*)-acetate by the careful selection of a suitable lipase enzyme with a single step separation of the expensive catalyst by magnetic separation.

Superparamagnetic iron oxide nanoparticles (SPIONs) with narrow particle size distribution with an average diameter of 10 nm were synthesized by a co-precipitation method (see ESI†) of ferrous and ferric chloride salts following our previously reported protocol.¹³ SPIONs were then coated with a thick mesoporous silica shell using tetraethyl orthosilicate (TEOS) as a silica source, cetyltrimethyl ammonium bromide (CTAB) as a structure directing template in an alkaline medium (NH₄OH) by a modified protocol (see details in ESI†) from our earlier report.¹⁴

Fig. 1a presents the TEM image of the core-shell nanoparticles of around 270 nm in diameter where the core was formed with multiple SPIONs of each approximately 10 nm diameter and mesoporous silica shell (thickness ~ 135 nm) with mesopore diameter of approximately 3.5 nm. Small Angle X-ray Scattering (SAXS) data (see Fig. 1b and ESI,† Fig. S1) indicate the presence of mesopores of repeating distance of 3.7 nm which is similar to the mesopore diameter measured using TEM (Fig. 1a and see ESI,† Fig. S2) and nitrogen gas adsorption (Fig. 1c). The XRD pattern (see Fig. S3 in ESI†) of core iron oxide nanoparticles indicated the magnetite (Fe₃O₄) phase due to the characteristic finger print patterns. N₂ gas adsorption analysis (Fig. 1c) shows a typical type IV isotherm with a hysteresis loop confirming the mesoporous structure.

^a Centre for Materials Science, School of Forensic and Investigative Sciences,

University of Central Lancashire, Preston, PR1 2HE, UK

^b Surface Patterning Group, Institute of Nanotechnology and Bioengineering,

University of Central Lancashire, Preston, PR1 2HE, UK

^c School of Computing and Physical Sciences, University of Central Lancashire, Preston, PR1 2HE, UK

† Electronic supplementary information (ESI) available: Detail experimental protocol for the synthesis of novel core-shell nanoparticles, surface engineering of the core-shell nanoparticles by TPRE method, enzyme immobilization by covalent coupling, catalysis reaction condition including the product analysis using Gas Chromatographic and Mass Spectroscopy (GCMS) along with nitrogen gas adsorption data. See DOI: 10.1039/c4cc05358c

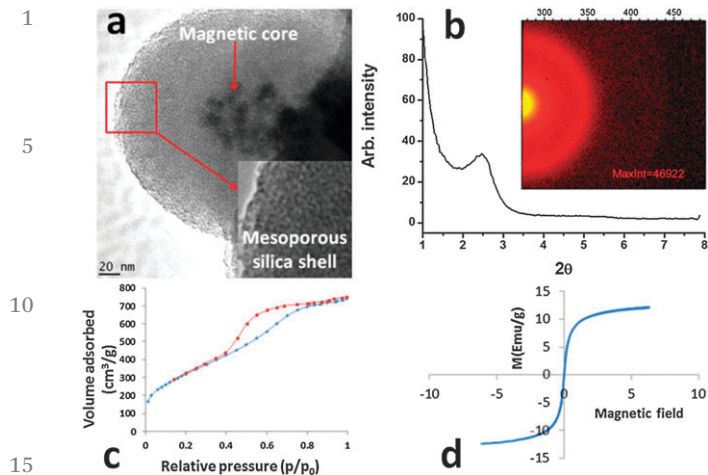


Fig. 1 TEM image (a), small angle X-ray scattering (b), N_2 adsorption-desorption isotherm (c) and magnetic measurement (d) of novel core-shell mesoporous superparamagnetic nanoparticles.

The BJH adsorption average pore diameter (see ESI†, pages S9–S13) of approximately 4 nm and the BET surface area was calculated to be $1187.8 \text{ m}^2 \text{ g}^{-1}$. The exceptional high surface area compared to our previously reported core-shell mesoporous silica-magnetite^{14,15} is the key in our study as a novel catalytic support. Fig. 1d presents the magnetic data using variable sample magnetometer (VSM). Zero remanence ($M_R = 0$) is an indication of superparamagnetism with low saturation magnetization value of approximately 12 emu g^{-1} . The low saturation magnetization value compared to a large value ($\sim 80 \text{ emu g}^{-1}$) of pure superparamagnetic magnetite nanoparticles can be due to the presence of a thick diamagnetic silica shell similar to our earlier report.¹⁵

Surface functionalisation of the core-shell nanoparticles with hydrophilic silica surface was carried out using aminopropyl triethoxy silane using novel TPPE method¹² (see ESI† for details) in order to create uniform layer of terminal $-\text{NH}_2$ functional groups. The $-\text{NH}_2$ groups were further converted to terminal aldehyde ($-\text{CHO}$) groups by reacting with glutaraldehyde solution in SSC buffer as reported earlier.^{6,12} The surface $-\text{CHO}$ modified nanoparticles were covalently coupled (see ESI† for experimental protocol) with $-\text{NH}_2$ groups of lipase (PFL and CRL) enzymes. The reaction scheme for the immobilisation of lipase with surface $-\text{CHO}$ groups has been reported⁶ earlier by our group using a different materials *i.e.* hierarchically ordered porous solid (HOPS).

The lipase immobilised core-shell nanoparticles and free lipases were used for the enzymatic hydrolysis of *meso*-cyclopent-2-en-1,4-diacetate under a biphasic reaction condition using a solvent mixture (80% hexane and 20% water) at 25°C with end-over-end rotation (40 rpm) for up to 48 hours reaction time (see ESI† for experimental details). A biphasic solvent was used in our experiment as the products are soluble in water and insoluble in hexane whereas the reactant is mostly soluble in hexane. Recently our group has established¹⁶ that a bi-phasic system is powerful compared to a single phase system for trans-esterification reaction using free lipase as catalyst. The activity of free and immobilised lipases in the conversion of *meso*-cyclopent-2-en-1,4-diacetate (reactant) and the

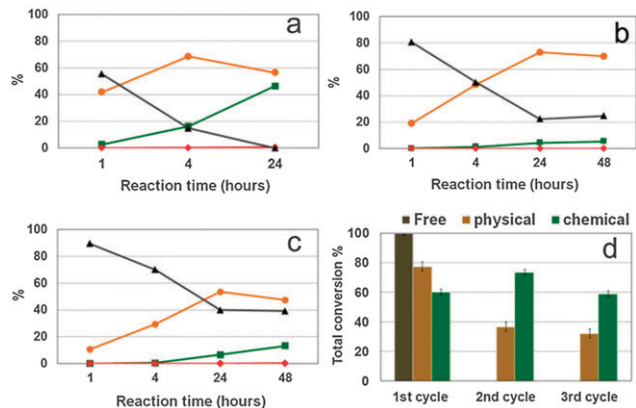


Fig. 2 Conversion of reactant and the formation of products in % at different reaction times by enzymatic hydrolysis by using free PFL (a), physically adsorbed PFL on core-shell nanoparticles (b), PFL immobilised on core-shell nanoparticles by covalent coupling via TPPE approach (c), and % conversion of reactant during the reusability test (d).

formation of products (see reaction Scheme 1) were determined by GC and GCMS analysis using a pre-established calibration curves (see ESI†, Fig. S4 to S7).

Fig. 2 and 3(a to c) present the percentage (%) of unreacted *meso*-cyclopent-2-en-1,4-diacetate (black lines) and the corresponding products such as 4-(*R*)-hydroxycyclopent-2-en-1-(*S*)-acetate (orange), 4-(*S*)-hydroxycyclopent-2-en-1-(*R*)-acetate (red) and cyclopent-2-en-1,4-diol (green) at different time intervals (1, 4, 24 and 48 h) by enzymatic hydrolysis using PFL (Fig. 2) and CRL (Fig. 3). Fig. 2d and 3d exhibit the percentage conversion of reactant during the reusability test (cycles 1 to 3). Free enzyme couldn't be recovered from the reaction mixture hence presented as one cycle (grey bar in Fig. 2d and 3d).

Percentage (%) conversion of reactant to various products increases with increasing the reaction times from 1 to 24 h for all materials using free enzymes or enzymes immobilised nanoparticles. Free PFL performed a complete conversion ($100 \mu\text{mol}$ of reactant per mg of enzyme) of reactant to products

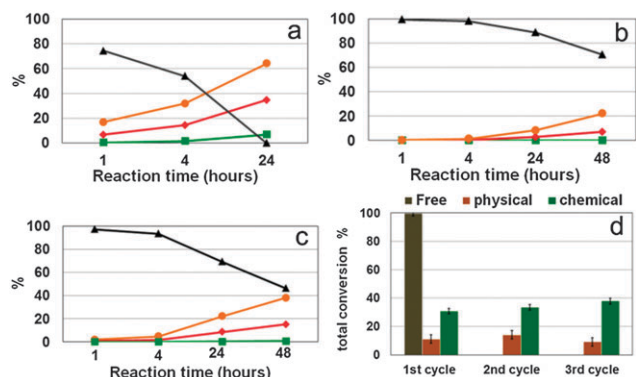


Fig. 3 Conversion of reactant and the formation of products in % at different reaction times by enzymatic hydrolysis using free CRL (a), physically adsorbed CRL on core-shell nanoparticles (b), CRL immobilised on core-shell nanoparticles by covalent coupling via TPPE approach, and % conversion of reactant during the reusability test using three different materials (d).

1 after 24 h of reaction whereas PFL immobilized (physically or
chemically) nanoparticles performed a partial conversion. Physically
adsorbed PFL materials performed around 78% conversion of
reactant (78 μmol of reactant per mg of enzyme) whereas chemically
5 immobilized PFL materials performed around 60% conversion of
reactant (60 μmol of reactant per mg of enzyme). It is important to
mention that free PFL or PFL immobilized (physically or chemically)
nanoparticles produced only 4-(*R*)-hydroxycyclopent-2-en-1-(*S*)-
acetate (**2a** of Scheme 1) without the formation of 4-(*S*)-hydroxycyclopent-2-
10 en-1-(*R*)-acetate (**2b** of Scheme 1). Increasing the reaction times from
1 to 24 h, the concentration of 4-(*R*)-hydroxycyclopent-2-en-1-(*S*)-
acetate decreases and the concentration of unwanted cyclopent-2-
en-1,4-diol (see Scheme 1, product **1**) increases due to the further
hydrolysis of acetate group to alcohol group. Physically adsorbed PFL
15 nanoparticles performed better in catalytic conversion of reactant to
desired product 4-(*R*)-hydroxycyclopent-2-en-1-(*S*)-acetate, however
when re-used the catalyst, the catalytic conversion reduced more
than 50% in the 2nd and 3rd cycles whereas chemically immobilized
PFL exhibited a stable catalytic activity. The higher catalytic activity of
20 physically adsorbed PFL in the 1st cycle could be due to the presence
of free enzymes in the reaction mixture by partial leaching from the
nanoparticles.

When CRL (free and immobilised forms) was used as an enzyme,
both optical isomers were formed (see Fig. 3). The ratio of 4-(*R*)-
25 hydroxycyclopent-2-en-1-(*S*)-acetate to 4-(*S*)-hydroxycyclopent-2-en-1-
(*R*)-acetate in the product mixture was around 2:1. Free CRL
exhibited the highest % conversion of reactant whereas physically
adsorbed CRL performed the lowest conversion. In conclusion, CRL
is catalytically less active compared to PFL but suitable for the
30 desymmetrisation of both acetoxy groups of the reactant in an
approximately 2:1 ratio.

The reactant *meso*-cyclopent-2-en-1,4-diacetate can form a
tetrahedral intermediate complex with the catalytic triad of the
lipase active site (Asp/Glu-His-Ser)¹⁷ and the mono-acetoxy
35 products (see Scheme S1 in ESI[†]) are hydrolyzed from the active
site. The hydrolysed acetoxy group from the diacetoxy reactant
remains bound to the serine residue in the active site as an
acyl-enzyme intermediate complex. After the acyl-enzyme
intermediate is formed, water hydrolyses the acetoxy group,
40 forming acetic acid in the reaction mixture, which enables the
acid-catalysed hydrolysis of the mono-acetoxy product **2a** to the
dihydroxy by-product **1** when the enzyme is PFL.

In summary, high surface area novel mesoporous core-shell
superparamagnetic nanoparticles have been fabricated and
45 used as a support for the immobilisation of two different

lipase enzymes *via* recently reported¹⁸ TPRES method. The
materials have been tested for one step formation of pharmaceu-
tically important optimal isomers 4-(*R*)-hydroxycyclopent-2-en-1-(*S*)-
acetate and 4-(*S*)-hydroxycyclopent-2-en-1-(*R*)-acetate by enzymatic
5 hydrolysis of *meso*-cyclopent-2-en-1,4-diacetate. It is important to
mention that enzyme PFL produced an optically pure enantiomer
4-(*R*)-hydroxycyclopent-2-en-1-(*S*)-acetate whereas CRL produced
both optical isomers (see Table S2 in ESI[†] for enantiomeric excess
value). Hence the combination of a suitable enzyme and a suitable
10 superparamagnetic support can be an important step for the
industrial production of pharmaceutically important optical iso-
mers with a single step separation of expensive enzymes from the
reaction mixture by an external magnetic field.

The authors thank to the Royal Society of Chemistry small
research funds (2010–2011 and 2011–2012) to carry out the
15 chiral reaction using a chiral column. MES thank to the
University of Central Lancashire for partial support on her
PhD scholarship.

Notes and references

- 1 G.-W. Xing, X.-W. Li, G.-L. Tian and Y.-H. Ye, *Tetrahedron*, 2000, **56**, 3517–3522.
- 2 P. W. Tardioli, R. Fernández-Lafuente, J. M. Guisán and R. L. C. Giordano, *Biotechnol. Prog.*, 2003, **19**, 565–574.
- 3 K. Hernandez and R. Fernandez-Lafuente, *Process Biochem.*, 2011, **46**, 873–878.
- 4 E. Karakuş and Ş. Pekyardımcı, *J. Mol. Catal. B: Enzym.*, 2009, **56**, 13–19.
- 5 A. Dyal, K. Loos, M. Noto, S. W. Chang, C. Spagnoli, K. V. P. M. Shafi, A. Ulman, M. Cowman and R. A. Gross, *J. Am. Chem. Soc.*, 2003, **125**, 1684–1685.
- 6 T. Sen, I. J. Bruce and T. Mercer, *Chem. Commun.*, 2010, **46**, 6807.
- 7 L. Costa, V. Brissos, F. Lemos, F. R. Ribeiro and J. M. S. Cabral, *Bioprocess Biosyst. Eng.*, 2008, **31**, 323–327.
- 8 K. Nagayama, N. Yamasaki and M. Imai, *Biochem. Eng. J.*, 2002, **12**, 231–236.
- 9 F. Theil, H. Schick, D. Weichert, K. Tannenberger and G. Klappach, *J. Prakt. Chem.*, 1991, **333**, 497–499.
- 10 F. Theil, H. Schick, M. A. Lapitskaya and K. K. Pivnitsky, *Liebigs Ann. Chem.*, 1991, **1991**, 195–200.
- 11 S. R. Ghorpade, R. K. Kharul, R. R. Joshi, U. R. Kalkote and T. Ravindranathan, *Tetrahedron: Asymmetry*, 1999, **10**, 891–899.
- 12 T. Sen and I. J. Bruce, *Sci. Rep.*, 2012, **2**, DOI: 10.1038/srep00564.
- 13 T. Sen, S. Magdassi, G. Nizri and I. J. Bruce, *Micro Nano Lett.*, 2006, **1**, 39.
- 14 T. Sen, A. Sebastianelli and I. J. Bruce, *J. Am. Chem. Soc.*, 2006, **128**, 7130–7131.
- 15 T. Sen and I. J. Bruce, *Microporous Mesoporous Mater.*, 2009, **120**, 246–251.
- 16 B. Hodgson, Peer-reviewed PhD thesis, University of Central Lancashire, Preston, UK, 2014.
- 17 S. Raza, L. Fransson and K. Hult, *Protein Sci.*, 2001, **10**, 329–338.