University of Central Lancashire UCLan

Central Lancashire Online Knowledge (CLoK)

Title	Component-resolved diagnostics in the clinical and laboratory investigation of allergy
Type	Article
URL	https://clok.uclan.ac.uk/id/eprint/30341/
DOI	https://doi.org/10.1177/0004563219877434
Date	2020
Citation	Callery, Emma L, Keymer, Catherine, Barnes, Nicholas A and Rowbottom, Anthony (2020) Component-resolved diagnostics in the clinical and laboratory investigation of allergy. Annals of Clinical Biochemistry: International Journal of Laboratory Medicine, 57 (1). pp. 26-35. ISSN 0004-5632
Creators	Callery, Emma L, Keymer, Catherine, Barnes, Nicholas A and Rowbottom, Anthony

It is advisable to refer to the publisher's version if you intend to cite from the work. https://doi.org/10.1177/0004563219877434

For information about Research at UCLan please go to http://www.uclan.ac.uk/research/

All outputs in CLoK are protected by Intellectual Property Rights law, including Copyright law. Copyright, IPR and Moral Rights for the works on this site are retained by the individual authors and/or other copyright owners. Terms and conditions for use of this material are defined in the http://clok.uclan.ac.uk/policies/

Check for updates

DOI: 10.1177/0004563219877434

Annals of Clinical Biochemistry

Component-Resolved Diagnostics in the Clinical and Laboratory Investigation of Allergy

Journal:	*Annals of Clinical Biochemistry*
Manuscript ID	ACB-19-195.R1
Manuscript Type:	Review Article
Date Submitted by the Author:	27-Aug-2019
Complete List of Authors:	Callery, Emma; Lancashire Teaching Hospitals NHS Foundation Trust, Department of Immunology Keymer, Catherine; Royal Liverpool University Hospital, Department of Immunology Barnes, Nicholas; Faculty of Biology Medicine and Health Rowbottom, Anthony; Lancashire Teaching Hospital, Immunology
Keywords:	Proteins < Analytes

SCHOLARONE™
Manuscripts

DOI: 10.1177/0004563219877434

Clinical Sciences Review Committee (CSRC)

Commissioned Review

CSRC Article Number	
Review Title	Component-Resolved Diagnostics in the Clinical and Laboratory Investigation of Allergy
Running Title	CRD in Clinical and Laboratory Investigations of Allergy
Author(s)	Emma L. Callery*[a] , Catherine Keymer [b] , Nicholas A. Barnes [c] & Anthony W. Rowbottom*[ad]
Author Affiliations Including email address for each author	[a] Department of Immunology, Lancashire Teaching Hospitals NHS Foundation Trust, Preston PR2 9HT, UK [b] Department of Immunology, Royal Liverpool University Hospital, Liverpool, L69 3GA, UK [c] Manchester Academy for Healthcare Scientist Education, Faculty of Biology Medicine and Health, The University of Manchester, Manchester, M13 9PL [d] School of Medicine, University of Central Lancashire, Preston PR1 2HE, UK * Corresponding authors emma.callery@lthtr.nhs.uk anthony.rowbottom@lthtr.nhs.uk
Word Count	4997 (excluding abstract and references)

Declaration of Interests	
Funding	The authors received no financial support for the research, authorship, and/or publication of this review article.
Ethical Approval	N/A
Guarantor	Anthony W. Rowbottom

DOI: 10.1177/0004563219877434

Contributorship	Shared intellectual contributions, discussions, writing and revising of the manuscript
Acknowledgements	This article was prepared at the invitation of the Clinical Sciences Reviews Committee of the Association for Clinical Biochemistry and Laboratory Medicine
Key Words	Allergy, allergen, component resolved diagnostics, CRD, sensitisation, specific IgE, laboratory testing

DOI: 10.1177/0004563219877434

Abstract

The diagnosis and management of allergy is complex; the clinical symptoms associated with allergic reactions span a broad spectrum of severity, from mild hay fever-type symptoms though to life-threatening anaphylaxis. Obtaining an allergy-focused clinical history is therefore vital for identifying possible allergic triggers and directing testing. However this focus could be changing as scientific and technological advances have paved the way for developments within in vitro testing for allergy. With knowledge of allergens at the molecular level expanding, there are now the facilities to characterise the sensitisation profiles of allergy sufferers and determine the specific molecules (or components) against which the allergen-inducing immunoglobulin type E (IgE) proteins have been produced. This technology is termed component-resolved diagnostics (CRD).

We know that accurate identification of IgE specificity, the source of the causative allergen and knowledge of potential allergic cross-reactivities are required for optimal clinical management of allergy patients. These factors can make allergy a diagnostic challenge outside of a specialist centre, and contribute to the difficulties associated with requesting and interpreting allergy tests. The incorporation of CRD into current practice has provided a platform for patient-tailored risk stratification and improved the application of allergen-specific immunotherapy, revolutionising specialist management of these patients. This review discusses the roles of each type of testing in allergy management, and predictions for future pathways.

Background

Allergy is an increasingly prevalent phenomenon in the population. Allergy is typically associated with a small number of commonly encountered food types, pollens, drugs, and other sources. The molecules within these substances that illicit allergic reactions are known as allergens. (Laboratory approaches to assist in the diagnosis of allergic disease revolves around the detection of specific immunoglobulin E (sIgE), directed against likely causative sources of allergens based on an allergy-focussed patient history. The allergens in these tests are mixed in nature, as an extract from an allergen source will contain many different molecules. . Advances in laboratory techniques have led to the development of tests that detect sIgE directed against molecularly-defined allergenic components. The overarching term for this concept is component resolved diagnostics (CRD). The ability to distinguish between sIgE against unique components allows clinicians to identify true allergen sensitisation (production of sIgE towards a primary allergenic stimulus) from cross-reactivity (whereby sIgE against one allergen can bind to other allergens of similar molecular structure) with greater certainty.

DOI: 10.1177/0004563219877434

Following a recent survey into the use of CRD within allergy testing across the UK and Europe (1), it was apparent that use of CRD was becoming increasingly incorporated into routine diagnostics; however its clinical utility was potentially being compromised due to limited understanding and interpretive guidance for requesting clinicians. With wider implementation, the immunology laboratory must consider the practical aspects for CRD, both methodological and interpretative, prior to the introduction of these specialised tests. Given the need for an allergy-focussed patient history and the fact that CRD testing provides a wealth of complex allergomic data, we envisage that molecular allergology will remain in the specialist setting at the current time. However, in the future, allergy testing could be transformed with the assistance of artificial intelligence and machine learning algorithms that guide and direct primary care physicians through the complex interpretation of CRD and allergy diagnostics.

The aim of this article is to provide an overview of diagnostic allergy testing and to review the relevance of some of the recently introduced tests that require a clinical interpretation. It is outside the scope of this review to provide an in-depth evaluation on the applications of CRD. Instead this review will provide an overview of the indications and advantages of using CRD in allergy diagnosis and management.

The burden of allergy for healthcare

Allergic disease is one of the major causes of illness in developed countries and prevalence is increasing. The World Allergy Organisation (WAO) estimates that allergy prevalence of the whole population by country ranges between 10 - 40% (2). In Europe, an epidemiological review placed the prevalence of self-reported allergy at 17.3; thus nearly 1 in 5 people will experience some form of allergic reaction in their lifetime. In the UK, allergic disease affects about one in three of the population. In 13- to 14-year-old children, 32% report symptoms of asthma, 9% have eczema, and 40% have allergic rhinitis. The UK ranks highest in the world for asthma symptoms and is also near the top of the world ranking for allergic rhinitis and eczema. High and increasing trends are also apparent in nut allergy, anaphylaxis, occupational allergy (e.g. latex), and allergic reactions to drugs (3,4). In the 20 years prior to 2012 there was a 615% increase in the rate of hospital admissions for anaphylaxis in the UK (5).

IgE-driven allergic reactions

The vast majority of proteins we are exposed to daily are well tolerated. However, in some individuals, IgE immunoglobulins are produced in response to innocuous 'non-self' proteins encountered from various animal, plant and microbial sources. These misdirected immune responses result in allergic reactions.

Allergic reactions driven by IgE have three characteristic stages: sensitisation, elicitation and late phase. During sensitisation, an allergen is recognised by the immune system, causing production of sIgE. The sIgE then

DOI: 10.1177/0004563219877434

circulates in the blood to sites of connective tissue (such as the skin and lungs) where they bind to the complementary high affinity IgE receptors on mast cells. In the elicitation phase, further exposure to the same allergen results in cross-linking of sIgE molecules bound to the IgE receptors through binding of the allergen to two adjacent sIgE molecules. This induces the mast cell to release proinflammatory mediators including histamine, tryptase, prostaglandins, leukotrienes and complement anaphylatoxins that cause the clinical symptoms associated with an allergic reaction. In late phase responses, further release of biochemical mediators and recruitment of eosinophils and basophils plays a key role in amplification of the allergic reaction (6,7).

The severity of allergic reactions is dependent upon the nature of the allergen, the susceptibility of the patient and the route of exposure. Mild symptoms may be limited to hives, rhinitis, or minor swelling (usually affecting the lips, tongue and orbital region); whereas severe allergic reactions can induce systemic anaphylaxis. This variation provides a clinical dilemma in managing individual patients. Patients whose allergic triggers and clinical symptoms are consistent with the potential to develop more serious complications requiring assessment from specialist services need to be distinguished from those patients who can be confidently managed in primary care.

Exposure to allergic triggers

Allergens are most frequently encountered via ingestion, inhalation, or via a compromised skin barrier (Table 1). Food allergies elicit allergic reactions in the gastrointestinal (GI) tract upon consumption (8) (9). Airborne particles (e.g. pollen) cause allergic reactions in the respiratory system or upon contact with the eyes (10), whilst allergens that cross the skin barrier typically cause pruritic rashes and hives (11). Each of these exposure routes may progress to systemic reactions including anaphylaxis. The extent of allergic reaction is graded by severity of clinical manifestations (12).

The clinical approach to allergy diagnosis

In any patient with suspected clinical allergy, investigation is directed by a thorough allergy history which includes details of symptoms and time interval between exposure and onset of clinical symptom. Skin prick testing (SPT) is still a widely used diagnostic procedure and most commonly the first line investigative procedure used in allergy centres across Europe (13). These *in vivo* tests involve the cutaneous administration allergen extracts and the presence of a localised allergic response indicates a positive or negative result when compared with a control response. SPTs have been shown to have high sensitivity for identifying sensitisation

DOI: 10.1177/0004563219877434

in oral challenge-positive individuals, compared with *in vitro* specific allergen testing (14,15), and are reliable tests for respiratory allergy (16). However, allergen panels for skin prick testing can vary considerably, and studies are limited by the availability of defined allergens (17). The lack of standardisation in the manufacturing of allergen extracts for use in skin testing has also recently been highlighted and is currently being addressed in Europe (18,19). In addition, biological variation between patients affects the test performance. The sensitivity of skin testing is also affected by the threshold for positivity, and it is therefore vital that this is established locally based on equipment and technique or protocol used (20).

The laboratory approach to allergy diagnostics

Following an allergy-focussed patient history to identify potential triggers, and subsequent skin prick testing, the detection of specific IgE using *in vitro* methods, in particular allergen-specific IgE tests to allergen extracts are commonly used. The *in vitro* tests for specific IgE, as with *in vivo* SPT, use whole allergen extracts for analysis therefore are negatively affected by the heterogeneous nature of purified allergens. This problem is overcome through use of CRD, as the IgE being measured in this case is directed against separate allergen components, and therefore avoids any interference from minor or non-specific allergens. These issues could have a significant impact on the usefulness of tests in clinical practice and highlight the need for specialised allergy services alongside a standardised approach when assessing patients, particularly when selecting patients for immunotherapy in consideration of test results. Whilst this approach remains clinically relevant for the routine mono-sensitised allergic patients, an improved diagnostic approach remains a clinical need, particularly for more complex cases which mainly arise due to the complexities of protein components of allergens, and their respective allergenicity. Subsequent testing using molecular-based allergen components may follow later, in an approach termed the 'top-down' diagnostic approach (21).

The increased availability and use of CRD is not only beginning to have an impact on the investigation of these challenging patients but is also unravelling further scientific understanding of the immunological signalling pathways involved in IgE-mediated hypersensitivity. Given the high levels of clinically variability observed between allergy patients, and even between patients with the same apparent allergy, it would appear that multiple factors and signalling pathways are at play following allergen binding and cross-linking IgE on the surface of mast cells and basophils, which can influence the overall clinical outcome.

For example, the molecular data generated from undertaking CRD in patients with peanut allergy has enabled a level of risk stratification to be realised. This has improved diagnosis by introducing a more clear-cut decision points for peanut CRDs and has reduced the number of challenge tests undertaken in clinic (22). These changes are having significant consequences for the patient and, whilst the precise signalling pathways triggered following the binding of each peanut component to mast cell surface IgE have yet to be elucidated, it raises

DOI: 10.1177/0004563219877434

the question of whether a single common cell signalling pathway is activated for all allergens, or in fact multiple, cross-talking pathways are triggered depending on the inciting molecule(s).

As we obtain more molecular data for allergens and begin to scratch the surface as to the signalling pathways activated, we have become aware that particular molecules within allergen proteins can exhibit a hierarchy or allergenic dominance, allowing risk stratification to be determined for certain allergies (23,24). However this now raises further questions as to whether the IgE-driven immune response triggered by every allergen follows a common sequence of signalling events in order to elicit the well-recognised allergic symptoms, or whether a hierarchy of allergen epitope dominance exists within allergen-driven signalling, which contains multiple cross-talking signalling pathways, activation of which is dependent on the particular allergen molecule-receptor binding.

One way of deciphering this complex enigma is to utilise technologies including CRD, specifically ISAC allergen microarrays. ISAC microarray consists of 112 characterised allergens in a single test. This provides significant additional information which can be made available to clinicians, but the interpretation of which requires highly specialised expertise to be of clinical value. Most likely due to this limitation, it has not as yet been fully supported by current guidelines for management of allergy, but has been shown to have significant value in appropriately selected patients (25), in particular with respect to patterns of sensitisation, and idiopathic anaphylaxis.

Promoting the use of CRD and associated platforms such as ISAC in a controlled manner will potentially expand our knowledge of the allergic response pathways, and may uncover sensitisations that are as yet unknown. This should not be underestimated given the asymptotic nature of allergens that are yet to be characterised.

How can CRD help?

Allergen families & prediction of severity

Up to 90% of food allergies are caused by a group of animal and plant sources collectively known as the “Big 8”: cow’s milk, eggs, peanuts, tree nuts (e.g. almonds, hazelnuts), fish, shellfish, soy and wheat (26). The same allergen source can elicit different types of allergic response amongst individuals. For example, peanuts can cause fatal allergic reactions in some individuals, whilst others only experience local mild reactions (27).

This variation can be attributed to specific allergenic components and their biochemical properties. Each source contains a number of different proteins, or allergen components, which can cause an allergic reaction; any two individual patients could be allergic to the same or different allergenic proteins within a source. Knowledge of the allergenic components linked to these reactions can help resolve the complex nature of allergy.

DOI: 10.1177/0004563219877434

Components from different sources can be grouped into allergen families based on shared molecular characteristics (28). Molecular analysis of the ~700 known allergens has demonstrated that the majority of allergens can be distributed into few protein families and possess a limited number of biochemical functions (29). Determining the sensitising component, and its associated allergen family, can aid the clinician in determining the level of risk. This will ultimately improve the management of allergy patients; high-risk patients can be appropriately given and trained to use adrenaline injector pens, whilst reassurance can be given to lower risk patients, preventing unnecessary diet restrictions, anxiety, and referrals to tertiary care.

Cross-reactivity between allergens from different sources is explained by the homogeneity exhibited within these protein families (30). High levels of cross-reactivity with other plant food allergens can result in allergic reactions when consumed (typically raw and/or unprocessed), despite not being originally sensitised to that source (31).The most prevalent plant-sourced allergens are distributed into distinct families: pathogenesis-related (PR) proteins, profilins, prolamins such as non-specific lipid transfer proteins (nsLTP), and seed storage proteins. Figure 1 illustrates the major allergen components found in peanut (Ara h) and links them to their associated protein families; the individual components (and protein families) are depicted in order of risk severity.

Allergenicity (the potential to trigger an allergic response) is reflected in the molecular structure of an allergen, with relevant features such as size, solubility, stability, and conformation contributing to the ability of a particular molecule to trigger an allergic response (32). A primary allergen is the original molecule that induces sensitisation and production of sIgE. Cross-reactive allergens can cause allergic reactions to molecules present in different sources. Cross-reactivity occurs due to similarities between amino acid sequence homology and 3D protein-folding of allergen components; shared structural conformations allow cross-reactive allergens to interact with IgE previously generated against the primary allergen, with the potential to induce an allergic response (32). Reactions to cross-reactive molecules may be less severe than the reactions to primary genuine allergens (21).

Cross-reactivity also occurs due to post-translational glycosylation or cross-reactive carbohydrate determinants (CCDs) which exhibit similarity between different sources such as plants, venom proteins and house dust mites. The production of sIgE against CCDs can give a broad sensitisation profile, causing multiple positive responses by SPT or *in vitro* IgE measurements (33). These cases can be difficult to interpret for the requesting clinicians; however as CCD do not usually associate with clinical symptoms, the ability to identify sensitisation to CCD can help eliminate clinically irrelevant positive results from true major allergen sensitisation.

Allergen molecules

DOI: 10.1177/0004563219877434

Genuine or major allergenic molecules are classified as those causing species-specific sensitisation and subsequent IgE-mediated immune reactions to its corresponding allergen source for most allergic individuals. The definition of a major allergen is based on the prevalence of IgE or positive skin prick test to the total allergen extract however this does not take into consideration the allergenic risk for eliciting an immune response. Generally, in allergic patients major allergenic molecules bind to IgE and trigger responses in >50% of individuals with an allergy to its source, whereas minor allergens induce allergic reaction in <50% of allergic patients (21,34).

For example, sensitisation to Bet v 1, the major allergen of *Betula verrucos,* (birch pollen), is found in over 90% of birch pollen allergy patients in Europe, with minor panallergen molecules Bet v 2 and Bet v 4 found to be responsible for sensitisation of 44.6% and 9.4 % of birch allergy patients respectively (35–37). The term 'panallergen' refers to widely distributed ubiquitous protein molecules occurring across multiple species and allergen sources, and includes molecules such as profilins and calcium-binding proteins (31). The clinical relevance of panallergen sensitisation is considered limited, although can present problems if patients develop symptomatic multiple sensitives (38,39). The key aspect of all diagnostic allergy investigations, the allergy-directed clinical history, will ultimately guide interpretation of serological results; however CRD can have a significant influence on future management options.

Clinical example: Pollen-associated food allergy/Oral allergy syndrome (Bet v 1 homologues)

A well-known group of cross-reacting plant allergens are the Bet v 1 homologues, belonging to the PR-10 family of proteins. As illustrated in figure 2, cross-reactive Bet v 1 homologues are found in a number of allergens from pollen, fruits, nuts, vegetables and legumes. Exposure, usually of raw, unprocessed allergen-containing foods, can result in a localised, symptomatic IgE-mediated allergic response in patients identified to have an allergy to birch pollen, driven by the major allergen Bet v 1. Using CRD to define primary sensitisation to PR-10 allergen molecules can aid diagnosis of oral allergy syndrome. Furthermore, CRD can be helpful in identifying birch allergy patients with sensitisation to the minor allergen Bet v 2, thus potential cross-reactivity to components from the profilin family. As profilins are the most widely distributed proteins of the allergen families, cross-reactivity can occur from a wide range of unrelated species, including trees, weeds, grasses, fruits, vegetables and nuts, but will be distinct from those of the Bet v 1 homologues. Molecule-based approaches enable distinction between genuine sensitisation and the less clinically relevant IgE cross-reactivity due to panallergens or carbohydrate determinants. Polysensitivity within birch allergy patients can also be identified using CRD; for which the determination of the primary sensitising source can help to direct appropriate management, including suitability for AIT.

DOI: 10.1177/0004563219877434

An official list of allergens is published by the WHO-International Union of Immunological Societies (IUIS) allergen nomenclature sub-committee (http://www.allergen.org). This list currently contains approximately 880 proteins, drawn from a variety of sources, each proven to have evidence of causing allergic reactions and IgE-binding capacity from the sera of at least 5 patients. Allergen names are assigned by the allergen nomenclature sub-committee for consistent reproduction, compromising a reduced form of the taxonomic name (for example, peanut, *Arachis hypogea*, is abbreviated to Ara h), followed by a number to identify the protein family (40).

Diagnostic accuracy in polysensitisation and cross-reactivity

Allergy diagnosis is as complex as finding needle in a hay-stack. The patient may report several potential allergenic triggers and it's often difficult to pin-point the exact culprit without detailed allergy-focused clinical history and the additional laboratory diagnostics. Although sIgE testing is widely used in clinical practice especially within primary care settings, the obtained serological results can often further complicate matters, particularly if sensitisation to several different allergens is detected (co- or polysensitisation).

A major advantage of CRD includes assisting in the prediction of response by specifying 'genuine' as well as the 'cross-reactive' components in polysensitised individuals; such resolution is not possible using conventional testing. This has been demonstrated for several allergens including food, pollen, venom and idiopathic anaphylaxis (39). Use of CRD in these patients not only improves the diagnostic accuracy but also provides additional information around individuals' sensitisation profile which can assist risk stratification (illustrated in figure 1). Co-sensitisation is distinct from cross-reactivity, and occurs when IgE-driven responses are triggered by major allergenic molecules from two or more unrelated sources. The determination of genuine co-reactivity and cross-reactivity using CRD can have a significant impact on the interpretation of allergy results, particularly in complex cases whereby patients demonstrate presence of sIgE against multiple antigens. Major advantages of CRD include determination of the primary major allergen component (and protein family) to aid determination of reaction severity; the prediction of additional cross-reactivities and allergic reactions following exposure to sources containing molecular allergen homologues; and the identification of CCD-based cross-reactivity (which are unlikely to elicit reactions).

Furthermore, there will be consequences for the appropriate administration of allergy immunotherapy; as responses will vary depending on whether genuine or cross-reactive allergen sensitisation is determined.

Immunotherapy and scope for personalised treatment

Although exact immunological mechanism underlying the development of immune tolerance, and thus response to allergen-specific immunotherapy remains to be fully elucidated, it is accepted that the allergen component specificity to which the patient is sensitised plays a fundamental role (41). As highlighted earlier,

DOI: 10.1177/0004563219877434

a considerable challenge for diagnostic investigations is associated with the extracts used to represent the allergen source. For both serological (*in vitro*) or skin-prick (*in vivo*) testing, these are traditionally based on crude extracts of the allergen source, and therefore contain a mixture of relevant and less clinically relevant allergen components. Immunotherapy products face a similar problem, in addition to the fact that standardisation of allergen concentration is usually only reported for major allergen molecules. This is likely to explain the variation in treatment responses between patients as some allergen molecules to which a patient is sensitised to will be poorly represented in the therapeutic product (42). In light of this, it is vital to identify the allergen components to which patients are responding to, as this will massively assist in anticipating the efficacy of treatment (43).

For instance, not all patients will be suitable candidates for immunotherapy as they will not be sensitised to the major allergens found in commercial immunotherapy extracts, or, due to heterogeneity between therapeutic extracts, the major allergens they are susceptible to are poorly represented (42). Prediction of treatment response using CRD would prevent inappropriate use of expensive immunotherapy in patients shown to respond to minor allergens, levels of which are found at much lower concentrations in commercial AIT (often unstandardized) and therefore are unlikely to induce allergen tolerance. CRD could also be used to reduce severe adverse responses during treatment. Some of the minor allergen components can be found in high concentrations within commercial IAT and sudden exposure on commencement of treatment could potentially trigger life-threatening anaphylaxis (44). These situations could be avoided if patients found to be sensitised to these molecular components can be identified prior to starting treatment.

Further considerations for diagnostic allergy testing

In comparison with the heterogeneous nature of SPT and specific IgE tests, the products used in molecular component *in vitro* allergy tests are highly purified and the constituents of the component tests are fully characterised with no other minor antigens present. Single allergens are used (rather than multiple allergen extracts with varying compositions), and testing is carried out in laboratories where analytical performance is tightly regulated. However, due to the availability of allergens, the repertoire of these assays are restricted by the expansive existence of allergens which cannot feasibly be reflected in analytical tests using current methods (16,17).

Both SPT and *in vitro* diagnostic techniques are restricted in their use of allergenic source extracts. By their nature, they are a mix of proteins and other molecules from an animal, plant or microbial source. Thus detection of a positive response only identifies the source material, but not the specific protein to which the immune response has raised a specific IgE against. This lack of specificity causes difficultly in accurately

DOI: 10.1177/0004563219877434

identifying whether positive results are linked to reaction severity and predicting the likelihood of experiencing future allergic reactions to similar molecules in other allergenic sources (17,21).

An absence of international calibration standards for specific IgE assays, including allergen component testing, poses further analytical problems with inconsistencies, as a reference standard is only available for the measurement of total IgE (WHO 11/234). Instead, the units measured in sIgE assays are converted into quantitative sIgE antibody levels (kUA/l), using a reference curve calibrated to the WHO standard for total IgE. The assumptions made in the process (around the binding avidity and avidity of sIgE and allergen extract being comparable to the antibody binding when generating the total IgE reference curve) introduces one of many sources of error, resulting in potential quantitative inaccuracies up to 10% (45). These aforementioned factors have implications for laboratory testing and quantitation sIgE. Of particular importance, and relevant in order to achieve UKAS accreditation is the ability to define the measurement of uncertainty (MoU) within allergy diagnostics. Adherence to ISO 15189 laboratory standards requires that each laboratory must define their uncertainty budget for each analyte measured, meaning the inherent error of a result from analytical and biological differences. The lack of standardisation makes defining this measurement with confidence a challenge for laboratories.

Multiple studies have also compared the performance of CRD with SPTs and challenge tests (gold standard provocation testing) for diagnosis of allergy, but a consensus has yet to be reached as the analytical performance varies depending on the allergen (17). When considering this in the clinical pathway, it would seem reasonable that both testing methods are used, particularly whilst there is an absence of appropriate clinical trials providing evidence of efficacy of treatment by allergen-specific immunotherapy based on components (17,21).

The future

As with most diagnostic testing the immediate clinical context and past medical history are essential in order to interpret laboratory results. Often in the case of allergy, which is not monospecific, the presence of multiple reactivities to a range of allergens presents a diagnostic challenge. This is particularly evident when tests are requested by non-specialists such as general practitioners in the community. This lack of understanding often leads to wide variation in practice and inappropriate referrals to specialist allergy services and the greater use of emergency departments. The getting right first time (GIRFT) initiative (46) seeks to address these issues and improve the quality of care within the NHS. By reducing variation and promoting best practice it is predicated that there will be improved patients outcomes and opportunities for cost efficiencies. The challenge for allergy will be how best to support healthcare providers in the community given an ever increasing workload and limited time available for patient individual consultations.

DOI: 10.1177/0004563219877434

The NHS Five Year Forward plan (47,48) identifies the need for more GPs who are supported by other professionals. It is hoped that this be will accelerated by partnerships between services in the community and hospital setting. How this might be achieved for allergy diagnostics remains unclear given the shortage of allergy specialists. It is apparent that the current ways of delivering education and information are struggling to have a significant impact which will continue until alternatives ways are established. This situation is likely to be compounded with the wider use of CRD. It is therefore important that new ways of delivering allergy services across integrated pathways are developed. Following the publication of the Industrial Strategy (49) and, in particular, the Health Sector deal (50) it is clear one such area might be the use of machine learning and artificial intelligence. The creation of case-based algorithms might provide opportunities for clinical standardisation and reduction in the unwarranted variation of allergy diagnostics. The use of machine learning and big data approaches might also provide a framework to support education across all healthcare providers potentially operating to a single governance structure. The use of exception rules provided by new cases would help refine the clinical utility of such an approach especially as new knowledge is uncovered by the wider adoption of CRD testing. Empowering patients to be more responsible for their care is a key objective in the NHS Five year Forward plan (48) and the greater use of artificial intelligence might offer a means by which this can be achieved on a large scale.

What does the future hold for the allergy patient? Given the growth in services available over the internet and the influence of social media it is especially likely that patients will continue to present well informed and potentially with a series of results from unconventional sources. Providing the healthcare provider, in whatever setting, with clear succinct information at the point of consultation will be central for providers of allergy tests to ensure the appropriate test is performed for the appropriate patient at the appropriate time

Conclusion/Summary

At the current point in time the application and routine use of CRD remain in its infancy, in need of immunology specialists, laboratories and requesting clinicians to evaluate the wealth of new evidence around their use in the absence of standardised protocols. CRD will undoubtedly add value to understanding the IgE response on a molecular level in addition to expanding the current armoury of tests available for allergy investigation. The enhanced use of CRD by clinicians and allergy specialists is now required in order to obtain useful clinical and laboratory information in terms of sensitivity, specificity, PPV, NPV and risk assessment. This is evidenced in the wealth of recently published literature, however there is still a long way to go before CRD is being routinely used, confidently interpreted and adding value to individual patient cases on a regular basis.

DOI: 10.1177/0004563219877434

Funding

The authors received no financial support for the research, authorship, and/or publication of this article.

DOI: 10.1177/0004563219877434

References

1. Saleem R, Keymer C, Patel D, Egner W, Rowbottom AW. UK NEQAS survey of allergen component testing across the United Kingdom and other European countries. Clin Exp Immunol. 2017 Jun 1;188(3):387–93.

2. Pawankar R. Allergic diseases and asthma: a global public health concern and a call to action. World Allergy Organ J. 2014;7(1):12.

3. Royal College of Physicians. Allergy services: still not meeting the unmet need. London: RCP; 2010. (Report of the Joint Royal College of Physicians and Royal College of Pathologists Working Party).

4. Royal College of Physicians. Allergy: the unmet need: a blueprint for better patient care: a report of the Royal Colleges of Physicians Working Party on the Provision of Allergy Services in the UK. London: RCP; 2003.

5. Turner PJ, Gowland MH, Sharma V, Ierodiakonou D, Harper N, Garcez T, et al. Increase in anaphylaxis-related hospitalizations but no increase in fatalities: an analysis of United Kingdom national anaphylaxis data, 1992-2012. J Allergy Clin Immunol. 2015 Apr;135(4):956–963.e1.

6. Delves PJ. Hypersensitivity: IgE-Mediated (Type I). In: John Wiley & Sons Ltd, editor. eLS [Internet]. Chichester, UK: John Wiley & Sons, Ltd; 2017 [cited 2018 Nov 16]. p. 1–8. Available from: http://doi.wiley.com/10.1002/9780470015902.a0000965.pub3

7. Galli SJ, Tsai M. IgE and mast cells in allergic disease. Nat Med. 2012 May;18(5):693.

8. Matsuo H, Yokooji T, Taogoshi T. Common food allergens and their IgE-binding epitopes. Allergol Int Off J Jpn Soc Allergol. 2015 Oct;64(4):332–43.

9. Abrams EM, Sicherer SH. Diagnosis and management of food allergy. CMAJ Can Med Assoc J J Assoc Medicale Can. 2016 Oct 18;188(15):1087–93.

10. Brożek JL, Bousquet J, Agache I, Agarwal A, Bachert C, Bosnic-Anticevich S, et al. Allergic Rhinitis and its Impact on Asthma (ARIA) guidelines-2016 revision. J Allergy Clin Immunol. 2017 Oct;140(4):950–8.

11. Sicherer SH, Leung DYM. Advances in allergic skin disease, anaphylaxis, and hypersensitivity reactions to foods, drugs, and insects in 2014. J Allergy Clin Immunol. 2015 Feb;135(2):357–67.

12. Niggemann B, Beyer K. Time for a new grading system for allergic reactions? Allergy. 2016 Feb;71(2):135–6.

13. Cardona V, Demoly P, Dreborg S, Kalpaklioglu AF, Klimek L, Muraro A, et al. Current practice of allergy diagnosis and the potential impact of regulation in Europe. Allergy. 2018 Feb;73(2):323–7.

14. Alessandri C, Zennaro D, Scala E, Ferrara R, Bernardi ML, Santoro M, et al. Ovomucoid (Gal d 1) specific IgE detected by microarray system predict tolerability to boiled hen's egg and an increased risk to progress to multiple environmental allergen sensitisation. Clin Exp Allergy J Br Soc Allergy Clin Immunol. 2012 Mar;42(3):441–50.

15. Ott H, Baron JM, Heise R, Ocklenburg C, Stanzel S, Merk H-F, et al. Clinical usefulness of microarray-based IgE detection in children with suspected food allergy. Allergy. 2008 Nov;63(11):1521–8.

DOI: 10.1177/0004563219877434

16. Jensen-Jarolim E, Jensen AN, Canonica GW. Debates in allergy medicine: Molecular allergy diagnosis with ISAC will replace screenings by skin prick test in the future. World Allergy Organ J. 2017;10(1):33.

17. Larenas-Linnemann D, Luna-Pech JA, Mösges R. Debates in Allergy Medicine: Allergy skin testing cannot be replaced by molecular diagnosis in the near future. World Allergy Organ J [Internet]. 2017 Dec [cited 2017 Oct 9];10(1). Available from: http://waojournal.biomedcentral.com/articles/10.1186/s40413-017-0164-1

18. Chapman MD, Ferreira F, Villalba M, Cromwell O, Bryan D, Becker W-M, et al. The European Union CREATE project: a model for international standardization of allergy diagnostics and vaccines. J Allergy Clin Immunol. 2008 Nov;122(5):882–889.e2.

19. van Ree R, Chapman MD, Ferreira F, Vieths S, Bryan D, Cromwell O, et al. The CREATE project: development of certified reference materials for allergenic products and validation of methods for their quantification. Allergy. 2008 Mar;63(3):310–26.

20. Peters RL, Gurrin LC, Allen KJ. The predictive value of skin prick testing for challenge-proven food allergy: a systematic review. Pediatr Allergy Immunol Off Publ Eur Soc Pediatr Allergy Immunol. 2012 Jun;23(4):347–52.

21. Matricardi PM, Kleine-Tebbe J, Hoffmann HJ, Valenta R, Hilger C, Hofmaier S, et al. EAACI Molecular Allergology User's Guide. Pediatr Allergy Immunol. 2016 May 1;27:1–250.

22. Eller E, Bindslev-Jensen C. Clinical value of component-resolved diagnostics in peanut-allergic patients. Allergy. 2013 Feb;68(2):190–4.

23. Shreffler WG, Lencer DA, Bardina L, Sampson HA. IgE and IgG4 epitope mapping by microarray immunoassay reveals the diversity of immune response to the peanut allergen, Ara h 2. J Allergy Clin Immunol. 2005 Oct 1;116(4):893–9.

24. Thomas WR. Hierarchy and molecular properties of house dust mite allergens. Allergol Int. 2015 Oct;64(4):304–11.

25. Heaps A, Carter S, Selwood C, Moody M, Unsworth J, Deacock S, et al. The utility of the ISAC allergen array in the investigation of idiopathic anaphylaxis. Clin Exp Immunol. 2014 Aug;177(2):483–90.

26. Sicherer SH, Sampson HA. Food allergy: A review and update on epidemiology, pathogenesis, diagnosis, prevention, and management. J Allergy Clin Immunol. 2018 Jan;141(1):41–58.

27. Vereda A, van Hage M, Ahlstedt S, Ibañez MD, Cuesta-Herranz J, van Odijk J, et al. Peanut allergy: Clinical and immunologic differences among patients from 3 different geographic regions. J Allergy Clin Immunol. 2011 Mar;127(3):603–7.

28. Hoffmann-Sommergruber K, Mills ENC. Food allergen protein families and their structural characteristics and application in component-resolved diagnosis: new data from the EuroPrevall project. Anal Bioanal Chem. 2009 Sep;395(1):25–35.

29. Radauer C, Bublin M, Wagner S, Mari A, Breiteneder H. Allergens are distributed into few protein families and possess a restricted number of biochemical functions. J Allergy Clin Immunol. 2008 Apr;121(4):847–852.e7.

30. Popescu F-D. Cross-reactivity between aeroallergens and food allergens. World J Methodol. 2015 Jun 26;5(2):31–50.

DOI: 10.1177/0004563219877434

31. Hauser M, Roulias A, Ferreira F, Egger M. Panallergens and their impact on the allergic patient. Allergy Asthma Clin Immunol Off J Can Soc Allergy Clin Immunol. 2010 Jan 18;6(1):1.

32. Aalberse RC. Structural biology of allergens. J Allergy Clin Immunol. 2000 Aug 1;106(2):228–38.

33. Altmann F. Coping with cross-reactive carbohydrate determinants in allergy diagnosis. Allergo J Int. 2016;25(4):98–105.

34. Canonica GW, Ansotegui IJ, Pawankar R, Schmid-Grendelmeier P, van Hage M, Baena-Cagnani CE, et al. A WAO - ARIA - GA^{2}LEN consensus document on molecular-based allergy diagnostics. World Allergy Organ J. 2013 Oct 3;6(1):17.

35. Lizaso MT, García BE, Tabar AI, Lasa E, Echechipía S, Álvarez MJ, et al. Comparison of conventional and component-resolved diagnostics by two different methods (Advia-Centaur/Microarray-ISAC) in pollen allergy. Ann Allergy Asthma Immunol. 2011 Jul;107(1):35–41.

36. Movérare R, Westritschnig K, Svensson M, Hayek B, Bende M, Pauli G, et al. Different IgE reactivity profiles in birch pollen-sensitive patients from six European populations revealed by recombinant allergens: an imprint of local sensitization. Int Arch Allergy Immunol. 2002 Aug;128(4):325–35.

37. Rossi RE, Monasterolo G, Monasterolo S. Detection of specific IgE antibodies in the sera of patients allergic to birch pollen using recombinant allergens Bet v 1, Bet v 2, Bet v 4: evaluation of different IgE reactivity profiles. Allergy. 2003 Sep;58(9):929–32.

38. Canis M, Gröger M, Becker S, Klemens C, Kramer MF. Recombinant marker allergens in diagnosis of patients with allergic rhinoconjunctivitis to tree and grass pollens. Am J Rhinol Allergy. 2011 Feb;25(1):36–9.

39. Treudler R, Simon JC. Overview of Component Resolved Diagnostics. Curr Allergy Asthma Rep. 2013 Feb 1;13(1):110–7.

40. Pomés A, Davies JM, Gadermaier G, Hilger C, Holzhauser T, Lidholm J, et al. WHO/IUIS Allergen Nomenclature: Providing a common language. Mol Immunol. 2018 Aug;100:3–13.

41. Douladiris N, Savvatianos S, Roumpedaki I, Skevaki C, Mitsias D, Papadopoulos NG. A molecular diagnostic algorithm to guide pollen immunotherapy in southern Europe: towards component-resolved management of allergic diseases. Int Arch Allergy Immunol. 2013;162(2):163–72.

42. Curin M, Reininger R, Swoboda I, Focke M, Valenta R, Spitzauer S. Skin prick test extracts for dog allergy diagnosis show considerable variations regarding the content of major and minor dog allergens. Int Arch Allergy Immunol. 2011;154(3):258–63.

43. Chan SK, Leung DYM. Dog and Cat Allergies: Current State of Diagnostic Approaches and Challenges. Allergy Asthma Immunol Res. 2018 Mar;10(2):97–105.

44. Barber D, Moreno C, Ledesma A, Serrano P, Galán A, Villalba M, et al. Degree of olive pollen exposure and sensitization patterns. Clinical implications. J Investig Allergol Clin Immunol. 2007;17 Suppl 1:11–6.

45. Kleine-Tebbe J, Jakob T. Molecular allergy diagnostics using IgE singleplex determinations: methodological and practical considerations for use in clinical routine. Allergo J Int. 2015 Oct 1;24(6):185–97.

DOI: 10.1177/0004563219877434

46. Briggs T. Getting it Right First Time: improving the quality of orthopaedic care within the National Health Service in England. [Internet]. British Orthopaedic Association, London; 2012 [cited 2018 Nov 22]. Available from: https://www.boa.ac.uk/pro-practice/getting-it-right-first-time/

47. NHS England. NEXT STEPS ON THE NHS FIVE YEAR FORWARD VIEW [Internet]. 2017. Available from: https://www.england.nhs.uk/wp-content/uploads/2017/03/NEXT-STEPS-ON-THE-NHS-FIVE-YEAR-FORWARD-VIEW.pdf

48. NHS England. Five Year Forward View [Internet]. 2014. Available from: https://www.england.nhs.uk/ourwork/futurenhs/

49. BEIS. UK Industrial Strategy: Building a Britain fit for the future [Internet]. GOV.UK; 2017. Available from: https://www.gov.uk/government/publications/uk-industrial-strategy-a-leading-destination-to-invest-and-grow

50. BEIS. Industrial Strategy: Life Sciences Sector Deal [Internet]. GOV.UK; 2017. Available from: https://www.gov.uk/government/publications/uk-industrial-strategy-a-leading-destination-to-invest-and-grow

51. Mari A, Scala E, Palazzo P, Ridolfi S, Zennaro D, Carabella G. Bioinformatics applied to allergy: Allergen databases, from collecting sequence information to data integration. The Allergome platform as a model. Cell Immunol. 2006 Dec;244(2):97–100.

DOI: 10.1177/0004563219877434

Table Legends:

Table 1: Common routes of exposure to allergenic proteins. Protein allergens may be encountered by a variety of routes. Severity of reaction is determined by type of allergen, its native integrity and the level of exposure.

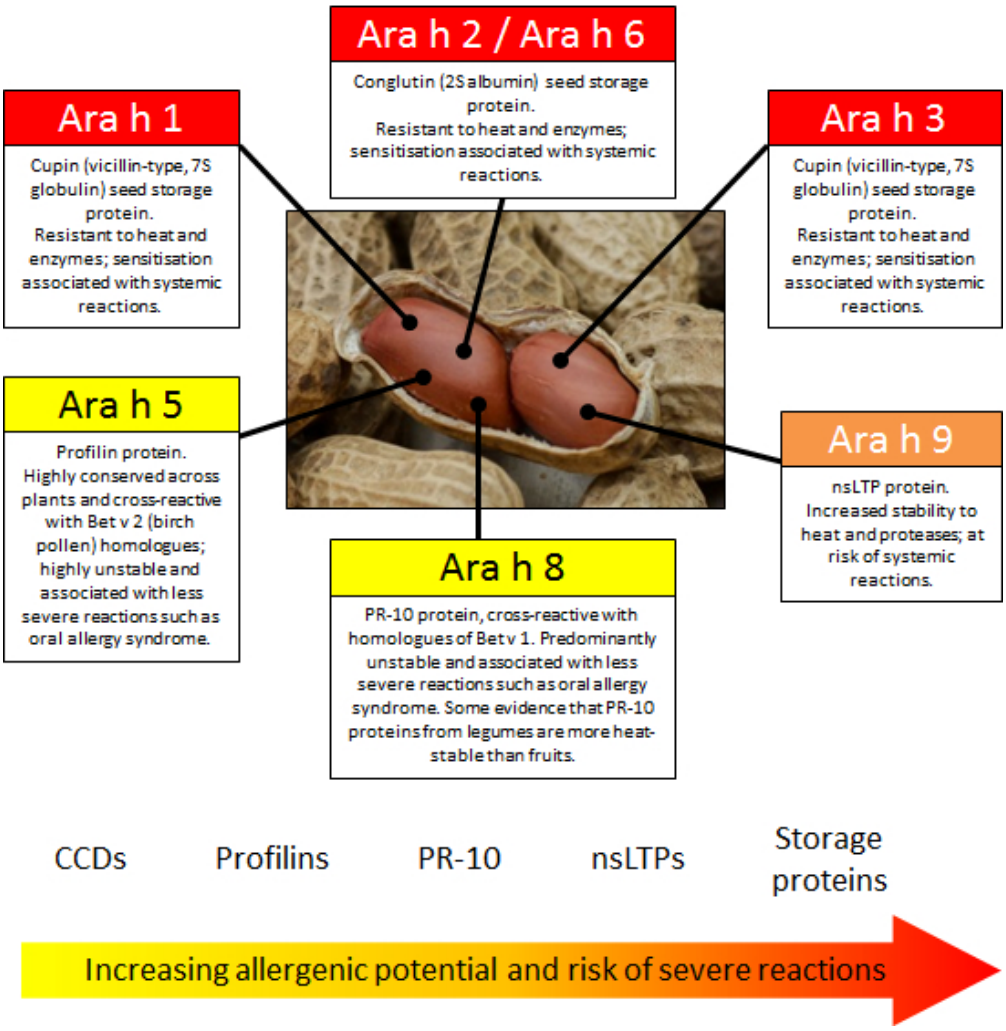
Figure Legends:

Figure 1. Allergenic potential of peanut (Arachis hypogea) components. A number of proteins have been identified as allergens within the peanut. Identification of the specific component responsible for allergic sensitisation can help to guide clinicians in stratifying patients as high- or low-risk for systemic reactions. Image obtained from sources that permit free-to-use reproduction for commercial and non-commercial purposes.

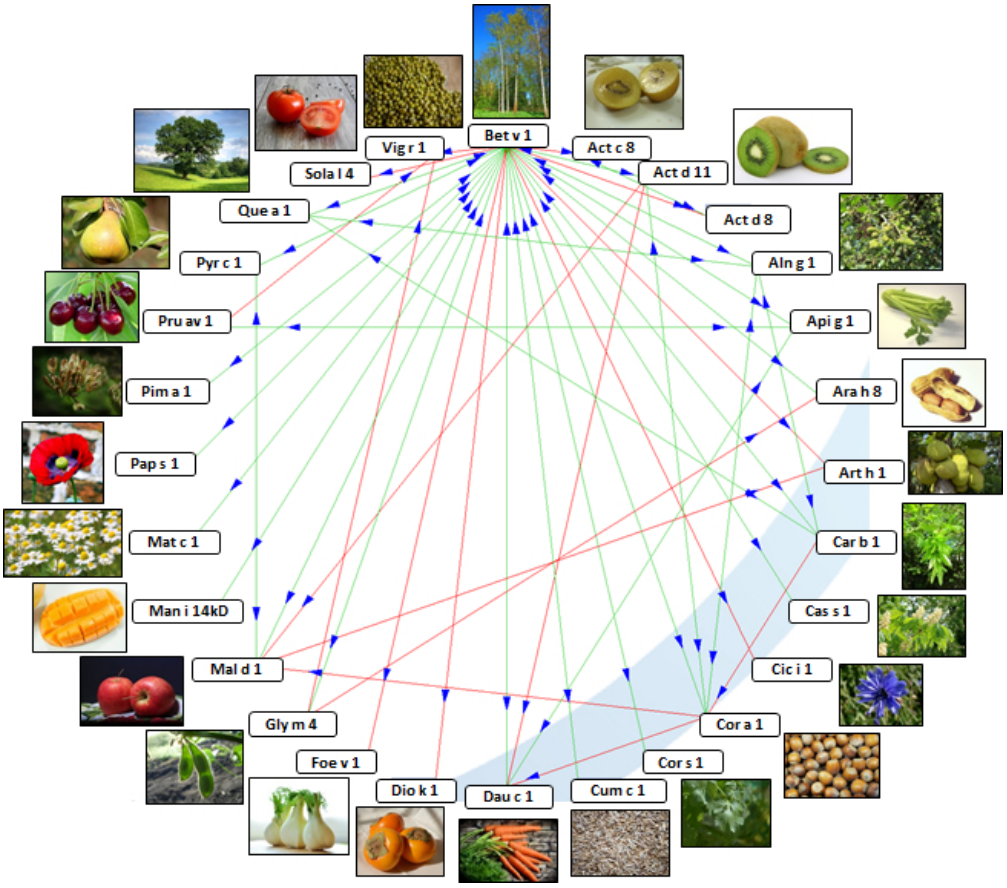
Figure 2: IgE cross-reactivity of Bet v 1. The major birch pollen allergen, Bet v 1, is a PR-10 protein. Sensitisation to Bet v 1 is associated with cross-reactivity to one or more allergenic food sources that contain related PR-10 proteins in over 70% of birch pollen allergy cases, causing oral allergy syndrome. Sensitisation to Bet v 2, a profilin protein, has a cross-reactive pattern of allergenic food sources distinct to that associated with Bet v 1. Identifying the specific allergenic component of birch pollen to which the patient has been sensitised can guide recommendations for avoidance of likely cross-reactive triggers of oral allergy syndrome caused by ingestion of foods with homologous PR-10 proteins. Reciprocal cross-reactivity between allergen components is demonstrated by double-headed arrows, and unilateral cross-reactivity is demonstrated by single-headed arrows. This figure was generated using the online Allergome O-ring tool (51); all images were obtained from sources that permit free-to-use reproduction for commercial and non-commercial purposes.

.

DOI: 10.1177/0004563219877434



DOI: 10.1177/0004563219877434



DOI: 10.1177/0004563219877434

Exposure Route	Types of Allergen	Allergic Responses
Gastrointestinal tract	Various food types (e.g. peanuts, shellfish, soy)	Oedema (lips, tongue) Vomiting
Respiratory tract	Pollen (grass, tree) Animal dander House dust mites	Rhinoconjunctivitis Allergic asthma Bronchoconstriction
Skin	Venom (bee, wasp) Injectable drugs	Pruritic rashes Urticaria Angioedema

160x48mm (96 x 96 DPI)

DOI: 10.1177/0004563219877434

Figure 1

Image	Licence	URL
Bet v 1	Free for commercial use No attribution required	https://pixabay.com/photos/birch-tree-birch-tree-birch-grove-3337239/
Act c 8	All photos on Pexels can be used for free for commercial and non-commercial use. Attribution is not required.	https://www.pexels.com/photo/fruit-golden-kiwi-yellow-755312/
Act d 11 / 8	Free for commercial use No attribution required	https://pixabay.com/photos/breakfast-cookery-cooking-food-1239438/
Aln g 1	Free for commercial use No attribution required	https://pixabay.com/photos/alnus-glutinosa-alder-common-alder-844513/
Api g 1	Free for commercial use No attribution required	https://pixabay.com/photos/celery-vegetables-healthy-vitamins-74333/
Ara h 8	Free for commercial use No attribution required	https://pixabay.com/photos/peanuts-nuts-snack-nutrition-1046136/
Art h 1	Free for commercial use No attribution required	https://pixabay.com/photos/jackfruit-tree-african-green-2108869/
Car b 1	Free for commercial use No attribution required	https://pixabay.com/photos/carpinus-betulus-european-hornbeam-846553/
Cas s 1	Free for commercial use No attribution required	https://pixabay.com/photos/horse-chestnut-flowering-3374068/
Cic i 1	CC0 Public Domain Free for commercial use	https://www.maxpixel.net/Chicory-Cichorium-Intybus-Common-Chicory-2541624
Cor a 1	Free for commercial use No attribution required	https://pixabay.com/photos/hazelnut-shell-brown-nuts-food-2923204/
Cor s 1	Free for commercial use No attribution required	https://pixabay.com/photos/real-coriander-coriander-115373/
Cum c 1	CC0 Public Domain Free for commercial use	https://www.maxpixel.net/Whole-Cumin-Pepper-Aromatic-Cumin-1049551
Dau c 1	Free for commercial use No attribution required	https://pixabay.com/photos/carrots-vegetables-harvest-healthy-2387394/
Dio k 1	Free for commercial use No attribution required	https://pixabay.com/photos/persimmons-fruit-fresh-colorful-1456186/
Foe v 1	Free for commercial use No attribution required	https://pixabay.com/photos/fennel-vegetables-fennel-bulb-food-1311691/
Gly m 4	Free for commercial use No attribution required	https://cdn.pixabay.com/photo/2015/10/20/22/13/soy-998566__340.jpg
Mal d 1	Free for commercial use No attribution required	https://pixabay.com/photos/apple-red-fruit-fruits-decoration-1506119/
Man I 14kD	Free for commercial use No attribution required	https://pixabay.com/nl/photos/mango-plak-white-geel-ge%C3%AFsoleerde-2471837/
Mat c 1	Free for commercial use No attribution required	https://pixabay.com/photos/chamomile-summer-nature-796381/
Pap s 1	Free for commercial use No attribution required	https://cdn.pixabay.com/photo/2017/07/31/18/59/opium-poppy-2560032__340.jpg
Pim a 1	Free for commercial use No attribution required	https://pixabay.com/photos/anise-seeds-gap-fruits-umbelliferae-2578534/

DOI: 10.1177/0004563219877434

Pru av 1	Free for commercial use No attribution required	https://pixabay.com/photos/cherry-sweet-cherry-red-fruit-167341/
Pyr c 1	Free for commercial use No attribution required	https://pixabay.com/photos/pear-leaves-fruit-delicious-branch-1586866/
Que a 1	Free for commercial use No attribution required	https://pixabay.com/photos/tree-oak-landscape-view-field-402953/
Sol l 4	Free for commercial use No attribution required	https://pixabay.com/photos/tomato-vegetables-eat-salt-pepper-2096306/
Vig r 1	Free for commercial use No attribution required	https://pixabay.com/photos/mung-beans-moong-beans-green-gram-390017/

Figure 2

Image	Licence	URL
Ara h	CC0	https://pxhere.com/en/photo/1048637