

Central Lancashire Online Knowledge (CLoK)

Title	Could Upregulated Hsp70 Protein Compensate for the Hsp90-Silence-Induced Cell Death in Glioma Cells?
Type	Article
URL	https://clock.uclan.ac.uk/id/eprint/13834/
DOI	https://doi.org/10.1155/2014/652643
Date	2014
Citation	Munje, Chinmay, Shervington, Leroy Alexander, Khan, Zarine Jamsher and Shervington, Amal A (2014) Could Upregulated Hsp70 Protein Compensate for the Hsp90-Silence-Induced Cell Death in Glioma Cells? International Journal of Brain Science, 2014. ISSN 2356-6965
Creators	Munje, Chinmay, Shervington, Leroy Alexander, Khan, Zarine Jamsher and Shervington, Amal A

It is advisable to refer to the publisher's version if you intend to cite from the work.
<https://doi.org/10.1155/2014/652643>

For information about Research at UCLan please go to <http://www.uclan.ac.uk/research/>

All outputs in CLoK are protected by Intellectual Property Rights law, including Copyright law. Copyright, IPR and Moral Rights for the works on this site are retained by the individual authors and/or other copyright owners. Terms and conditions for use of this material are defined in the <http://clock.uclan.ac.uk/policies/>

Research Article

Could Upregulated Hsp70 Protein Compensate for the Hsp90-Silence-Induced Cell Death in Glioma Cells?

Chinmay Munje, Leroy Shervington, Zarine Khan, and Amal Shervington

Brain Tumour North West, Faculty of Science and Technology, University of Central Lancashire, Preston PR1 2HE, UK

Correspondence should be addressed to Amal Shervington; aashervington@gmail.com

Received 13 May 2013; Accepted 19 September 2013; Published 2 January 2014

Academic Editor: Toshio Matsuda

Copyright © 2014 Chinmay Munje et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The molecular chaperone heat shock protein 90 alpha (Hsp90 α) has been recognized in various tumours including glioma. This pilot study using a proteomic approach analyses the downstream effects of Hsp90 inhibition using 17-allylamino-17-demethoxygeldanamycin (17AAG) and a short hairpin RNA (shRNA) oligonucleotide targeting *hsp90 α* (sh*hsp90 α*) in the U87-MG glioma cell line. Preliminary data coupled with bioinformatic analysis identified several known and unknown Hsp90 client proteins that demonstrated a change in their protein expression after Hsp90 inhibition, signifying an alteration in the canonical pathways of cell cycle progression, apoptosis, cell invasion, angiogenesis, and metastasis. Members of the glycolysis pathway were upregulated, demonstrating increased dependency on glycolysis for energy source by the treated glioma cells. Upregulated proteins also include Hsp70 and members of its family such as Hsp27 and gp96, thereby suggesting the role of Hsp90 co-chaperones in compensating for Hsp90 function after Hsp90 inhibition. Considering Hsp70's role in antiapoptosis, it was postulated that a combination therapy involving a multitarget approach could be carried out. Consequently inhibition of both Hsp90 and Hsp70 in U87-MG glioma cells resulted in 60% cell death indicating the importance of combination therapy for glioma therapeutics.

1. Introduction

Heat shock protein Hsp90 is upregulated in several tumours including glioma, and thus targeting its function may provide new therapeutic aspects [1, 2]. Hsp90 is a highly conserved molecular chaperone present in eukaryotic cytosol. It has been proposed to play a vital role in tumorigenesis, maintenance of transformation and regulation of several key proteins involved in apoptosis and survival and growth pathways [3]. These pathways are exploited in tumours where Hsp90 chaperoning contributes towards drug resistance [4], metastasis [5], and cell survival [6]. The synthesis of several natural and chemical inhibitors along with RNA inhibition using siRNA or shRNA to silence Hsp90 has been undertaken.

Previous studies in our laboratory have shown that enhanced chemosensitivity is attained upon transcription inhibition of the inducible Hsp90 subunit *hsp90 α* by siRNA, suggesting that inhibiting *hsp90 α* expression by shRNA could possibly be a favourable therapeutic approach compared to conventional chemotherapies as it is target specific and has reduced toxicity. Furthermore, a combination treatment of

siRNA followed by Temozolomide (TMZ) (200–400 μ M) after 48 hours was significantly more effective, suggesting the combination as a possible effective form of therapy [7]. Given its functions as a molecular chaperone and its expression in gliomas, *hsp90 α* may represent a promising target for the development of novel therapeutic strategies. This study investigated Hsp90 chaperone inhibition with (a) 17-allylamino-17-demethoxygeldanamycin (17AAG), an analogue of Geldanamycin (GA) and a potent Hsp90 inhibitor and (b) short hairpin RNA (shRNA) oligonucleotide targeted against *hsp90 α* .

In this study, proteins were extracted from the U87-MG cell lysate to characterize the changes transpired after Hsp90 inhibition with 17AAG and shRNA (*hsp90 α*) using a differential proteomic analysis. Proteins expressed in wild-type U87-MG cells (control) were compared with the proteins expressed in U87-MG cells upon subsequent silencing of *hsp90 α* (U87-MG-sh*hsp90 α*) and the proteins expressed in U87-MG cells after 17AAG treatment (U87-MG-17AAG).

This approach elucidates the changes caused in the Hsp90 chaperone system after inhibition using 17AAG or

shRNA oligonucleotide targeting *hsp90α* in glioma. Though the significance of Hsp90's glioma regulation has been well documented, the downstream effect of Hsp90 inhibition at the various physiological and signalling pathways in glioma is still ambiguous. Silencing *hsp90α* at the genetic level (shRNA) and at the protein level (17AAG) to identify potential downstream pathways and proteins affected and/or controlled by Hsp90 in glioma is a novel approach. Therefore, this pilot study aims to silence Hsp90 by using 17AAG and shRNA targeting *hsp90α* and attempts to identify the potential pathways and proteins altered in glioma.

This study also explores the role of Hsp70 as a co-chaperon for Hsp90. Hsp70 is the most predominant and conserved class of the Hsps [8], and it functions as an antiapoptotic protein [9]. It forms a multichaperone complex with Hsp90 and is involved in folding and refolding of newly synthesized or misfolded proteins [9, 10]. Hsp70 with Hsp90 plays vital roles in tumour immunity by preventing apoptosis and regulating the generation of stable complexes with cytoplasmic tumour antigens bestowing antitumour immunity [11, 12]. In tumours, Hsp70 is upregulated and plays a key role in the regulation of several pathways such as cell proliferation, metastasis, invasion, and death [13]. The over expression of Hsp70 has been associated with poor prognosis, and reduced response to tumour therapeutics [14]. Members of the Hsp70 family act at multiple points in the apoptotic pathway and inhibit cell death [15–17].

2. Materials and Methods

2.1. Cell Culture. The human brain cell line U87-MG (glioblastoma) used in this study was purchased from European Collection of Cell Cultures (ECCAC, UK). U87-MG was cultured in Eagle's minimum essential medium (EMEM) (Sigma, UK) and was supplemented with 10% (v/v) foetal bovine serum (Gibco-BRL, UK) and 2 mM L-glutamine (Sigma, UK). Cells were cultivated in 25 or 75 cm² culture-treated polystyrene flasks and were maintained at 37°C with 5% CO₂ in filtered air. Cells were closely monitored and when a monolayer growth of 70–80% confluence was obtained, the cells were scraped and subcultured.

2.2. shRNA Transfection. Purified and sequence verified pGFP-V-RS expression plasmids with *hsp90α* specific shRNA cassettes were obtained from Origene, USA. The sequence of the shRNA verified to silence *hsp90α* was shRNA (NM_005348; targeted exon 10 at nucleotides 2137–2166 bp) (sense) 5'-CTCTCAAGGACTACTGCACCAGA3' (anti-sense) 5'-GAGAGTTCCTGATGACGTGGTCTTACTTC3'. The shRNA plasmids were diluted with 50 μL of dH₂O to get a final concentration of 100 ng/μL, from which 1 μg of the shRNA expression plasmid DNA was then transfected into the cells according to the manufacturer's protocol using MegaTran 1.0 solution. The cells were then incubated in a 5% CO₂ incubator for 48 hours before being used for further analysis.

2.3. 17AAG and KNK437 Treatment. 17AAG (InvivoGen, USA) and KNK437 (Merck Bioscience, UK) were solubilised in DMSO (Sigma-Aldrich, UK). U87-MG cells were treated for 48 hours with inhibitory concentration (IC₅₀) values of 17AAG (0.25 μM) and KNK437 (55 nM) (unpublished results) and then with a combination of IC₅₀ values of 17AAG and KNK437. The relative IC₅₀ values were determined using point to point analysis. CellTitre-Glo one solution assay (Promega, UK) was used to determine the percentage of viable cells present in culture. The assay quantifies the level of the present ATP which is indicative of the presence of metabolically active cells. The raw data of the luminescence units were plotted against the different drug(s) concentration used, and regression analysis was used to determine the IC₅₀ of the drug(s).

2.4. Akt/PKB Kinase Activity. The Akt/PKB kinase activity was assayed using a solid-phase enzyme-linked immune-absorbent assay kit (Assay Deigns, UK) that detects the Akt/PKB activity in the solution phase. Proteins were extracted from the control cells (U87-MG Wild-Type) and treated cells (U87-MG-17AAG; U87-MG-sh*hsp90α*) using CellLytic MT mammalian tissue lysis buffer (Sigma, UK). Bradford's assay was used to determine the protein concentration and, 1 μg of protein from each sample was used to perform the Akt/PKB kinase activity assay following the manufacturer's protocol.

2.5. Flow Cytometry. The control and treated cells (>1 × 10⁶ cells) were collected by scraping. The cells were washed 3 times with 0.1% bovine serum albumin (BSA) made in 0.1 M phosphate buffered saline (PBS). After each wash, the cells were centrifuged at 1000 rpm for 5 min at 4°C, and the supernatant was discarded. The cells were permeabilised by incubating with 0.1% of 0.1 M Triton X-100 in PBS for 15 min in the dark followed by 3 washes with 0.1% BSA. The blocking solution (made up with 5% rabbit serum in 0.1 M PBS) was added to the cells and incubated for 30 min at 4°C. The cells were centrifuged and the blocking solution was discarded, and Hsp90α specific primary antibody, (rat monoclonal IgG2a (1:50) made up in 5% rabbit serum with 0.1 M PBS) was added and incubated for 30 min. The primary antibody was removed after 3 washes with 0.1% BSA. Hsp90α secondary antibody {rabbit anti-rat IgG conjugated with fluorescein isothiocyanate (FITC) (1:130)} prepared similar to the primary antibody preparation was added to the cell sample and incubated in the dark for 30 min. The negative control was prepared by excluding the labelling with primary antibody. The cells were washed 3 times and resuspended in 0.1% BSA. The cell suspension was filtered to remove clumps using a sterile filter. The samples were stored at 4°C in the dark until being analysed on the flow cytometer. Throughout the experimental process the samples and the reagents were kept on ice to minimize cell death. The samples during each wash were gently pipetted to achieve a single cell suspension.

2.6. Proteomic Analysis. Two-dimensional difference gel electrophoresis (2D-DIGE) and protein identification (ID)

were performed by Applied Biomix, Inc (Hayward, CA). Both the control and treated cell pellets were obtained by scraping, and the culture medium was removed by washing the pellets 3 times with washing buffer (10 mM Tris-HCl, 5 mM magnesium acetate, pH 8.0). Also, 200 μ L of 2D cell lysis buffer (30 mM Tris-HCl, pH 8.8, containing 7 M urea, 2 M thiourea, and 4% CHAPS) was added to 10 mg of cultured cell pellet. The mixture was sonicated at 4°C followed by shaking for 30 min at room temperature and then centrifuged for 30 min at 14000 rpm. The supernatant was collected, and Bio-Rad protein assay method was used to measure the protein concentration.

The protein extracts (~25–50 μ g) from the control and treated U87-MG cell samples were subjected to labelling using CyDye DIGE fluors (Cy2, Cy3 or Cy5). Proteins were separated using two-dimensional polyacrylamide gel electrophoresis (2D-PAGE) under denaturing conditions. The gels were stained and scanned using Typhoon TRIO (Amersham BioSciences, US) following protein separation. Differential regulation amidst the proteins was analyzed using Image Quant software (Amersham Bioscience, USA) by merging the images obtained. DeCyder analysis was performed for automated detection, background subtraction, quantitation, normalization, and intergel matching and spot picking. Protein identification was carried out using a mass spectrometer, in particular Matrix-assisted laser desorption/ionization-time of flight (MALDI-TOF). The peptide masses derived from the mass spectrometric analysis were matched with peptide fingerprints of known proteins in a protein sequence database using search engine MASCOT 2.0 (Matrix Science, US).

2.7. Statistical Analysis. All values were expressed as mean $\pm$ standard deviation for three independent experiments. Student's *t*-test and paired *t*-test were used to test the difference between the samples. Values of significance between **P* < 0.05 and ***P* < 0.001 were considered to be statistically significant.

3. Results

Hsp90 α inhibition was initially compared by studying the protein level of Hsp90 α in the control (wild-type U87-MG cells) and the treated cells with shhsp90 α and 17AAG referred to as U87-MG-shhsp90 α and U87-MG-17AAG, respectively, using fluorescence-activated cell sorting (FACS) analysis with a flow cytometer (Figure 1 and Table 1). These results demonstrate that both treatments can significantly inhibit the expression of Hsp90 α in the U87-MG glioma cell line (***P* < 0.001). The results also demonstrate that 17AAG works more effectively than shRNA.

Previous studies suggested the importance of Hsp90 in the maintenance of AKT kinase activity [18]; thus correlating the Hsp90 client protein Akt/PKB kinase activity levels is indicative to the activity of Hsp90, determined using the Akt/PKB kinase activity assay kit.

Akt/PKB kinase activity assay kit was used to measure the activity of Akt/PKB kinase in the control (wild-type U87-MG

TABLE 1: Hsp90 regulation level after silencing in U87-MG cells.

Samples	Kinase activity for 1 μ g Protein	Hsp90 α expression [%]
U87-MG (Control)	234.51 $\pm$ 4	76 $\pm$ 0.7
U87-MG-17AAG	44.53 $\pm$ 0.1**	42.7 $\pm$ 0.2**
U87-MG-shRNA hsp90 α	95.16 $\pm$ 1.9**	64.1 $\pm$ 1.4**

Data values are mean $\pm$ standard deviation, *n* = 3; **P* < 0.05 and ***P* < 0.001 are considered to be statistically significant, with * being significant and ** being very significant.

Note: Hsp90 protein levels were determined by FACS analysis, and Akt/PKB kinase activities were measured in the control (wild-type U87-MG cells), treated cells (U87-MG-shhsp90 α and U87-MG-17AAG), and its activity.

cells) and treated cells (U87-MG-shhsp90 α and U87-MG-17AAG) (Table 1). The Akt/PKB kinase activity was significantly reduced upon inhibition of Hsp90 by 17AAG and shRNA oligonucleotide targeted against hsp90 α . Statistical analysis using paired-sample *t*-test demonstrated ***P* < 0.001 for treated cells compared to untreated cells in U87-MG cell line showed a significant decrease in Akt kinase activity after hsp90 α inhibition using 17AAG and shhsp90 α . Note that shhsp90 α was less effective compared to 17AAG.

The U87-MG glioma cells were treated with 17AAG and transfected with shRNA targeting hsp90 α to inhibit Hsp90 function. Proteins were isolated from the samples, and proteomic analysis was carried out. A comprehensive proteomic study was further performed on the control (wild-type U87-MG) and the treated (U87-MG-17AAG and U87-MG-shhsp90 α) cell samples using 2D-DIGE and MALDI-TOF mass spectrometry. Proteomic analysis revealed 96 spots to be differentially expressed by a volume ratio of 1.5-fold or greater after treatment across the three samples tested (Figure 2). The spots were selected upon DeCyder analysis, and some of the spots were differentially modified between the three treated groups to show a slight shift in molecular weights. Mass spectrophotometric analysis was carried out using MALDI-TOF and for protein identification, and 36 spots showing $\geq$ 2-fold change were selected (Table 2). The protein identification was carried out on the basis of peptide fingerprint mass mapping (MS data) and peptide fragmentation mapping (tandem mass spectrometry (MS/MS) data). For the identification of proteins from their primary sequence databases, MASCOT search engine was used. From the 36 spots identified, 33 were identified as human proteins, while the other 3 were not elucidated (identified). More than one spot was identified as the same protein upon protein identification. This could have been attributed to the presence of different isoforms of the same protein following posttranslational modifications such as phosphorylation or methylation which changes the proteins isoelectric point (pI) and/or molecular weight (MW) causing the spots to shift along with protein fragmentation.

The U87-MG glioma cells were treated with IC₅₀ values of 17AAG (0.25 μ M) and KNK437 (55 nM) and then with a combination of IC₅₀ values of 17AAG and KNK437 (Figure 3). CellTiter-Glo one solution assay (Promega, UK) was used (as described previously) to determine the number of viable

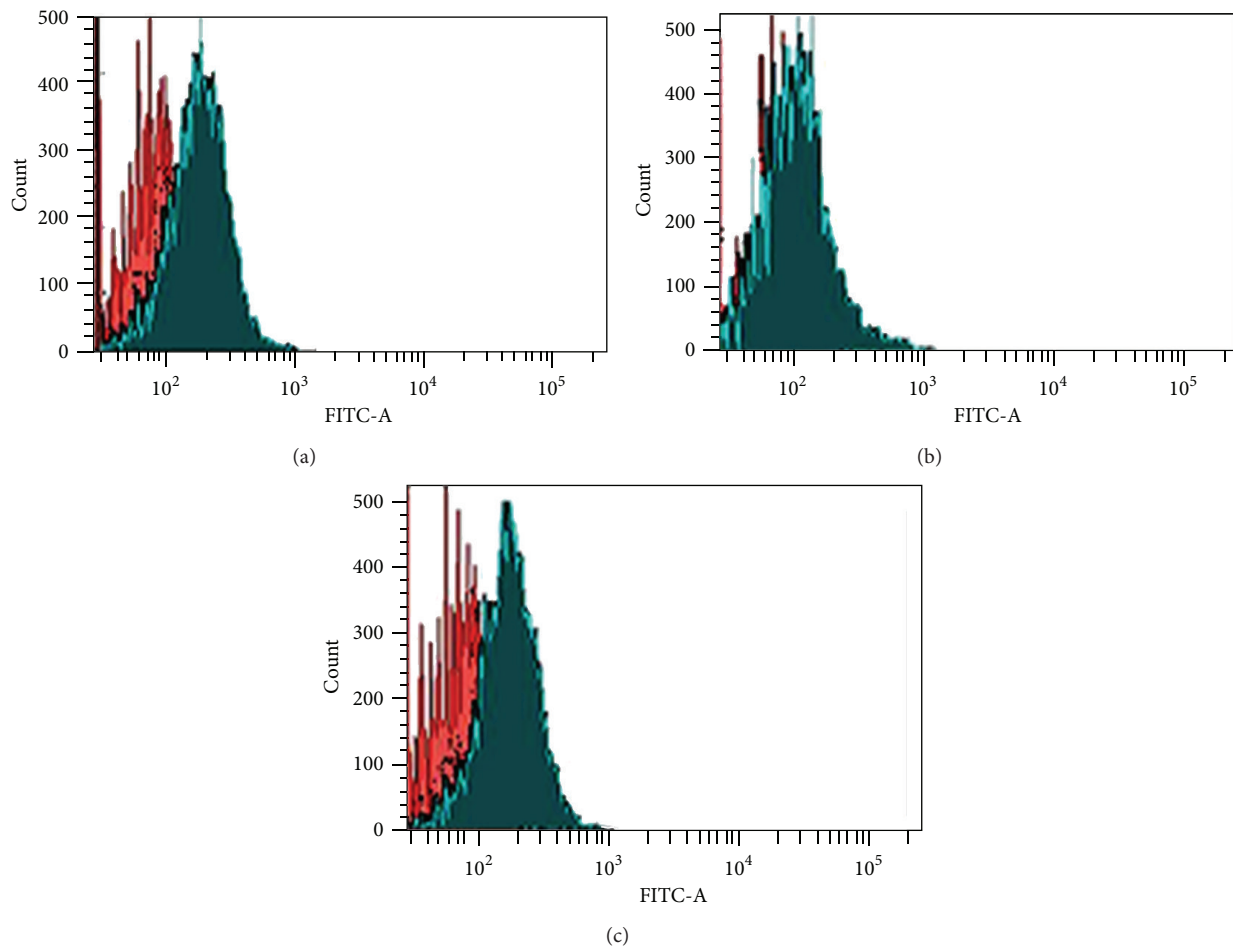


FIGURE 1: Flow cytometric analysis of control and treated U87-MG cell line for Hsp90 α detection. Hsp90 α antigen is detected with FITC that conjugated secondary antibody upon flow cytometric analysis; (a) control (wild-type U87-MG cells), (b) U87-MG cells treated with 17AAG, and (c) U87-MG cells treated with shRNA targeted against hsp90 α . Overlay image of negative (red) with the positive (blue). These flow cytometric readings are typical of 3 such different experiments.

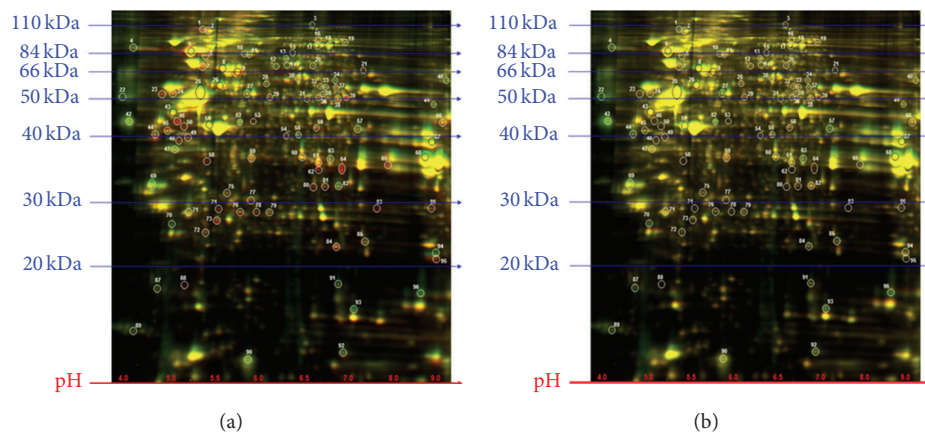


FIGURE 2: 2D-DIGE protein profile. Representation of the 2D-DIGE gel showing (a) control U87-MG and U87-MG-17AAG and (b) control U87-MG and U87-MG-shhsp90 α . Spots were picked automatically by spots volume ratio ≥ 2 .

TABLE 2: Proteins identified by mass spectrophotometry.

Protein	Accession number	Molecular function	Biological process	U87-MG-17AAG/U87-MG	U87-MG-sh <i>hsp90α</i> /U87-MG
Proteins upregulated by inhibition of Hsp90 by 17AAG and shRNA targeted towards <i>hsp90α</i>					
HSP70-1 (homosapiens)	gi 4529893	Chaperone activity	Protein metabolism	7.8	1.3
Hexokinase 1, isoform CRA_d (homosapiens)	gi 119574708	Catalytic activity	Metabolism, energy pathways	4.5	1.9
Tumor rejection antigen (gp96) 1 variant (homosapiens)	gi 62088648	Heat shock protein activity	Protein metabolism	3.1	1.7
Vimentin variant 3 (homosapiens)	gi 167887751	Structural constituent of cytoskeleton	Cell growth and/or maintenance	2	1.2
Pyruvate kinase, muscle (homosapiens)	gi 127795697	Kinase activity	Metabolism, energy pathways	6.6	2.9
Vimentin variant 3 (homosapiens)	gi 167887751	Structural constituent of cytoskeleton	Cell growth and/or maintenance	3.9	1.4
Heat shock 70 kDa protein 8 isoform 2 (homosapiens)	gi 24234686	Heat shock protein activity	Protein metabolism	4.2	1.2
Chain A, Crystal Structure of Aldose Reductase complexed with Dichlorophenylacetic acid	gi 119390284	Oxidoreductase activity	Metabolism, energy pathway	2.8	1.2
Annexin I (homosapiens)	gi 4502101	Calcium ion binding	Cell communication, signal transduction	16.8	1.7
Glyceraldehyde-3-phosphate dehydrogenase (homosapiens)	gi 31645	Catalytic activity	Metabolism, energy pathway	8	1.9
Ubiquitin carboxyl-terminal Esterase L1 (ubiquitin thiolesterase), Isoform CRA_c (homosapiens)	gi 119613387	Ubiquitin specific protease activity	Protein metabolism	2.2	1
Heat shock protein 27 (homosapiens)	gi 662841	Chaperone activity	Protein metabolism	2.7	1.1
Heat shock protein 27 (homosapiens)	gi 662841	Chaperone activity	Protein metabolism	2.7	1.2
Actin related protein 2/3 complex Subunit 2 (homosapiens)	gi 5031599	Cytoskeletal protein binding	Cytoskeleton organization and biogenesis	2.9	1
Heat shock 70 kDa protein 8 isoform 2 variants (homosapiens)	gi 62896815	Heat shock protein activity	Protein metabolism	4.5	1.3
Proteins downregulated by inhibition of Hsp90 by 17AAG and shRNA targeted towards <i>hsp90α</i>					
X-ray repair complementing defective repair in Chinese hamster cells 6 (Ku autoantigen, 70 kDa) (homosapiens)	gi 169145198	DNA binding	Regulation of nucleobase, nucleoside, nucleotide, and nucleic acid metabolism, DNA repair.	−3.7	−1.2
Vimentin (homosapiens)	gi 62414289	Structural constituent of cytoskeleton	Cell growth and/or maintenance	−2.3	−3.4
Vimentin (homosapiens)	gi 62414289	Structural constituent of cytoskeleton	Cell growth and/or maintenance	−2.1	−1.3
SERPINE1 mRNA binding protein 1 isoform 1	gi 66346679	RNA binding	Regulation of nucleobase, nucleoside, nucleotide, and nucleic acid metabolism.	−2.2	−1.4

TABLE 2: Continued.

Protein	Accession number	Molecular function	Biological process	U87-MG-17AAG/U87-MG	U87-MG-sh <i>hsp90α</i> /U87-MG
Calumenin isoform a precursor (homosapiens)	gi 4502551	Calcium ion binding	Cell communication, Signal transduction	-1.9	-2.5
Phosphoglycerate kinase 1 (homosapiens)	gi 4505763	Catalytic activity	Metabolism, Energy pathways	-1.2	-2
Aldolase A, fructose-bisphosphate, isoform CRA.b	gi 119600342	Lyase activity	Metabolism, Energy pathways	-2	-1.6
Predicted Glyceraldehyde-3-phosphate Dehydrogenase-like 6 (homosapiens)	gi 169208088	Catalytic activity	Metabolism, Energy pathways	-2.5	-1.8
Tropomyosin 4-anaplastic lymphoma kinase fusion protein (homosapiens)	gi 13274400	Structural constituent of cytoskeleton	Cell communication, Signal transduction	-2	-1.4
Eukaryotic translation initiation factor 3, subunit 12	gi 10801345	Translation regulator activity	Protein metabolism	-2.7	-1.6
Transgelin 2 (homosapiens)	gi 4507357	Unknown	Unknown	-3	-1
Chain A, Cyclophilin B complexed with (d-(Cholinylester)ser8)-Cyclosporin	gi 1310882	Unknown	Unknown	-3.7	-1.8
Proteins differentially expressed by inhibition of Hsp90 by 17AAG and shRNA targeted towards <i>hsp90α</i>					
Collagen, type VI, alpha 1 precursor (homosapiens)	gi 87196339	Extracellular matrix structural constituent	Cell growth and/or maintenance	2.1	-1.5
Chain A, structure of human Annexin A2 in the presence Of calcium ions	gi 56967118	Calcium ion binding	Signal transduction, Cell communication	2.1	-1.1
Heat shock protein beta-1 (homosapiens)	gi 4504517	Chaperone activity	Protein metabolism	2.3	-1.2
Chain A, Human manganese superoxide dismutase mutant Q143n	gi 2780818	Superoxide dismutase activity	Cell proliferation, Anti-apoptosis, Cell growth and/or maintenance	2.7	-1.2
Chain A, crystal structure of Ca2+-bound form of Des3-23alg-2	gi 211939086	Unknown	Apoptosis	2.3	-1.1
Transgelin 2 (homosapiens)	gi 4507357	Unknown	Unknown	2.4	-1.1

Note: The lists of identified peptides having ≥ 2 -fold change were searched against MASCOT database for the corresponding proteins. Proteins were categorized according to their change in expression after treatment with 17AAG and shRNA targeting *hsp90α* in U87-MG glioma cell line. Protein name, accession number, protein molecular function, biological function, and fold change (volume ratio treated/control) as generated by MASCOT and human protein research database are summarized.

cells present. The results show that combining 17AAG and KNK437 resulted in an increased cell death rate of U87-MG glioma cells (** $P < 0.001$).

4. Discussion

Hsp90 is upregulated in several human cancers, and targeting its function could be of therapeutic importance [2]. Previous studies have shown the presence of Hsp90 activity in glioma cell lines and tissues but not in normal brain cell lines or tissues [1, 19]. Hsp90 inhibitors exhibit antitumour activity by binding to Hsp90 and inducing proteosomal degradation of Hsp90 [20, 21]. As for 17AAG, a benzoquinone antibiotic derived from GA is a potent Hsp90 inhibitor, it dissociates the Hsp90 complex and leads to the inactivation of Hsp90 and

eventually results in its degradation [22, 23]. Furthermore, it has been reported to inhibit tumour growth in cell lines and it is being examined in preclinical trials [22, 24, 25].

Cohorts of U87-MG cells were treated with 17AAG and shRNA to target *hsp90α* in this study. The Akt/PKB kinase activity was significantly reduced by 81 and 59.4%, (** $P < 0.001$), and the flow cytometric analysis showed a reduction in Hsp90 protein levels by approximately 44 and 16% after 17AAG and sh*hsp90α* treatments, respectively (** $P < 0.001$). The reduction of Akt/PKB kinase protein was in agreement with previous studies which reported a dose dependent decrease in Hsp90 client proteins, namely, Akt after exposure, to 17AAG in murine neural stem cells and glioma stem cells [26]. Based on the previous literature reports and the laboratory findings, the inhibition of Hsp90 protein is a

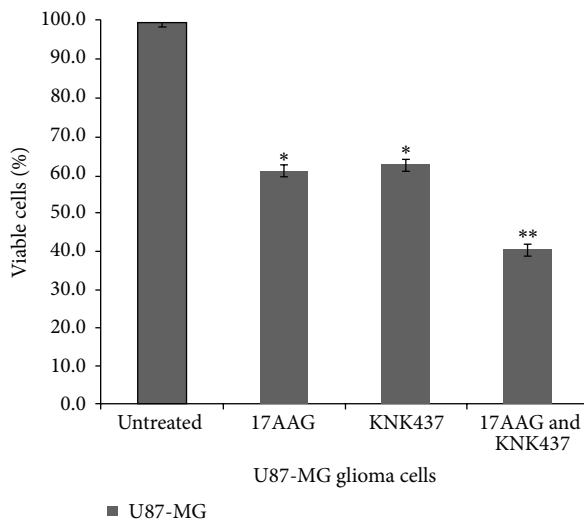


FIGURE 3: Cell viability assessment of U87-MG with 17AAG and KNK437. The percentage of viable U87-MG cells after treatment with inhibitors of Hsp90 and Hsp70, 17AAG, and KNK437, respectively, and a combination of the two. Data values are mean $\pm$ standard deviation, $n = 3$; * $P < 0.05$ and ** $P < 0.001$ are considered to be statistically significant.

more effective therapeutic approach than silencing *hsp90* gene since 17AAG showed a better silencing profile.

To understand the mechanisms involved or to characterize the changes caused at the cellular protein levels by inhibition of Hsp90, a differential proteomic analysis comparing control cells (wild-type U87-MG) and treated cells (U87-MG-sh*hsp90α* and U87-MG-17AAG) was performed. The 2D-DIGE technique was carried out to separate the proteins, while MALDI-TOF mass spectrometry was used for protein identification. Based on a 2-fold cut off, the analysis identified 36 proteins, while MALDI-TOF analysis identified 33 proteins with altered expressions showing $> 99\%$ confidence levels (3 protein being listed as unknown proteins).

On Hsp90 inhibition 15 proteins were found to be upregulated, of which 6 proteins are members of Hsp70 family together with Hsp27 and the gp96 which is a member of the Hsp90 family. These molecular chaperones serve as cochaperones to Hsp90 function [27, 28], thereby suggesting the role of Hsp90 cochaperones in compensating for Hsp90 function after Hsp90 inhibition. However, further research needs to be carried out to confirm these postulations. Members of the glycolysis pathways were also upregulated, demonstrating increased dependency on glycolysis for energy by the treated glioma cells. This phenomenon is described as the “Warburgs effect”, and it is an important phenomenon during malignant transformation [29]. The other proteins reported to be upregulated were vimentin, aldose reductase complexed with dichlorophenoxyacetic acid, annexin 1, ubiquitin thiolesterase isoform CRA.d, and actin related protein 2/3 complex subunit 2. Most of the proteins upregulated are involved in protein metabolism, energy pathways, cell growth and/or maintenance, metabolism, cell communication, signal

transduction, and cytoskeleton organization and biogenesis. Their upregulation could be attributed to their functions which may assist U87-MG glioma cells to survive after Hsp90 inhibition. Thus, Hsp90 inhibition may be sublethal, and there is a need for a multitarget approach to glioma therapy.

As discussed earlier, it was observed that 17AAG inhibits Hsp90 greater than shRNA targeting *hsp90α*. A significant difference between the increase in some of the key proteins, namely, Hsp70-1, Hsc70, hexokinase 1, pyruvate kinase, GAPDH, and annexin 1 was observed between the two treatments, that is, 17AAG and shRNA targeting *hsp90α*.

Several proteins such as Ku autoantigen, vimentin, serpinel mRNA binding protein 1, calumenin, phosphoglycerate kinase, aldolase A, tropomyosin 4-anaplastic lymphoma kinase fusion protein, eukaryotic translation initiation factor 3, transgelin 2, cyclophilin B complexed with (d-(cholinester)ser8)-cyclosporin, and GAPDH (predicted) were downregulated. Some of the proteins downregulated after Hsp90 inhibition are involved in various cellular responses such as DNA repair, protein synthesis, and cell mobility. Most of the proteins downregulated such as Ku70, SERPINE1, PGK1, and transgelin after Hsp90 inhibition are normally upregulated in several tumours including gliomas [30–33]. Thus, their subsequent downregulation after Hsp90 inhibition could be of therapeutic importance. However, further work should be carried out to understand the interactions between the downregulated proteins and Hsp90 in the treatment of gliomas. Though certain proteins like vimentin and GAPDH are present in both upregulated and downregulated lists of proteins, it could be argued that they are different isoforms/variants of the protein (as seen in Table 2) which were possibly altered by negative and/or positive feedback mechanism and, hence, show presence in both upregulated and the downregulated lists. Further studies should be carried out to further understand the mechanism.

Proteomic analysis in this study showed induction of the Hsp70 family members after Hsp90 inhibition. Therefore, it can thereby be postulated that inhibition of Hsp70 together with Hsp90 could be of therapeutic importance in glioma therapy. Based on this hypothesis, U87-MG glioma cells were treated with both a Hsp90 inhibitor, that is, 17AAG and a Hsp70 inhibitor, that is, N-formyl-3,4-methylenedioxybenzylidene-gamma-butyrolactam (KNK437). KNK437 is a benzylidene lactam compound which affects the induction of Hsp70 [34]. In addition to Hsp70, KNK437 has a considerable effect on the activation of the transcription factor, heat shock factor protein 1 (HSF1) and potentially inhibits the activity of Hsp40 and Hsp105, however, little is known about KNK437 and its mechanism [34]. This study in the light of previous finding examined the combined effect of inhibiting Hsp90 and Hsp70 with 17AAG and KNK437, respectively, in U87-MG glioma cell line.

Members of Hsp70 family act at multiple points in the apoptotic pathway and inhibit cell death [15–17]. Thus, even though Hsp90 was inhibited, the induction of Hsp70 after inhibition could have resulted in survival of the glioma cells (U87-MG). It can therefore be postulated that the inhibition of Hsp70 and Hsp90 would be of a therapeutic importance in glioma therapy. Based on this hypothesis, U87-MG glioma

cells were treated with both the Hsp90 inhibitor, that is, 17AAG, and the Hsp70 inhibitor, that is, KNK437. The IC₅₀ level of KNK437 was reported to be 55 nM, after 48 hours in the U87-MG cell line. Cell viability analysis demonstrated a higher percentage of U87-MG glioma cell death (~60%) (***P* < 0.001) when treated with a combination of 17AAG and KNK437 compared to treatment with 17AAG alone which resulted in ~39% cell death. When U87-MG glioma cells were treated with only KNK437, there was ~37% cell death. However, the effects of 17AAG and KNK437 are not synergetic but rather additive in nature. Thus, it could be suggested that combination therapy involving inhibition of both Hsp90 and Hsp70 could be used for glioma therapy.

5. Conclusion

In conclusion, the pilot study showed that the treatment of U87-MG glioma cells with 17AAG and shRNA to target *hsp90α*, effectively reduced Hsp90α activity and subsequently reduced the Akt/PKB kinase activity in U87-MG cell line. These results suggest that inhibition of Hsp90 protein activity could be applied in the future glioblastoma multiforme (GBM) therapy.

The efficacy of Hsp90 inhibitors can be compromised by the induction of antiapoptotic Hsp70 isoforms as an off-target effect of Hsp90 inhibition. Subsequently, Hsp90 and Hsp70 were both inhibited by treating U87-MG cells with 17AAG and KNK437, respectively. An induced U87-MG cell death rate was observed when both Hsp90 and Hsp70 were simultaneously inhibited. These findings suggest the applying of a combination therapy in the management of GBM. We propose a further analysis of the effects of inhibiting Hsp90 and Hsp70 in combination for glioma studies. It is equally important to determine the cellular and subcellular mechanisms, whereby KNK437 and 17AAG can induce cell death. Although proteomic screening identifies useful downstream effects of Hsp90 inhibition, the targets need further validation. The changes in protein expression and IPA analysis are indicative of alterations in multiple cell signalling pathways which need to be investigated.

Abbreviations

17AAG:	17-Allylamino-17-demethoxygeldanamycin
2D-DIGE:	Two-dimensional fluorescence difference gel electrophoresis
FACS:	Fluorescence-activated cell sorting
Hsp:	Heat shock protein
IC ₅₀ :	Half-maximal inhibitory concentration
KNK437:	N-formyl-3,4-methylenedioxy-benzylidene-gamma-butyrolactam
MALDI-TOF:	Matrix-assisted laser desorption/ionization time-of-flight
MS/MS:	Tandem mass spectrometry
shRNA:	Short hairpin RNA
SDS-PAGE:	Sodium dodecyl sulfate polyacrylamide gel electrophoresis.

Conflict of Interests

The authors would like to indicate that there is no conflict of interests.

Acknowledgment

This work was supported by a grant from the Sydney Driscoll Neuroscience Foundation (SDNF).

References

- [1] A. Shervington, N. Cruickshanks, R. Lea, G. Roberts, T. Dawson, and L. Shervington, "Can the lack of HSP90α protein in brain normal tissue and cell lines, rationalise it as a possible therapeutic target for gliomas?" *Cancer Investigation*, vol. 26, no. 9, pp. 900–904, 2008.
- [2] D. C. Altieri, "Coupling apoptosis resistance to the cellular stress response: the IAP-Hsp90 connection in cancer," *Cell Cycle*, vol. 3, no. 3, pp. 255–256, 2004.
- [3] L. Neckers, "Heat shock protein 90: the cancer chaperone," *Journal of Biosciences*, vol. 32, no. 3, pp. 517–530, 2007.
- [4] L. E. Cowen and S. Lindquist, "Hsp90 potentiates the rapid evolution of new traits: drug resistance in diverse fungi," *Science*, vol. 309, no. 5744, pp. 2185–2189, 2005.
- [5] B. K. Eustace, T. Sakurai, J. K. Stewart et al., "Functional proteomic screens reveal an essential extracellular role for hsp90α in cancer cell invasiveness," *Nature Cell Biology*, vol. 6, no. 6, pp. 507–514, 2004.
- [6] A. Rodina, M. Vilenchik, K. Moulick et al., "Selective compounds define Hsp90 as a major inhibitor of apoptosis in small-cell lung cancer," *Nature Chemical Biology*, vol. 3, no. 8, pp. 498–507, 2007.
- [7] N. Cruickshanks, L. Shervington, R. Patel, C. Munje, D. Thakkar, and A. Shervington, "Can hsp90α-targeted siRNA combined with TMZ be a future therapy for glioma?" *Cancer Investigation*, vol. 28, no. 6, pp. 608–614, 2010.
- [8] R. Oehler, B. Schmierer, M. Zellner, R. Prohaska, and E. Roth, "Endothelial cells downregulate expression of the 70 kDa heat shock protein during hypoxia," *Biochemical and Biophysical Research Communications*, vol. 274, no. 2, pp. 542–547, 2000.
- [9] D. Lanneau, A. de Thonel, S. Maurel, C. Didelot, and C. Garrido, "Apoptosis versus cell differentiation: role of heat shock proteins HSP90, HSP70 and HSP27," *Prion*, vol. 1, no. 1, pp. 53–60, 2007.
- [10] N. A. Seidberg, R. S. B. Clark, X. Zhang et al., "Alterations in inducible 72-kDa heat shock protein and the chaperone cofactor BAG-1 in human brain after head injury," *Journal of Neurochemistry*, vol. 84, no. 3, pp. 514–521, 2003.
- [11] S. K. Calderwood, J. R. Theriault, and J. Gong, "Message in a bottle: role of the 70-kDa heat shock protein family in anti-tumor immunity," *European Journal of Immunology*, vol. 35, no. 9, pp. 2518–2527, 2005.
- [12] E. Schmitt, M. Gehrmann, M. Brunet, G. Multhoff, and C. Garrido, "Intracellular and extracellular functions of heat shock proteins: repercussions in cancer therapy," *Journal of Leukocyte Biology*, vol. 81, no. 1, pp. 15–27, 2007.
- [13] J. Nylandsted, K. Brand, and M. Jäättelä, "Heat shock protein 70 is required for the survival of cancer cells," *Annals of the New York Academy of Sciences*, vol. 926, pp. 122–125, 2000.
- [14] S. K. Calderwood, M. A. Khaleque, D. B. Sawyer, and D. R. Ciocca, "Heat shock proteins in cancer: chaperones of

- tumorigenesis," *Trends in Biochemical Sciences*, vol. 31, no. 3, pp. 164–172, 2006.
- [15] S. Frese, M. Schaper, J.-R. Kuster et al., "Cell death induced by down-regulation of heat shock protein 70 in lung cancer cell lines is p53-independent and does not require DNA cleavage," *Journal of Thoracic and Cardiovascular Surgery*, vol. 126, no. 3, pp. 748–754, 2003.
 - [16] M. Pocaly, V. Lagarde, G. Etienne et al., "Overexpression of the heat-shock protein 70 is associated to imatinib resistance in chronic myeloid leukemia," *Leukemia*, vol. 21, no. 1, pp. 93–101, 2007.
 - [17] J. Kaur, J. Kaur, and R. Ralhan, "Induction of apoptosis by abrogation of HSP70 expression in human oral cancer cells," *International Journal of Cancer*, vol. 85, no. 1, pp. 1–5, 2000.
 - [18] S. Sato, N. Fujita, and T. Tsuruo, "Modulation of Akt kinase activity by binding to Hsp90," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 97, no. 20, pp. 10832–10837, 2000.
 - [19] M. D. Siegelin, A. Habel, and T. Gaiser, "17-AAG sensitized malignant glioma cells to death-receptor mediated apoptosis," *Neurobiology of Disease*, vol. 33, no. 2, pp. 243–249, 2009.
 - [20] T. W. Schulte, W. G. An, and L. M. Neckers, "Geldanamycin-induced destabilization of Raf-1 involves the proteasome," *Biochemical and Biophysical Research Communications*, vol. 239, no. 3, pp. 655–659, 1997.
 - [21] W. G. An, T. W. Schulte, and L. M. Neckers, "The heat shock protein 90 antagonist geldanamycin alters chaperone association with p210(bcr-abl) and v-src proteins before their degradation by the proteasome," *Cell Growth and Differentiation*, vol. 11, no. 7, pp. 355–360, 2000.
 - [22] K. S. Bisht, C. M. Bradbury, D. Mattson et al., "Geldanamycin and 17-allylamino-17-demethoxygeldanamycin potentiate the *in vitro* & *in vivo* radiation response of cervical tumor cells via the heat shock protein 90-mediated intracellular signaling and cytotoxicity," *Cancer Research*, vol. 63, no. 24, pp. 8984–8995, 2003.
 - [23] M. K. Hadden, D. J. Lubbers, and B. S. J. Blagg, "Geldanamycin, radicicol, and chimeric inhibitors of the Hsp90 N-terminal ATP binding site," *Current Topics in Medicinal Chemistry*, vol. 6, no. 11, pp. 1173–1182, 2006.
 - [24] A. M. Burger, H.-H. Fiebig, S. F. Stinson, and E. A. Sausville, "17-(Allylamino)-17-demethoxygeldanamycin activity in human melanoma models," *Anti-Cancer Drugs*, vol. 15, no. 4, pp. 377–387, 2004.
 - [25] M. Braga-Basaria, E. Hardy, R. Gottfried, K. D. Burman, M. Saji, and M. D. Ringel, "17-Allylamino-17-demethoxygeldanamycin activity against thyroid cancer cell lines correlates with heat shock protein 90 levels," *Journal of Clinical Endocrinology and Metabolism*, vol. 89, no. 6, pp. 2982–2988, 2004.
 - [26] C. M.-E. Sauvageot, J. L. Weatherbee, S. Kesari et al., "Efficacy of the HSP90 inhibitor 17-AAG in human glioma cell lines and tumorigenic glioma stem cells," *Neuro-Oncology*, vol. 11, no. 2, pp. 109–121, 2009.
 - [27] K. D. Dittmar and W. B. Pratt, "Folding of the glucocorticoid receptor by the reconstituted hsp90-based chaperone machinery. The initial hsp90-p60·hsp70-dependent step is sufficient for creating the steroid binding conformation," *Journal of Biological Chemistry*, vol. 272, no. 20, pp. 13047–13054, 1997.
 - [28] Y. Morishima, P. J. M. Murphy, D.-P. Li, E. R. Sanchez, and W. B. Pratt, "Stepwise assembly of a glucocorticoid receptor·hsp90 heterocomplex resolves two sequential ATP-dependent events involving first hsp70 and then hsp90 in opening of the steroid binding pocket," *The Journal of Biological Chemistry*, vol. 275, no. 24, pp. 18054–18060, 2000.
 - [29] O. Warburg, "On the origin of cancer cells," *Science*, vol. 123, no. 3191, pp. 309–314, 1956.
 - [30] H.-C. Wang, C.-S. Liu, C.-F. Chiu et al., "Significant association of DNA repair gene Ku80 genotypes with breast cancer susceptibility in Taiwan," *Anticancer Research*, vol. 29, no. 12, pp. 5251–5254, 2009.
 - [31] S. Martin, E. C. Cosset, J. Terrand, A. Maglott, K. Takeda, and M. Dontenwill, "Caveolin-1 regulates glioblastoma aggressiveness through the control of $\alpha 5 \beta 1$ integrin expression and modulates glioblastoma responsiveness to SJ749, an $\alpha 5 \beta 1$ integrin antagonist," *Biochimica et Biophysica Acta*, vol. 1793, no. 2, pp. 354–367, 2009.
 - [32] S. Kreth, J. Heyn, S. Grau, H. A. Kretzschmar, R. Egensperger, and F. W. Kreth, "Identification of valid endogenous control genes for determining gene expression in human glioma," *Neuro-Oncology*, vol. 12, no. 6, pp. 570–579, 2010.
 - [33] M. Shirahata, K. Iwao-Koizumi, S. Saito et al., "Gene expression-based molecular diagnostic system for malignant gliomas is superior to histological diagnosis," *Clinical Cancer Research*, vol. 13, no. 24, pp. 7341–7356, 2007.
 - [34] S.-I. Yokota, M. Kitahara, and K. Nagata, "Benzylidene lactam compound, KNK437, a novel inhibitor of acquisition of thermotolerance and heat shock protein induction in human colon carcinoma cells," *Cancer Research*, vol. 60, no. 11, pp. 2942–2948, 2000.

