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| Title | Resveratrol prevents brain edema, blood-brain barrier permeability, and altered aquaporin profile in autism animal model | | | | | |
|----------|--|--|--|--|--|--|
| Type | Article | | | | | |
| URL | https://clok.uclan.ac.uk/id/eprint/38465/ | | | | | |
| DOI | https://doi.org/10.1002/jdn.10137 | | | | | |
| Date | 2021 | | | | | |
| Citation | Deckmann, Iohanna, Santos-Terra, Júlio, Fontes-Dutra, Mellanie, Körbes-Rockenbach, Marília, Bauer-Negrini, Guilherme, Brum Schwingel, Gustavo, Riesgo, Rudimar, Bambini-Junior, Victorio and Gottfried, Carmen (2021) Resveratrol prevents brain edema, blood-brain barrier permeability, and altered aquaporin profile in autism animal model. International Journal of Developmental Neuroscience, 81 (7). pp. 579-604. ISSN 0736-5748 | | | | | |
| Creators | Deckmann, Iohanna, Santos-Terra, Júlio, Fontes-Dutra, Mellanie, Körbes- Rockenbach, Marília, Bauer-Negrini, Guilherme, Brum Schwingel, Gustavo, Riesgo, Rudimar, Bambini-Junior, Victorio and Gottfried, Carmen | | | | | |

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RESVERATROL PREVENTS BRAIN EDEMA, BLOOD-BRAIN BARRIER PERMEABILITY AND ALTERED AQUAPORIN PROFILE IN AUTISM ANIMAL MODEL

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This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process which may lead to differences between this version and the Version of Record. Please cite this article as doi: 10.1002/jdn.10137

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Funding statement: This work was supported by Instituto Nacional de Ciência e Tecnologia em Neuroimunomodulação (INCT-NIM #465489/2014-1), Rio de Janeiro, Brazil; Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES) and Fundo de Incentivo à Pesquisa e Eventos do Hospital de Clínicas de Porto Alegre (FIPE-HCPA #13-0047). We would also like to thank Fluxome (Stenløse, Denmark) for the generous gift of *trans*-resveratrol.

Conflict of Interest: The authors declare that they have no conflict of interest.

Data availability statement: "Research data are not shared."

Ethical statement: All procedures were approved by the local Ethics Commission on the Use of Animals (CEUA-UFRGS 36229) and performed according to ethical principles in accordance with the NIH Guide for the Care and Use of Laboratory Animals, as well as Brazilian Arouca Law (11,794, of October 8, 2008).

Author contributions: ID, JS-T, MF-D, MK-R, GB-N, GBS, RR, VB-J and CG: experimental design and intellectual contribution. RR, VB-J and CG: acquisition of financial resources. ID, GB-N and GBS: edema and Evans blue analyses. ID, JS-T, MF-D and MK-R: immunofluorescence and western blotting analyses. ID, JS-T, MF-D, VB-J and CG: data discussion and manuscript preparation.

Abbreviations: aCC: anterior cingulate cortex (in the medial prefrontal cortex); AmR: amygdala region; AQP: aquaporin; ASD: autism spectrum disorder; BBB: blood-brain barrier; CA: cornu ammonis (CA1-CA3, in the hippocampus); CNS: central nervous system; GFAP: glial fibrillary acidic protein; IL: infralimbic cortex (in

the medial prefrontal cortex); MIA: maternal immune activation; mPFC: medial prefrontal cortex;

PrL: prelimbic cortex (in the medial prefrontal cortex); pSSA: primary somatosensory area; RSV: resveratrol; VPA: valproic acid.

Highlights:

- Autism Spectrum Disorder (ASD) is hallmarked by neuroimmune background;
- ASD animal model replicates the greater brain volume described in ASD patients;
- Resveratrol (RSV) prevents blood-brain barrier alterations and edema formation;
- RSV restore AQP4 levels to control levels;
- RSV promotes a functional amelioration in astrocytes.

Abstract

Autism Spectrum Disorder can present a plethora of clinical conditions associated with the disorder, such as greater brain volume in the first years of life in a significant percentage of patients. We aimed to evaluate the brain water content, the blood-brain barrier permeability, and the expression of aquaporin 1 and 4, and GFAP in a valproic acid-animal model, assessing the effect of resveratrol. On postnatal day 30, Wistar rats of the valproic acid group showed greater permeability of the bloodbrain barrier to the Evans blue dye and a higher proportion of brain water volume, prevented both by resveratrol. Prenatal exposition to valproic acid diminished aquaporin 1 in the choroid plexus, in the primary somatosensory area, in the amygdala region and in the medial prefrontal cortex, reduced aquaporin 4 in medial prefrontal cortex and increased aquaporin 4 levels in primary somatosensory area (with resveratrol prevention). Valproic acid exposition also increased the number of astrocytes and GFAP fluorescence in both primary somatosensory area and medial prefrontal cortex. In medial prefrontal cortex, resveratrol prevented the increased fluorescence. Finally, there was an effect of resveratrol per se on the number of astrocytes and GFAP fluorescence in the amygdala region and in the hippocampus. Thus, this work demonstrates significant changes in blood-brain barrier permeability, edema formation, distribution of aquaporin 1 and 4, in addition to astrocytes profile in

the animal model of autism, as well as the use of resveratrol as a tool to investigate the mechanisms involved in the pathophysiology of Autism Spectrum Disorder.

Keywords: Autism spectrum disorder. Valproic acid. Resveratrol. Water content. Blood-brain barrier. Aquaporin.

1. INTRODUCTION

Autism Spectrum Disorder (ASD) was first characterized in the 1920s by the Russian psychiatrist Grunya Sukharewa, who described 6 children with autistic characteristics (Ssucharewa, 1926; Zeldovich, 2018). Currently, ASD is a highly prevalent neurodevelopmental disorder characterized by 1) deficits in communication and social interaction and 2) the presence of repetitive behaviors and restricted interests/activities (APA, 2013), affecting 1:54 children up to 8 years old in the USA (Maenner et al., 2020).

Despite research advances on this disorder, the etiology of ASD remains unknown. However, epidemiological observations suggest that environmental factors, such as valproic acid (VPA), are closely related to the onset of ASD (Christensen et al., 2013; Roullet et al., 2013; Smith & Brown, 2014). Besides the core symptoms, several conditions associated with ASD are described, including greater brain volume in the first years of life (Aylward et al., 2002) affecting around 20% of ASD patients (Sacco et al., 2015).

For a long time, it was hypothesized that the Central Nervous System (CNS) was an immunologically privileged location (Tambur & Roitberg, 2005) due to blood-brain barrier (BBB), a selective barrier composed, among others, by astrocytes (Abbott, 2013). Several studies relate astrocytic dysfunctions to psychiatric disorders, including ASD (Mony et al., 2016; Zeidán-Chuliá et al., 2014). Increased reactive gliosis, proliferation of glial cells in the brain of ASD individuals (Petrelli et al., 2016) as well as the association between ASD and genes related to the activation of glia and the immune system (Voineagu et al., 2011), stressing the role of astrocytes in ASD and in BBB impairments.

Besides astrocytes, the BBB's dynamic can be affected by the water channels aquaporins (AQP). AQP1 is expressed mainly in the apical membrane of the choroid plexus (involved in the production of cerebrospinal fluid), in addition to glial membranes, astrocytes and ependymal cells (Benga & Huber, 2012; Oshio et al., 2003). On the other hand, AQP4 is the most common water channel in the CNS,

present in greater quantity in terminal feet of astrocytes that surround blood vessels (essential constituent of BBB) (Nagelhus & Ottersen, 2013; Papadopoulos & Verkman, 2007; Xiao & Hu, 2014), playing an important role in removing water from the cerebral parenchyma, in addition to assisting the potassium buffering (Benga & Huber, 2012).

Few studies have been conducted related to BBB and AQP alteration in ASD individuals, indicating decreased cerebellar AQP4 (*postmortem*) (Fatemi et al., 2008), and no alteration in serum (Kalra et al., 2015), as well as a huge cerebellar permeability in the VPA-animal model of ASD (Kumar & Sharma, 2016b, 2016a; Kumar et al., 2015). Considering the increased brain volume and the possible impairment of neural barrier systems, as well as the pro-inflammatory and pro-oxidant processes already observed in ASD individuals, molecules with antioxidant and anti-inflammatory properties become important targets for the study of neuroprotective mechanisms in ASD.

The *trans*-resveratrol (RSV, 3,5,4'-trihydroxystilbene) is a naturally occurring polyphenolic compound present in grapes, peanuts and red wine, having several biological effects (Frémont, 2000; Vang et al., 2011). Several studies emphasizing the protective and therapeutic roles of RSV in several pathologies (Berman et al., 2017; Koushki et al., 2020), highlighting the anti-inflammatory (J.-A. Lee et al., 2015; Sánchez-Fidalgo et al., 2010), antioxidant (Mohammadshahi et al., 2014) and neuroprotective effects (Quincozes-Santos & Gottfried, 2011; Tang, 2010) of RSV.

Our research group have been demonstrated the preventive effect of prenatal treatment with RSV in sociability and sensory deficits in the offspring of VPA-animal model (Bambini-Junior et al., 2014; Fontes-Dutra et al., 2018), as well as in microRNA levels (Hirsch et al., 2018). These results enable the use of RSV as both a reliable method for understanding the pathophysiology of ASD and an assisting tool in the study of biological routes and structures involved in its etiology.

The mechanisms that mediate the brain volume dynamics are largely unknown; thus, we proposed an investigation of factors possibly associated with the formation of brain edema in ASD in an animal model of autism. Thus, we aimed to evaluate the proportion of brain fluid volume, the BBB permeability, as well as to analyze AQP 1 and 4 and GFAP+-astrocytes in 30-day-old animals of the animal model of autism induced by prenatal exposure to VPA, evaluating the possible therapeutic effect of RSV.

2. EXPERIMENTAL PROCEDURE

2.1 Animals

Wistar rats from the Center for Reproduction and Experimentation of Laboratory Animals (CREAL-UFRGS) of the Federal University of Rio Grande do Sul (UFRGS) were used, kept in standard animal facilities conditions, with food and water *ad libitum*, light/dark cycle 12 hours, constant temperature (22°C ± 1°C) and a maximum of four animals per housing box. All procedures were approved by the local Ethics Commission on the Use of Animals (CEUA-UFRGS 36229) and performed according to ethical principles in accordance with the NIH Guide for the Care and Use of Laboratory Animals, as well as Brazilian Arouca Law (11,794, of October 8, 2008).

The animals' euthanasia procedure followed the Euthanasia Practice Guidelines of the National Council for Animal Experimentation Control (Normative Resolution N. 13, 2013). Euthanasia was performed by anesthetic overdose with ketamine and xilasine, supplied in concentrations three times higher (300 mg/kg and 40 mg/kg, respectively) than the concentration required to obtain an anesthetic-surgical plan.

2.2 Animal model and RSV treatment

The animals were mated overnight and, in the next morning, the presence of sperm was verified in the vaginal canal of the females; when fertilization was confirmed, the embryonic day 0.5 (E0.5) was determined. From E6.5 to E18.5, pregnant rats received RSV (trans-resveratrol, Fluxome, Stenløse, Denmark - 3.6 mg/kg) or dimethylsulfoxide (DMSO P.A. - vehicle, equivalent volume of RSV injection in a proportion 1:1 of DMSO and saline) subcutaneously. At E12.5, pregnant rats received a single injection of VPA (sodium valproate, Sigma-Aldrich, USA - 600mg/kg) or saline solution (0.9% - vehicle) via intraperitoneal (i.p.), as previously described (Bambini-Junior et al., 2014). On postnatal day 21 (P21), the litter was weaned and, at P30, male rats were euthanized by anesthetic overdose. The total number of animals used in the study was 24 control, 28 RSV, 25 VPA, and 22 RSV+VPA divided randomly among experiments, generated from the following number of dams: 5 control, 7 RSV, 14 VPA, and 12 RSV+VPA. As we use a maximum of 1 male from the same litter in each group, the excess offspring per litter was destined to other projects in the lab). The litters were randomly divided so that sibling animals were not part of

the same experiment (n of animals is n of litters). The loss rate for the VPA groups was 50% in this protocol.

2.3 Tissue preparation and analysis

2.3.1 Brain water content

Immediately after the euthanasia, the brains were removed, weighed, and placed in a drying oven at 60°C. After 72h, brains were reweighted and the brain water content was measured considering the difference between wet tissue weight (w) and dry weight (d) (Wei et al., 2015). We considered the animal body weight since the animals prenatally VPA-exposed presented lower body weight throughout the development as described by Schneider et al., 2005 (Schneider & Przewłocki, 2005) as follows: {[(wet weight - dry weight)/wet weight]×100/animal body weight}.

2.3.2 Evans blue dye permeability

The animals were injected via i.p. with Evans blue 2% solution (4 mg/kg) diluted in saline solution 0,9% (Kumar et al., 2015) and, after 2 hours, were anesthetized and subjected to transcardiac perfusion with saline solution and 4% paraformaldehyde. The brain was removed, post-fixed in 4% paraformaldehyde and preserved in sucrose (15% and 30%). The tissues were kept in an ultrafreezer (-80°C) until coronal slices (25 µm) were made in cryostat (Leica Microsystems). The brain coordinates following Paxinos Atlas (5th edition): bregma 3.72/3.24 (medial prefrontal cortex - mPFC) and -2.92/-3.00 (primary somatosensory area - pSSA, amygdala region - AmR, hippocampus and choroid plexus).

The slices were also incubated with DAPI solution for marking nuclei (diluted in 1:10,000 in saline solution 0.9%) for 10 minutes, followed by 5 washes with 0.1 M pH 7.4 PBS (3 minutes each) and adding the Fluoshield® mounting medium and the coverslip. The images were obtained in a 20x magnification using a confocal microscope (Olympus FV1000 – Olympus FluoView 4.0 Viewer) at the Center for Microscopy and Microanalysis (CMM-UFRGS) and the fluorescence was analyzed using the ImageJ® software.

2.3.3 Immunofluorescence

After anesthesia, the animals were euthanized by transcardiac perfusion with saline solution 0.9% and 4% paraformaldehyde, the brain was removed, preserved and cut as already described.

The technique was performed according to a previous protocol (Fontes-Dutra et al., 2018) and followed the following steps: 1) tissue permeabilization with PBS-Triton 0.1% or 0.3% (according to primary antibody); 2) 3 washes with PBS; 3) blocking with PBS-Triton 0.1% (or 0.3%) containing 5% bovine serum albumin (BSA); 4) incubation with primary antibodies for 48h at 4°C in PBS-Triton 0.1% (or 0.3%) BSA 1% solution; 5) 5 washes with PBS buffer; 6) incubation with secondary antibodies for 2 hours at room temperature; 7) 5 washes with PBS; 8) incubation with DAPI solution (1:10,000 - 10 minutes); 9) 5 washes with PBS followed by addition of Fluoshield® mounting medium and coverslip. The images were obtained as previously described. Two trained researchers did manual analysis of both fluorescence distribution and cell counting in the brain regions and subregions, blinded for the experimental groups, using the ImageJ® software.

All primary antibodies were chosen according to the previous data from references cited in the manufacturer datasheets. All reagent information were detailed in Supplementary Table 1. Representative images of the AQP1 and AQP4 labeling can be seen in Supplementary Figure 1 and Supplementary Figure 2, respectively.

2.3.4 Western blotting

Following to the euthanasia, the brain was removed and the mPFC, pSSA, AmR and hippocampus were dissected.

The samples were homogenized in a buffer containing protease inhibitor, 10% SDS, EDTA 100 mM, TRIS/HCl buffer 500 mM pH 8. The total proteins were quantified by the Lowry method (Lowry et al., 1951) and the samples prepared in a buffer containing glycerol, bromophenol blue, TRIS/HCl buffer and β-mercaptoethanol. 40μg of protein was applied in 10% polyacrylamide gel, separated by one-dimensional electrophoresis and transferred to nitrocellulose membranes for the detection of the AQP1 and AQP4 immunocontents. The membranes were blocked in 5% BSA dissolved in a pH 7.5 TRIS buffer (TBS) with 0.1% Tween-20 (TTBS) and incubated overnight at 4°C with the primary antibodies.

After incubation, the membranes were washed with TTBS and incubated with secondary antibodies, followed by 3 washes. The substrate SuperSignal® West Pico (Thermo Fisher Scientific) was used on the membranes and the chemiluminescent signal was detected using ImageQuant LAS 4000 (GE HealthCare Life Sciences). The quantification of the relative immunocontent was performed with the ImageJ software (v. 1.51) and the data was normalized by the housekeeping protein β -actin. All reagent information were detailed in Supplementary Table 1.

2.4 Statistical analysis

The data were analyzed using the IBM SPSS Statistics 20 program (IBM SPSS, Armonk, NY, USA). Kolmogorov-Smirnov and Shapiro-Wilk tests of normality were applied to determine data distribution. The data of "edema" and "BBB permeability to Evans blue dye" had a non-normal distribution; therefore, a non-parametric test was performed for independent samples (Kruskal-Wallis). Immunofluorescence and western blotting data showed normal data distribution, using the two-way ANOVA test followed by Sidak's post-test. When there was an interaction effect, pairwise comparison was analyzed in the post hoc; when there was no effect, the effect of exposure to factors (VPA or RSV) was analyzed.

The graphs were made using the GraphPad Prism 6 program. Data were reported as median ± interquartile range (IQR) for the non-parametric test and mean ± standard deviation for the parametric test. p <0.05 was considered statistically significant.

3. RESULTS

3.1. Prenatal administration of RSV prevents the alterations induced by prenatal exposure to VPA in body weight and in proportional brain water content at P30

Prenatal exposure to VPA increased brain water content (p = 0.003, Figure 1A), decreased body weight (p = 0.001, Figure 1B) and increased proportion of brain fluid (p = 0.002, Figure 1C). In this experiment, prenatal treatment with RSV was able to prevent these alterations. Detailed statistics are shown in Table 1.

3.2. Prenatal administration of RSV prevents the increased BBB permeability induced by VPA at P30

The VPA group had increased BBB permeability to Evans blue dye (representative images in Figure 2) in the choroid plexus (p = 0.009), in the pSSA, both in layers II/III (p = 0.004) and IV/V (p = 0.005), in subregions of the mPFC when compared to the control and/or RSV group) anterior cingulate cortex (aCC) (II/III: p = 0.001; IV/V: p = 0.013); 2) prelimbic cortex (PrL) (II/III: p = 0.019; IV/V: p = 0.022); and 3) infralimbic cortex (IL) (II/III: p = 0.028; IV/V: p = 0.016). RSV was able to prevent the permeability alterations in these regions. No significant difference was observed in dye permeability in the hippocampus (p = 0.134) or in the AmR (p = 0.050). Detailed statistics are shown in Table 2.

3.3. Prenatal exposure to VPA changes choroid plexus morphology and decreases AQP1 distribution at P30

The choroid plexus from both groups exposed to VPA had a huge morphological alteration (indicated by the white arrow in Figure 3A) and decreased AQP1 labeling (Figure 3A-3B, control: 1244 ± 279.3 ; RSV: 1053 ± 131.9 ; VPA: 867.1 ± 76.11 ; RSV+VPA: 798.6 ± 221.3 ; F (3, 16) = 5.345, p VPA = 0.0022). These alterations were not prevented by RSV.

3.4. Prenatal administration of RSV prevents the VPA-induced increase in AQP4-immunocontent and distribution in pSSA at P30.

Prenatal exposure to VPA decreased AQP1 content in pSSA (Figure 4A, Control: 1.594±0.073; RSV: 1.342±0.210; VPA: 1.148±0.180; RSV+VPA: 0.834±0.185; F (1, 12) = 31.39; p VPA = 0.0001) in deep layers (II/III; p interaction = 0.9098; IV/V: p VPA = 0.0005) with no preventive effect by RSV (Table 3). Interestingly, the VPA group had increased AQP4 content in pSSA (Figure 4E, control: 1.445±0.0256; RSV: 1.086±0.149; VPA: 2.391±0.368; RSV+VPA: 1.457±0.314; F (1, 12) = 5.128; p interaction = 0.0429) in deep layers (II/III: p interaction = 0.4255; IV/V: p interaction = 0.0011) with a significant preventive effect of RSV. Detailed statistics of immunofluorescence are shown in Table 3.

3.5. mPFC has decreased levels of both AQP1 and AQP4 induced by prenatal exposure to VPA

Despite no change was observed in the AQP1 immunocontent in mPFC from VPA group (Figure 4C, control: 1.221 ± 0.185 ; RSV: 1.124 ± 0.182 ; VPA: 1.081 ± 0.389 ; RSV+VPA: 0.975 ± 0.195 ; F (1, 12) = 0.00095; p interaction = 0.9759), both groups exposed prenatally to VPA presented decreased distribution of this protein in all subregions of the mPFC, with no preventive effect of RSV: aCC – layers II/III (p VPA = 0.0043) and IV/V (p VPA = 0.0010); PrL – layers II/III (p VPA = 0.0010) and IV/V (p VPA < 0.0001); IL – layers II/III (p VPA = 0.0009) and IV/V (p VPA = 0.0004) (Table 4).

The VPA group had decreased content of AQP4 (Figure 4G, control: 0.816±0.196; RSV: 1.019±0.130; VPA: 0.387±0.164; RSV+VPA: 0.946±0.507; F (1, 12) = 6.808; p RSV = 0.0228) with decreased distribution of this protein in all subregions of the mPFC: aCC – layers II/III (p interaction = 0.0421) and IV/V (p interaction = 0.0125); PrL – layers II/III (p interaction = 0.0449) and IV/V (p interaction = 0.0167); IL – layers II/III (p interaction = 0.0412) and IV/V (p interaction = 0.0171). Detailed statistics of immunofluorescence are shown in Table 4.

3.6. Intrauterine exposure to VPA induces a decrease in AQP1 in the AmR

Despite no differences were observed in AQP1 immunocontent in the AmR among experimental groups (Figure 4B, Control: 0.918±0.214; RSV: 1.089±0.399; VPA: 0.871±0.113; RSV+VPA: 0.842±0.277; F (1, 12) = 0.6424; p interaction = 0.4384), both groups that received VPA had decreased distribution of AQP1 (p VPA = 0.0004) (Table 5). No changes were observed in both AQP4 content (Figure 4F, Control: 1.192±0.457; RSV: 1.288±0.333; VPA: 0.943±0.108; RSV+VPA: 1.512±0.521; F (1, 12) = 1.484; p interaction = 0.2466) and in distribution among groups (p interaction = 0.5988). Detailed statistics of immunofluorescence are shown in Table 5.

3.7. Prenatal exposure to VPA does not alter the expression profile of AQP1 and AQP4 in the hippocampus, but the RSV treatment had a *per se* effect

No changes were observed in the hippocampal content of AQP1 (Figure 4D, Control: 1.031 ± 0.275 ; RSV: 0.927 ± 0.273 ; VPA: 0.941 ± 0.214 ; RSV+VPA: 0.977 ± 0.238 ; F (1, 12) = 0.3097; p interaction = 0.5881). However, a *per se* effect of

RSV was observed, with decreased AQP1 distribution in dentate gyrus and CA2 region: Dentate gyrus (p RSV = 0.0412); CA1 (p RSV = 0.0518); CA2 (p RSV = 0.0330); CA3 (p RSV = 0.0810) (Table 6).

No changes were observed in both AQP4 content (Figure 4H, Control: 0.533 ± 0.271 ; RSV: 0.422 ± 0.221 ; VPA: 0.291 ± 0.106 ; RSV+VPA: 0.360 ± 0.116 ; F (1, 12) = 0.8866; p interaction = 0.365) and distribution among groups: dentate gyrus (p interaction = 0.0387); CA1 (p interaction = 0.0786); CA2 (p interaction = 0.1113); CA3 (p interaction = 0.0490). Detailed statistics of immunofluorescence are shown in Table 6.

3.8 RSV treatment improves the functional but not morphological aspect of astrocytic changes induced by prenatal exposure to VPA

3.8.1 Primary somatosensory area

An increased number of GFAP+-astrocytes was observed in layers II/III (p VPA = 0.0038) and in layers IV/V (p VPA = 0.0038) in both groups that were prenatally exposed to VPA (representative image in Figure 5). As expected, this data was reflected by increased GFAP immunofluorescence per area (II/III: p VPA = 0.0270; IV/V: p RSV = 0.0380). Detailed statistics are shown in Table 7.

3.8.2 Medial prefrontal cortex

This region showed different effects between the upper (II/III) and deeper (IV/V) layers. In upper layers, exposition to VPA increases the number of astrocytes, with no preventive effect of RSV (aCC: p VPA = 0.0135; PrL: p VPA = 0.0269; IL: p VPA = 0.0278), which was reflected in the GFAP immunofluorescence values (aCC: p VPA = 0.0379; PrL: p VPA = 0.0604; IL: p interaction = 0.5621).

In deeper layers, both groups exposed to VPA increased the number of GFAP+-astrocytes (aCC: p VPA = 0.0091; PrL: p VPA = 0.0004; IL: p VPA = 0.0018). However, we observed increased GFAP immunofluorescence in the VPA group, with a significantly preventive effect of RSV (aCC: p interaction = 0.0171; PrL: p interaction = 0.2853; IL: p interaction = 0.0455). Detailed statistics are shown in Table 8.

3.8.3 Amygdala region

In this region, we observed a *per* se effect of RSV, with decreased number GFAP+-astrocytes (p interaction = 0.0017) and GFAP immunofluorescence (p interaction = 0.0450) in RSV group. Detailed statistics are shown in Table 9.

3.8.4 Hippocampus

Another *per se* effect of RSV was observed in number of GFAP+-astrocytes (dentate gyrus: p interaction = 0.0162; CA1: p interaction = 0.1926; CA2: p RSV = 0.0001; CA3: p interaction = 0.0461) and GFAP immunofluorescence values (dentate gyrus: p interaction = 0.0096; CA1: p interaction = 0.0172; CA2: p interaction = 0.0214; CA3: p interaction = 0.0058). Detailed statistics are shown in Table 10.

4. DISCUSSION

A significant percentage of ASD patients presents increased brain volume in the first years of life, followed by an apparent normalization of this volume in late childhood (Aylward et al., 2002; Bartholomeusz et al., 2002; Emerson et al., 2017; Hazlett et al., 2011). Recent evidence in animal models has been highlighting the association of maternal inflammatory processes during critical embryonic development with excessive brain growth and with the triggering of ASD-associated behavior in the offspring (Le Belle et al., 2014). In fact, maternal immune activation (MIA) contributes to the onset of several neuropsychiatric disorders, including ASD (Estes & McAllister, 2016). Thus, we postulate that the fingerprint caused by prenatal exposure to VPA could involve mechanisms of MIA since animals from VPA-animal model present enhanced levels of IL-1 β , IL-6, and TFN- α in the hippocampus and other brain regions (Deckmann et al., 2018), besides enhanced TFN- α levels and microglial activation after prenatal VPA exposure (Zamberletti et al., 2019).

In the present study, we demonstrated that the prenatal exposure to VPA increased the absolute brain water content, providing a clearer overview of the higher liquid volume in the brain of the VPA group even though the body structure was smaller. The preventive effect of RSV against these changes, in addition to the alterations in the proteins AQP1 and AQP4 according to brain region, opens new clues about the mechanisms associated to the brain volume changes in ASD patients.

The VPA group presented evident BBB permeability in brain regions related to the neocortex: choroid plexus (directly in contact with neocortex), pSSA (layers II/III

and IV/V) and all subregions of the mPFC (aCC, PrL and IL, in superficial and deeper layers). In all of these regions, RSV treatment was able to prevent the BBB permeability. BBB damage can be a pivotal event for brain edema development, which could be both initiated and regulated by several pro-inflammatory mediators (among them cytokines and chemokines) that coordinate the extent of leukocyte entry to the brain parenchyma, causing loosening of the tight junctions and vasogenic edema (Stamatovic et al., 2006). RSV, known by its antioxidants and anti-inflammatory properties, could be acting as a neuroprotective molecule in different pathways during embryonic development. In a model of cerebral ischemia-reperfusion, RSV attenuates BBB dysfunctions and reverses the brain water accumulation by the regulation of matrix metallopeptidase 9 (MMP-9) (Wei et al., 2015). MMP-9 is an enzyme with zincdependent proteolytic activity that has the ability to break down collagen IV (which composes basal lamina) and whose increased levels were associated with neurodevelopmental disorders, including ASD (Reinhard et al., 2015). Both VPA and RSV are able to act in an epigenetic way, mainly modulating the histone activity. The histone deacetylase inhibition (HDACi) - an effect of VPA - is able to induce several effects on the BBB stabilization by the deregulation of important transcription factors associated with BBB formation like SOX7, SOX18, TAL1, and ETS1 (Roudnicky et al., 2020), as well as is known to interfere in the immune system, inducing increased transcription of proinflammatory genes associated with the NFkappaB pathway, for example (Rahman et al., 2004). This is important since inflammatory mediators are known to increase the BBB permeability, leading to an inflammatory infiltrate in the CNS, and are associated with neurodevelopmental disorders, such as ASD. Complementary, VPA-prenatal exposure promotes systemic inflammation, and is known that the maternal immune activation (MIA) animal model is associated with BBB disruption (Simões et al., 2018). The early treatment with RSV (beginning at E6.5) probably reduced BBB alterations by the attenuating HDACi induced by VPA through the activating Sirt and promoting modulations of proteins like MMP9 and TIMP1 (Moussa et al., 2017; Sawda et al., 2017; Wei et al., 2015). Therefore, RSV acts as a stabilizer of the transcription, preventing both BBB cell alterations (directly) and the shift for a proinflammatory status in the immune system (indirectly).

We investigated the expression and distribution of aquaporins, important water channels, in different regions of CNS. These proteins perform several roles, but one of the main ones is to facilitate the movement of water, both in and out of the CNS (Rosu et al., 2019). The VPA group decreased the AQP1 distribution in the choroid plexus, in the deeper layers of pSSA, in the AmR and in all of the subregions of mPFC analyzed. Choroid plexus is crucial given its role in the production and release of CSF and it has been shown that in animals knocked out for AQP1 there was a reduction of up to 25% in the rate of cerebrospinal fluid secretion (Oshio et al., 2003). In addition, a morphological alteration was observed in the insertion of the choroid plexus in the third ventricle. A similar lesion was observed in a model of cerebral ischemic edema (Akdemir et al., 2016), but here in the present study, we do not consider any association between this morphological alteration and the pathophysiology of ASD, which may simply be a teratogenic effect of VPA *per se*.

The VPA group increased the AQP4 content in deeper layers of the pSSA (prevented by RSV) whereas the mPFC presented decreased levels without prevention by the RSV. Alterations on the AQP4 profile in *postmortem* brain tissue of ASD individuals have already been reported (Fatemi et al., 2008), including a discrete reduction in Broadmann area 9 (BA9 - equivalent of the frontal cortex) and an increase in the BA40 (parietal cortex, where is the pSSA). They also showed increased connexin 43 levels (a protein present in astrocytic gap junction) in BA9, representing an increase in neuroglial signaling and an improvement of cell-cell communication in the frontal lobe (an integrative area) (Fatemi et al., 2008). Moreover, AQP4 knockout mice have reduced brain swelling in cytotoxic edema, whilst there is a significantly worse result in the case of vasogenic brain edema (Papadopoulos & Verkman, 2007). In the VPA model, it is still not elucidated what type of brain edema is present. Lastly, there are findings linking AQP4 with neuroimmune modulation (Ikeshima-Kataoka, 2016), which represents an important clue in the ASD pathophysiology, since the immune component of this disorder is both relevant and well established (Gottfried et al., 2015).

Noteworthy, as there are few studies on the dynamics of AQP1 and AQP4 in ASD, we think it is important to clarify some points. Despite the common sense that AQP4 is more widely expressed in the brain than AQP1, we observed higher fluorescence levels in AQP1 compared to AQP4 in the control groups in some regions (pSSA and AmR). In astrocytes, the distribution of AQP4 is predominantly in the endfeet projections surrounding vessels; however, its brain concentration varies according to region, with higher levels in the cerebellum and lower expression in the hippocampus, diencephalons, and cortex (Hubbard et al., 2015). Undoubtedly, AQP1

is mainly expressed in the choroid plexus; nevertheless, it is also expressed under normal conditions in other brain structures, such as the brain stem, cerebellum, brain cortex, hippocampus, hypothalamus, and olfactory bulb (Li et al., 2020; Qiu et al., 2014). Beyond that, studies demonstrate neuronal localization of AQP1 in mouse cortical slices, as well as increased cortical levels of AQP1 in Alzheimer's disease models. Moreover, this same work also demonstrates that wild-type animals at 60 days old, present higher amounts of AQP1 protein than AQP4 in cortical homogenates (Park et al., 2021). Finally, in the amygdala region, the human protein atlas demonstrates the mRNA expression of AQP1 (The Human Protein Atlas, n.d.). These studies indicate that the concentration and distribution of AQP 1 and 4 vary according to cell type/domain and brain region, which corroborates our present study regarding the brain region.

The increased GFAP-immunofluorescence and number of GFAP+-astrocytes in mPFC and pSSA by prenatal exposure to VPA corroborate with previous studies showing neuroglial activation in both ASD patients and animal models (Bristot Silvestrin et al., 2013; Edmonson et al., 2014; Vargas et al., 2005; Zhao et al., 2019). Here, we observed an important preventive effect of RSV in the mPFC, demonstrated in the decrease of GFAP immunofluorescence. Based on previous data, that indicate the neuroprotective effect of lower doses of RSV in hippocampus slices (emphasizing the important role of RSV in improving glutamate uptake by astrocytes and modulate the synaptic plasticity) (Bobermin et al., 2012; De Almeida et al., 2008; Quincozes-Santos et al., 2013; Quincozes-Santos & Gottfried, 2011), and an improvement in neuroinflammation (an ASD hallmark) in an ASD-animal model (Ahmad et al., 2018; Bhandari & Kuhad, 2017), it would be possible that also in this context, we have a beneficial effect on astrocyte metabolism and function, since this treatment is effective in ameliorates several behavioral impairments in VPA animal model (Bambini-Junior et al., 2014; Fontes-Dutra et al., 2018; Hirsch et al., 2018).

Despite some studies demonstrated no alterations in astrocyte parameters in ASD post mortem tissues (T. T. Lee et al., 2017; Morgan et al., 2014), animal models of fragile X syndrome (a disorder with a high prevalence of ASD) present a specific disruption in the constitution of the deeper layers, besides presenting an increased number of astrocytes (F. Lee et al., 2019). Therefore, alterations in the laminar constitution could influence directly the distribution of astrocytes. The dynamics of cortical disorganization is widely described in ASD. An event like acute

neuroinflammation, with increased levels of brain cytokines, may contribute to synaptic reorganization, which results in long-term alterations regarding hyperexcitability of the whole neural circuitry (Clarkson et al., 2017). One of the most relevant findings in patients with ASD is the identification of disturbance in the organization of the minicolumns (Casanova, 2007) and the presence of patches with loss of layer delimitation in the cortex (DeNardo et al., 2015; Stoner et al., 2014), being the deeper cortical layers the most affected.

In hippocampus and AmR, we observed only effects of RSV treatment. A possible explanation for this effect observed in the amygdala region is that the amygdalar nuclei originate at different times between embryonic day E10-E12 in rats, before the induction of the animal model (in E12.5), and during the prenatal treatment with RSV (between E6.5 to E18.5) (Soma et al., 2009). Although the embryonary origin of the hippocampus starting from E15 (Hayashi et al., 2015), the effects of prenatal exposure to VPA seem to be progressive and of late-onset (Santos-Terra, *unpublished observations*). Despite being a molecule with important neuroprotective effects already described, RSV was able to cause changes in the hippocampus at P30. Considering the progressive effect of VPA in adulthood, maybe RSV develops an earlier cellular background to better support the progressive damage induced by VPA.

One of the major theories regarding ASD pathophysiology refers to the electrophysiological changes, mainly the imbalance between excitation and inhibition. The presence of epilepsy or seizure episodes in approximately 30% of individuals with ASD reinforces the excitatory profile predominant in ASD (Spence & Schneider, 2009). In response to this hyperexcitability and chronic neuroinflammation, might be observed proliferation and hypertrophy of the astrocytes, which acquire a reactive profile (Poskanzer & Molofsky, 2018), due to several roles, including K + buffering. The extracellular K⁺ is critical for defining the resting potential of neurons and astrocyte membranes, and mechanisms for removing this ion from the synaptic cleft are vital to maintaining cerebral homeostasis (Bellot-Saez et al., 2017). One mechanism of K⁺ uptake by glial cells is through the action of internal rectifying channels of K⁺ (Kir) (Olsen et al., 2015). This is particularly important since AQP4 and Kik4.1 are highly overlapping channels in the astrocytic end-feet (Strohschein et al., 2011) and probably AQP4 is required to sustain efficient K+ clearance, considering the association of water flux alteration and increasing intensity of epileptic seizures (Amiry-Moghaddam et al., 2003), and a delay in K+ buffering in AQP4-null mice (Lu et al., 2008).

Considering all data, we hypothesized that the brain impairments induced by the VPA model include a neuroinflammation background triggered in the developing brain of the embryo, which contributes to the increased BBB permeability (and consequently edema due to the entry of water and inflammatory infiltrate). In consequence, there is a decrease in the levels of AQP1 and AQP4 to maintain water homeostasis in the brain. In parallel, neuroinflammation triggers the excitotoxicity process, leading to a reactive astrocytic phenotype. The increased astrocytic activity leads to an increased need for K+ buffering, which in turn increases Kir4.1 and, consequently, AQP4 levels in a region-specific manner (which, in turn, could be the main onset to brain edema formation). The fingerprinting caused by VPA happens in multiple regions; since pSSA is a primary processing area, both the impact caused by VPA and the prevention mechanisms by RSV may be more expressive and less complex than those occurring in mPFC, an associative and more complex region. Here, RSV prevents successfully the impairments regarding BBB permeability and the increase of AQP4 in the pSSA, as well as decreases GFAP antibody labeling in the mPFC, indicating a lower glial reactivity. Thereby, based on several shreds of evidence that point to RSV as a stabilizer of the neural environment, RSV could also normalize K+ levels and restructure synaptic connections in pSSA considering the colocalization of AQP4 and Kir4.1 channels.

5. CONCLUDING REMARKS

In summary, we demonstrated that prenatal exposure to VPA alters the bodyweight of the animals, as well as induces brain edema, and increases the permeability of BBB. In addition, there was an altered AQP profile in region-dependent in VPA-exposed animals and GFAP augmented expression. RSV was able to prevent important changes in GFAP+ astrocytes and in AQP4 in the pSSA. The neuroprotective role of the RSV in this model shed some light on pathways possibly associated with the alterations induced by VPA along with neuroimmune changes also observed in ASD individuals. Taken together, the present data emphasize the investigation of the mechanisms involved in the neuroimmunological issues as a promising strategy in the understanding of biological pathways in ASD pathophysiology.

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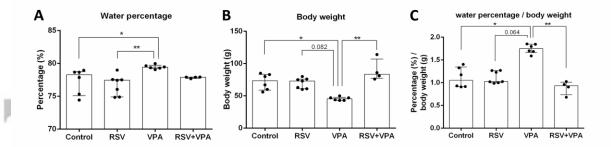


Figure 1 – Brain water content in an animal model of autism and the resveratrol effect (RSV). A) Percentage of water in the whole brain through the difference in wet weight and dry weight. B) Body weight, showing that the animals in the VPA group have lower weight throughout development. C) Proportion of brain fluid volume corrected by the difference in body weight of the animals. Values are shown in median±IQR. Statistical analyses: Kruskal-Wallis. N_{CON} : 5, N_{RSV} : 6, N_{VPA} : 6, $N_{RSV+VPA}$: 4. * p <0.05; ** p <0.001.

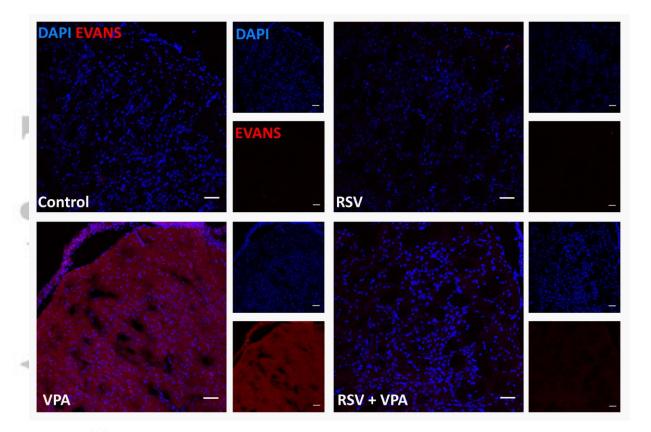


Figure 2 – Permeability of the blood-brain barrier to Evans blue dye. A) Representative images of the anterior cingulate cortex (II/III layers). The nuclear dye DAPI is stained in blue and the fluorescence of Evans blue in red. Scale bar: 50 μ m. Statistical analyses: Kruskal-Wallis. N_{CON}: 5, N_{RSV}: 7, N_{VPA}: 4, N_{RSV+VPA}: 4.

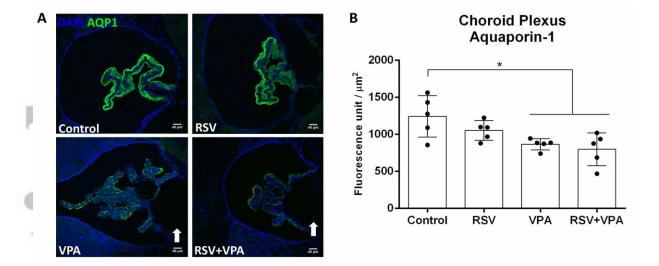


Figure 3 – Distribution of aquaporin 1 (AQP1) in the choroid plexus. (A) Representative images of the immunofluorescent label and (B) graphic representation of the expression of AQP1 in the choroid plexus. Scale bar: 40 μ m. Values are shown as mean of fluorescence \pm standard deviation. Statistical analysis: two-way ANOVA followed by Sidak. N = 5. * p <0.05. AQP1 is marked in green and nuclear DAPI dye in blue.



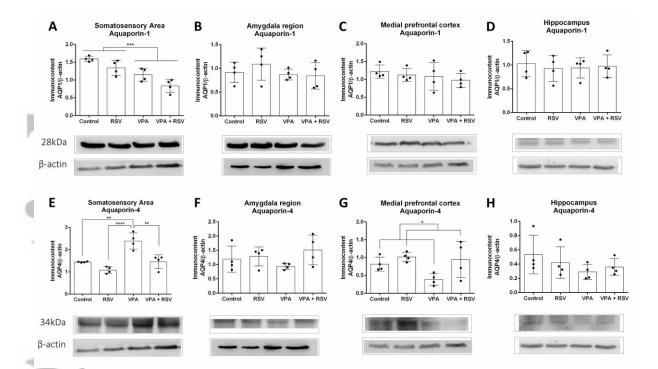


Figure 4 - Immunocontent of aquaporin 1 (AQP1) and AQP4 in different regions of the Central Nervous System. Quantification of AQP1 and AQP4 immunostaining (A)(E) in the primary somatosensory area, (B)(F) in the amygdala region, (C)(G) in the medial prefrontal cortex and (D)(H) in the hippocampus, respectively. The immunocontent of both AQP1 and AQP4 was normalized by the β-actin loading control. Values are shown as mean \pm standard deviation. Statistical analysis: Statistical analysis: two-way ANOVA followed by Sidak. N = 4. * p <0.05.

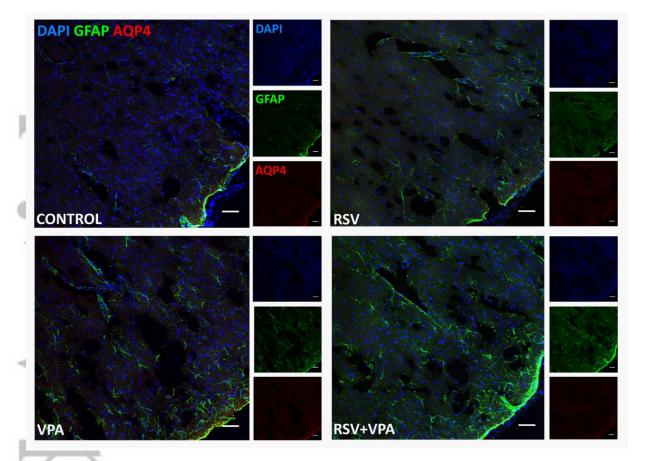


Figure 5 - Analysis of astrocytes in the primary somatosensory area. Representative images of the immunofluorescent labeling of DAPI, GFAP and AQP4 in the primary somatosensory area (II/III) in the different experimental groups. Statistical analysis: two-way ANOVA followed by Sidak. N_{CON} : 5, N_{RSV} : 6, N_{VPA} : 6, $N_{RSV+VPA}$: 5. GFAP is marked in green, AQP4 in red and DAPI nuclear dye in blue. Scale bar: 50 µm.

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Table 1 – Descriptive statistics of body weight and in proportional brain water content

| | CONTENT | | | | | |
|-----|---|-------------------|---------------|---|----------------------|---------|
| | Median (IQR) | Chi-Square, df | p Value | Mean rank | Pairwise comparisons | |
| % | | 14.019, 3 | 19, 3 0.003** | CON: 10.00 RSV: 6.86 VPA: 20.50 RSV+VPA: 11.25 | CON vs RSV: | >0.999 |
| | CON: 78.27 (75.10±78,74) | | | | CON vs VPA: | 0.044* |
| | RSV: 77.44 (74.89±77.56) | | | | CON vs RSV+VPA: | >0.999 |
| | VPA: 79.41 (79.23±79.73) | | | | RSV vs VPA: | 0.002** |
| | RSV+VPA: 77.91 (77.74±77.93) | | | | RSV vs RSV+VPA: | >0.999 |
| | | | | | VPA vs RSV+VPA: | 0.208 |
| g | CON: 73.50 (59.00±83.50) RSV: 73.00 (61.00±76.00) VPA: 45.50 (43.00±47.75) RSV+VPA: 83.50 (77.25±107.00) | 15.659, 3 | 0.001** | CON: 9.50 RSV: 10.86 VPA: 20.50 RSV+VPA: 5.00 | CON vs RSV: | >0.999 |
| | | | | | CON vs VPA: | 0.030* |
| | | | | | CON vs RSV+VPA: | >0.999 |
| | | | | | RSV vs VPA: | 0.064# |
| | | | | | RSV vs RSV+VPA: | >0.999 |
| | | | | | VPA vs RSV+VPA: | 0.002** |
| %/g | CON: 1.052 (0.914±1.347) | 14.699, 3 | 0.002** | CON: 14.33 RSV: 12.79 VPA: 3.50 RSV+VPA: 19.88 | CON vs RSV: | >0.999 |
| | | | | | CON vs VPA: | 0.034* |
| | RSV: 1.025 (0.998±1.268) | | | | CON vs RSV+VPA: | >0.999 |
| | VPA: 1.749 (1.663±1.847) | | | | RSV vs VPA: | 0.082# |
| | RSV+VPA: 0.934 (0.737±1.009) | | | | RSV vs RSV+VPA: | 0,570 |
| | | | | | VPA vs RSV+VPA: | 0.001** |

%: brain water content percentage; **g:** body weight; **%/g:** proportion of brain fluid. p <0.05 was considered significant. *p<0.05, **p<0.01. Statistical analyses: Kruskal-Wallis. N_{CON} : 5, N_{RSV} : 6, N_{VPA} : 6, $N_{RSV+VPA}$: 4.

Table 2 – Descriptive statistics of Evans blue dye

| | Median (IQR) | Chi-Square, df | p Value | Mean rank | Pairwise compa | risons |
|------------------|--|-------------------|---------|---|---|---|
| СР | CON: 98.08 (75.07±103,4) RSV: 77.09 (56.18±91.46) VPA: 253.6 (150.4±393.0) RSV+VPA: 72.00 (62.16±88.70) | 11.674, 3 | 0.009** | CON: 12.60 RSV: 7.64 VPA: 19.50 RSV+VPA: 7.30 | CON vs RSV: CON vs VPA: CON vs RSV+VPA: RSV vs VPA: RSV vs RSV+VPA: VPA vs RSV+VPA: | >0.999 0.584 >0.999 0.014* >0.999 0.020* |
| HIP | CON: 100.6 (81.16±122.5) RSV: 83.16 (63.64±97.31) VPA: 107.4 (57.4±180.5) RSV+VPA: 68.5 (55.03±75.59) | 5.572, 3 | 0.134 | CON: 14.90 RSV: 10.36 VPA: 13.25 RSV+VPA: 6.20 | CON vs RSV: CON vs VPA: CON vs RSV+VPA: RSV vs VPA: RSV vs RSV+VPA: VPA vs RSV+VPA: | >0.999 >0.999 0.149 >0.999 >0.999 0.542 |
| AmR | CON: 97.14 (90.83±115.4) RSV: 79.56 (66.98±94.11) VPA: 108.7 (46.77±159.2) RSV+VPA: 66.17 (44.93±83.03) | 7.812, 3 | 0.050 | CON: 15.60 RSV: 8.86 VPA: 14.75 RSV+VPA: 6.40 | CON vs RSV: CON vs VPA: CON vs RSV+VPA: RSV vs VPA: RSV vs RSV+VPA: VPA vs RSV+VPA: | 0.381 >0.999 0.114 0.778 >0.999 0.269 |
| pSSA (II/III) | CON: 123.00 (113.7±146.1) RSV: 88.71 (78.22±106.4) VPA: 838.6 (624.3±1019.0) RSV+VPA: 86.19 (78.83±112.4) | 13.541, 3 | 0.004** | CON: 13.8 RSV: 6.79 VPA: 19.50 RSV+VPA: 7.30 | CON vs RSV: CON vs VPA: CON vs RSV+VPA: RSV vs VPA: RSV vs RSV+VPA: VPA vs RSV+VPA: | 0.321 >0.999 0.585 0.006** >0.999 0.020* |
| pSSA (IV/V) | CON: 124.6 (115.4±142.0) RSV: 105.9 (62.22±118.5) VPA: 483,9 (321.5±578.7) RSV+VPA: 95.09 (91.75±120.7) | 12.855, 3 | 0.005** | CON: 13.40 RSV: 7.00 VPA: 19.50 RSV+VPA: 7.40 | CON vs RSV: CON vs VPA: CON vs RSV+VPA: RSV vs VPA: RSV vs RSV+VPA: VPA vs RSV+VPA: | 0.468 0.856 0.757 0.008** >0.999 0.022* |
| aCC (II/III) | CON: 100.9 (78.63±112.2) RSV: 77.78 (74.32±90.65) VPA: 370.1 (279.2±416.7) RSV+VPA: 71.57 (68.05±95.45) | 11.670, 3 | 0.001** | CON: 11.50 RSV: 8.21 VPA: 18.50 RSV+VPA: 5.25 | CON vs RSV: CON vs VPA: CON vs RSV+VPA: RSV vs VPA: RSV vs RSV+VPA: VPA vs RSV+VPA: | >0.999 0.465 0.690 0.033* >0.999 0.009** |
| aCC (IV/V) | CON: 86.73 (81.27±99.31) RSV: 78.16 (77.02±107.5) VPA: 423.9 (335.2±447.6) RSV+VPA: 80.25 (78.98±92.59) | 10.801, 3 | 0.013* | CON: 11.20 RSV: 6.86 VPA: 18.50 RSV+VPA: 8.00 | CON vs RSV: CON vs VPA: CON vs RSV+VPA: RSV vs VPA: RSV vs RSV+VPA: VPA vs RSV+VPA: | >0.999 0.391 >0.999 0.010* >0.999 0.071# |
| PrL (II/III) | CON: 78.56 (71.57±100.8) RSV: 79.78 (74.39±106.5) VPA: 378.2 (230.0±422.7) RSV+VPA: 73.94 (64.44±89.42) | 9.926, 3 | 0.019* | CON: 8.40 RSV: 9.71 VPA: 18.50 RSV+VPA: 6.50 | CON vs RSV: CON vs VPA: CON vs RSV+VPA: RSV vs VPA: RSV vs RSV+VPA: VPA vs RSV+VPA: | >0.999 0.065# >0.999 0.106 >0.999 0.024* |
| PrL (IV/V) | CON: 82.80 (78.15±99.0) RSV: 86.23 (71.51±108.7) VPA: 364.9 (172.0±378.0) RSV+VPA: 69.02 (63.39±89.87) | 9.653, 3 | 0.022* | CON: 9.70 RSV: 9.71 VPA: 18.00 RSV+VPA: 5.38 | CON vs RSV: CON vs VPA: CON vs RSV+VPA: RSV vs VPA: RSV vs RSV+VPA: VPA vs RSV+VPA: | >0.999 0.219 >0.999 0.152 >0.999 0.015* |

| | | | | | CON vs RSV: | >0.999 |
|----------|------------------------------|-----------|--------|---------------|-----------------|--------|
| | CON: 81.6 (80.25±93.42) | | | CON: 9.40 | CON vs VPA: | 0.141 |
| IL | RSV: 79.34 (71.30±104.2) | 9.060, 3 | 0.028* | RSV: 7.93 | CON vs RSV+VPA: | >0.999 |
| (II/III) | VPA: 232.9 (141.5±351.4) | 9.000, 3 | 0.026 | VPA: 18.38 | RSV vs VPA: | 0.029* |
| | RSV+VPA: 78.77 (68.97±106.5) | | | RSV+VPA: 8.50 | RSV vs RSV+VPA: | >0.999 |
| | | | | | VPA vs RSV+VPA: | 0.109 |
| | | | | | CON vs RSV: | >0.999 |
| | CON: 87.79 (86.19±104.8) | | | CON: 10.80 | CON vs VPA: | 0.313 |
| IL | RSV: 81.65 (76.40±108.2) | 10.289, 3 | 0.016* | RSV: 7.21 | CON vs RSV+VPA: | >0.999 |
| (IV/V) | VPA: 275.0 (153.4±336.0) | 10.209, 3 | 0.010 | VPA: 18.50 | RSV vs VPA: | 0.014* |
| | RSV+VPA: 87.05 (66.13±98.72) | | | RSV+VPA: 7.88 | RSV vs RSV+VPA: | >0.999 |
| | | | | | VPA vs RSV+VPA: | 0.066# |

II/III: upper cortical layers; **IV/V**: deeper cortical layers; **aCC**: anterior cingulate cortex; **AmR**: amygdala region; **CP**: choroid plexus; **HIP**: hippocampus; **IL**: infralimbic cortex; **IQR**: interquartile range; **PrL**: prelimbic cortex; **pSSA**: primary somatosensory area. p <0.05 was considered significant. *p<0.05, **p<0.01. Statistical analyses: Kruskal-Wallis. Ncon: 5, Nrsv: 7, Nvpa: 4, Nrsv+vpa: 4.

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Table 3 – Descriptive statistics of the AQP1 and AQP4 distribution profile in pSSA

| | pasa | | | |
|----------|----------------------|--|-----------------|----------|
| | Mean ± SD | F (DFn, DFd); p Value | Pairwise compa | risons |
| | | | CON vs RSV: | >0.999 |
| AQP1 | CON: 438.3±105.9 | Internation: F (4, 47) 0.042; n. 0.000 | CON vs VPA: | 0.934 |
| AQP1 | RSV: 455.8±121.3 | Interaction: $F(1, 17) = 0.013$; $p = 0.909$ | CON vs RSV+VPA: | 0.966 |
| (11/111) | VPA: 373.7±89.17 | VPA: F (1, 17) = 2,171; p = 0.158 RSV: F (1, 17) = 0,064; p = 0.804 | RSV vs VPA: | 0.744 |
| (11/111) | RSV+VPA: 380.2±111.2 | K3V. F(1, 17) = 0,004, p = 0.004 | RSV vs RSV+VPA: | 0.838 |
| | | | VPA vs RSV+VPA: | >0.999 |
| | | | CON vs RSV: | 0.969 |
| | CON: 556.3±140.8 | Interaction: F (1, 17) = 0.821; p = 0.377 | CON vs VPA: | 0.015* |
| AQP1 | RSV: 507.6±82.91 | VPA: F (1, 17) = 18.28; p = 0.0005*** | CON vs RSV+VPA: | 0.049* |
| (IV/V) | VPA: 337.6±80.89 | RSV: F (1, 17) = 0.0607; p = 0.808 | RSV vs VPA: | 0.039* |
| | RSV+VPA: 365.4±83.67 | | RSV vs RSV+VPA: | 0.139 |
| | 7 | | VPA vs RSV+VPA: | 0.997 |
| | | | CON vs RSV: | 0.801 |
| AQP4 | CON: 154.1±32.47 | Interaction: F (1, 18) = 0.6649; p = 0.4255 | CON vs VPA: | 0.799 |
| AGF | RSV: 194.3±55.26 | VPA: F (1, 18) = 0.8502; p = 0.3687 | CON vs RSV+VPA: | 0.789 |
| (11/111) | VPA: 194.4±52.50 | RSV: F (1, 18) = 0.8440; p = 0.3704 | RSV vs VPA: | >0.999 |
| (11/111) | RSV+VPA: 196.8±69.81 | (1, 10) = 0.0440, p = 0.0704 | RSV vs RSV+VPA: | >0.999 |
| | 100 | | VPA vs RSV+VPA: | >0.999 |
| | | | CON vs RSV: | 0.2348 |
| AQP4 | CON: 109.9±23.35 | Interaction: F (1, 19) = 14.75; p = 0.0011** | CON vs VPA: | 0.0052** |
| AQF4 | RSV: 147.5±21.48 | VPA: F (1, 19) = 2.573; p = 0.1252 | CON vs RSV+VPA: | 0.9966 |
| (IV/V) | VPA: 173.8±27.00 | RSV: F (1, 19) = 2.373, p = 0.1232 | RSV vs VPA: | 0.4338 |
| (14/4) | RSV+VPA: 123.7±28.31 | 1.0 v. 1 (1, 13) = 0.7 007, p = 0.4017 | RSV vs RSV+VPA: | 0.5920 |
| | | | VPA vs RSV+VPA: | 0.0252* |

II/III: upper cortical layers; **IV/V**: deeper cortical layers; **pSSA**: primary somatosensory area; **SD**: standard deviation. p <0.05 was considered significant. *p<0.05, **p<0.01, ***p<0.001. Statistical analyses: two-way ANOVA parametric test followed by Sidak. Ncon: 5, Nrsv: 6, Nvpa: 6, Nrsv+vpa: 5. "Mean" represents de mean of fluorescence.

Table 4 - Descriptive statistics of the AQP1 and AQP4 distribution profile in mPFC

| AQP1 CON: 440.8±84.40 RSV: 479A: 249.5±114.8 AQP1 CON: 444.4±79.13 RSV: 87V-VPA: 249.5±140.8 AQP1 CON: 444.4±79.13 RSV: 883.9±66.86 Pt. VPA: 226.2±52.99 RSV: VVPA: 238.9±12.0 AQP1 CON: 443.4±89.35 Interaction: F (1, 14) = 0.038; p = 0.869 Pt. VPA: 226.2±52.99 RSV: WVPA: 226.2±52.99 RSV: WVPA: 238.9±12.0 AQP1 CON: 443.4±89.3 RSV: 883.9±6.86 Pt. VPA: 226.2±52.99 RSV: WVPA: 238.9±12.0 AQP1 CON: 438.1±84.61 RSV: 865.1±95.19 RSV: VVPA: 238.9±12.0 AQP1 CON: 438.1±84.61 RSV: 418.4±89.54 IL VPA: 226.2±52.99 RSV: VVPA: 238.9±12.0 AQP1 CON: 438.1±84.61 RSV: 418.4±89.54 IVPA: 226.2±52.99 RSV: 400.5±94.07 RSV: F (1, 14) = 0.026; p = 0.869 VPA: F (1, 14) = 0.028; p = 0.869 VPA: F (1, 14) = 0.028; p = 0.869 VPA: F (1, 14) = 0.0091*** RSV: WS VPA: 0.099 VPA: F (1, 14) = 0.0091*** RSV: WS VPA: 0.099 VPA: F (1, 14) = 0.0091*** RSV: WS VPA: 0.099 VPA: F (1, 14) = 0.0091*** RSV: WS VPA: 0.0001*** RSV: WS VPA: 0.0000*** RSV: WS RSV: WPA: 0.0000*** RSV: WS VPA: | | Mean ± SD | F (DFn, DFd); p Value | Pairwise compa | risons |
|--|----------|--|---|-----------------|-----------|
| AQP1 CON: 440.9±8.94.0 (I/I/III) RSV+VPA: 296.9±10.8 (I/I/III) RSV+VPA: 298.9±10.8 (I/I/III) RSV+VPA: 298.9±10.8 (I/I/III) RSV+VPA: 298.9±10.8 (I/I/III) RSV+VPA: 298.9±10.8 (I/I/III) RSV+VPA: 299.9±138.2 (I/I/III) RSV+VPA: 298.9±10.8 (I/I/III) RSV+VPA: 299.9±138.2 (I/I/III) RSV+VPA: 299.9±138.2 (I/I/IIII) RSV+VPA: 299.9±138.2 (I/I/IIII) RSV+VPA: 298.9±10.9 (I/I/IIIII) RSV+VPA: 298.9±10.9 (I/I/IIII) RSV+VPA: 299.9±138.2 (I/I/IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII | | | , | | |
| RSV: 317,6459.90 VPA: F(1, 14) = 11.59; p = 0.0433* RSV + VPA: 240.3±89.26 RSV+VPA: 240.3±89.26 RSV+VPA: 240.3±89.26 RSV: F(1, 14) = 0.136 ; p = 0.717 RSV: 384.2±88.89 CON: 473.4±89.15 RSV: 874.29 = 0.010** RSV+VPA: 256.6±107.4 RSV: F(1, 14) = 0.0311; p = 0.863 RSV: 874.29 = 0.010** RSV+VPA: 256.6±107.4 RSV: F(1, 14) = 0.0311; p = 0.863 RSV: 874.26 = 0.010** RSV+VPA: 256.6±107.4 RSV: F(1, 14) = 0.0311; p = 0.863 RSV: 874.26 = 0.010** RSV-VPA: 256.6±107.4 RSV: F(1, 14) = 0.0311; p = 0.863 RSV: 874.26 = 0.010** RSV-VPA: 256.7±39.04 RSV: F(1, 14) = 0.031; p = 0.010** RSV-VPA: 256.7±39.04 RSV: F(1, 14) = 0.039; p = 0.845 RSV: 878.4VPA: 0.020* RSV-VPA: 20.3±110.8 RSV: 383.9±66.96 RSV: 874.249.5±114.8 RSV: 878.249.5±114.8 RSV: 878.249.5±114 | AQP1 | CON: 340 0+20 61 | | | |
| AQP1 CON: 440.8±84.40 RSV: F(1, 14) = 0.136 ; p = 0.717 RSV vs VPA: 0.264 | ,,,,,, | | | | |
| RSV+VPA: 240.3±89.26 RSV: F (1, 14) = 0.136 ; p = 0.717 RSV vs RSV+VPA: 0.301 VPA vs RSV+VPA: 0.999 CON vs RSV: 0.079 CON vs RSV: 0.995 CON vs RSV: 0.99 | aCC | | | | |
| AQP1 CON: 413.4±89.15 Interaction: F (1, 14) = 0.839; p = 0.375 CON vs RSV: 0.995 | | | RSV: $F(1, 14) = 0.136$; $p = 0.717$ | | |
| AQP1 CON: 443.4±89.15 | , , | | | | |
| AQP1 CON: 413.4±89.15 Interaction: F (1, 14) = 0.839; p = 0.375 CON vs RSV+VPA: 0.079# RSV + VPA: 213.5±60.90 RSV + VPA: 213.5±60.90 RSV + VPA: 213.5±60.90 RSV + VPA: 256.6±107.4 RSV + VPA: 256.7±39.04 RSV + VPA: 256.7±39.94 RSV + VPA: 256.9±14.8 RSV + VPA: 256.9±14.25 RSV + VPA: 256.9±19.19 RSV | | | | | |
| RSV: 384 2-68.89 | AQP1 | CON: 413.4+89.15 | | | |
| CON: 440.8±84.40 RSV: 400.3±110.8 RSV: F(1, 14) = 0.0311; p = 0.863 RSV vs VPA: 0.0207 VPA vs RSV+VPA: 0.997 VPA vs RSV+VPA: 0.997 VPA vs RSV+VPA: 0.998 CON vs RSV: 400.3±110.8 VPA: F(1, 14) = 0.0512; p = 0.486 CON vs RSV: 0.0985 CON vs RSV: 0.0985 VPA: F(1, 14) = 17.01; p = 0.0010** RSV: 400.3±110.8 VPA: F(1, 14) = 17.01; p = 0.0010** RSV: 0.050* VPA: RSV+VPA: 0.099 VPA: F(1, 14) = 3.034; p < 0.000*** VPA: RSV+VPA: 0.013* CON vs RSV: 0.013* CON vs RSV+VPA: 0.0007** VPA: RSV+VPA: 0.0007** VPA: RSV+VPA: 0.0000*** VPA: RSV+VPA: 0.0000*** VPA: RSV+VPA: 0.0000*** VPA: RSV+VPA: 0.0000** VPA: RSV+VPA: 0.0000*** VPA: RSV+VPA: 0.0 | | | | | |
| RSV+VPA: 256.6±107.4 RSV+VPA: 256.6±107.4 RSV: F (1, 14) = 0.031; p = 0.063 RSV ss RSV+VPA: 0.207 VPA vs RSV+VPA: 0.207 VPA vs RSV+VPA: 0.998 CON vs RSV: 0.985 CON vs RSV: 0.985 CON vs RSV: 0.985 CON vs RSV: 0.0985 CON vs RSV: 0.056# RSV vs PA: 0.068 RSV vs PA: 0.099 CON vs RSV: 0.099 VPA: F (1, 14) = 0.039; p = 0.845 RSV vs RSV+VPA: 0.999 VPA: F (1, 14) = 3.04; p = 0.869 VPA: Ss RSV+VPA: 0.999 VPA: F (1, 14) = 3.04; p = 0.869 VPA: Ss RSV+VPA: 0.099 VPA: F (1, 14) = 3.04; p = 0.0001*** RSV vs PA: 0.0030*** RSV vs RSV | aCC | VPA: 213.5±60.90 | | | |
| AQP1 CON: 440.8±84.40 | (IV/V) | RSV+VPA: 256.6±107.4 | RSV: F (1, 14) = 0.0311; p = 0.863 | | |
| AQP1 CON: 440,8±84.40 Interaction: F (1, 14) = 0.512; p = 0.486 VPA: 90.3±110.8 VPA: 226.7±39.04 RSV: 400.3±110.8 VPA: 226.7±39.04 RSV: F (1, 14) = 17.01; p = 0.0010** RSV vs VPA: 0.056# RSV vs VPA: 0.086# VPA: 249.5±114.8 RSV: F (1, 14) = 0.039; p = 0.845 RSV vs RSV+VPA: 0.168 VPA: 8SV+VPA: 0.168 VPA: 8SV+VPA: 0.168 VPA: 8SV+VPA: 0.168 VPA: 8SV+VPA: 0.099 VPA: F (1, 14) = 3.832; p = 0.0705 RSV: vs RSV+VPA: 0.003** VPA: 8SV+VPA: 0.0007** RSV: 383.0±66.96 VPA: F (1, 14) = 3.832; p = 0.0705 RSV: vs RSV+VPA: 0.0013* CON vs RSV: 0.739 CON vs RSV: 0.739 CON vs RSV: 0.739 CON vs RSV: 0.739 CON vs RSV: 0.0007** RSV: vs RSV+VPA: 0.0086* VPA: F (1, 14) = 1.550; p = 0.2336 VPA: F (1, 14) = 1.550; p = 0.2336 CON vs RSV: 0.780 CON vs RSV: 0.780 CON vs RSV: 0.780 CON vs RSV: 0.780 CON vs RSV: 0.0116* RSV: vs RSV+VPA: 0.294 VPA: F (1, 14) = 0.2064; p = 0.6565 RSV: vs RSV+VPA: 0.0996 CON vs RSV: 0.982 CON vs RSV+VPA: 0.294 VPA: RSV+VPA: 0.0326* CON vs RSV+VPA: 0.0326* CON vs RSV+VPA: 0.0326* CON vs RSV+VPA: 0.0326* CON vs RSV+VPA: 0.0996 CON vs RSV+VPA: 0.0326* CON vs RSV+VPA: 0.0996 CON vs RSV+V | ` ' | | | VPA vs RSV+VPA: | 0.979 |
| AQP1 CON: 440,8±84.40 Interaction: F (1, 14) = 0.512; p = 0.486 VPA: 90.3±110.8 VPA: 226.7±39.04 RSV: 400.3±110.8 VPA: 226.7±39.04 RSV: F (1, 14) = 17.01; p = 0.0010** RSV vs VPA: 0.056# RSV vs VPA: 0.086# VPA: 249.5±114.8 RSV: F (1, 14) = 0.039; p = 0.845 RSV vs RSV+VPA: 0.168 VPA: 8SV+VPA: 0.168 VPA: 8SV+VPA: 0.168 VPA: 8SV+VPA: 0.168 VPA: 8SV+VPA: 0.099 VPA: F (1, 14) = 3.832; p = 0.0705 RSV: vs RSV+VPA: 0.003** VPA: 8SV+VPA: 0.0007** RSV: 383.0±66.96 VPA: F (1, 14) = 3.832; p = 0.0705 RSV: vs RSV+VPA: 0.0013* CON vs RSV: 0.739 CON vs RSV: 0.739 CON vs RSV: 0.739 CON vs RSV: 0.739 CON vs RSV: 0.0007** RSV: vs RSV+VPA: 0.0086* VPA: F (1, 14) = 1.550; p = 0.2336 VPA: F (1, 14) = 1.550; p = 0.2336 CON vs RSV: 0.780 CON vs RSV: 0.780 CON vs RSV: 0.780 CON vs RSV: 0.780 CON vs RSV: 0.0116* RSV: vs RSV+VPA: 0.294 VPA: F (1, 14) = 0.2064; p = 0.6565 RSV: vs RSV+VPA: 0.0996 CON vs RSV: 0.982 CON vs RSV+VPA: 0.294 VPA: RSV+VPA: 0.0326* CON vs RSV+VPA: 0.0326* CON vs RSV+VPA: 0.0326* CON vs RSV+VPA: 0.0326* CON vs RSV+VPA: 0.0996 CON vs RSV+VPA: 0.0326* CON vs RSV+VPA: 0.0996 CON vs RSV+V | | | | | |
| RSV: 400.3±110.8 | AQP1 | CON: 440.8±84.40 | | | |
| PrL VPA: 226.7±39.04 VPA: 14.9 = 17.01; p = 0.0010* RSV vs VPA: 0.086# RSV vs RSV+VPA: 0.168 RSV vs RSV+VPA: 0.168 VPA: 28.0±68.79 RSV: F (1, 14) = 0.039; p = 0.845 RSV vs RSV+VPA: 0.168 VPA: RSV+VPA: 0.099 CON vs RSV: 0.739 CON vs RSV: 0.0146 RSV: SESV+VPA: 0.0146 RSV: SESV+VPA: 0.0146 RSV: SESV+VPA: 0.0000*** RSV: SESV+VPA: 0.0116 RSV: S | | | | CON vs RSV+VPA: | |
| AQP1 CON: 438.1±84.61 RSV+VPA: 249.9±1148.2 Interaction: F (1, 14) = 0.028; p = 0.869 VPA: S RSV+VPA: 0.0037** AQP1 CON: 434.4±79.13 RSV: 383.0±66.96 VPA: F (1, 14) = 3.832; p = 0.0705 RSV: VPA: 0.0007*** RSV+VPA: 189.3±85.19 RSV: F (1, 14) = 3.832; p = 0.0705 RSV+VPA: 0.0007*** RSV vs VPA: 0.146 RSV vs VPA: 0.146 RSV vs RSV+VPA: 0.0080** VPA: S RSV+VPA: 0.0080** VPA: F (1, 14) = 1.550; p = 0.2336 CON vs RSV: 0.780 CON vs RSV: 0.780 CON vs RSV+VPA: 0.00324* RSV: F (1, 14) = 0.2064; p = 0.6565 RSV vs VPA: 0.111 RSV vs VPA: 0.00324* RSV: F (1, 14) = 0.2064; p = 0.6565 RSV vs RSV+VPA: 0.0986 CON vs RSV+VPA: 0.0996 CON vs RSV+VPA: 0.0996 CON vs RSV+VPA: 0.0936* RSV: F (1, 14) = 0.2064; p = 0.0004*** RSV vs VPA: 0.111 RSV vs VPA: 0.00324* RSV: F (1, 14) = 0.459; p = 0.0004*** RSV vs VPA: 0.0132* RSV: F (1, 14) = 0.459; p = 0.0044** RSV: S VPA: 0.0132* RSV vs VPA: 0.00326* CON vs RSV+VPA: 0.0986 CON vs RSV+VPA: 0.0936* CON vs RSV+VPA: 0.0986 CON vs RSV+VPA: 0.00326* RSV: S VPA: 0.0132* RSV: S VPA: 0.0132* RSV: S VPA: 0.0132* RSV: S VPA: 0.0066* RSV vs VPA: 0.0066* RSV vs RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.0066* RSV vs RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.00326* RSV: S RSV+VPA: 0.0066* RSV vs RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.0066* RSV vs RSV+VPA: 0.00324* RSV vs RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.0066* RSV vs RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.0066* RSV vs RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.0066* RSV vs RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.00324* RSV: S RSV+VPA: 0.0066* RSV vs RSV+VPA: 0 | PrL | | | RSV vs VPA: | |
| AQP1 CON: 444.4±79.13 RSV: 383.0±66.96 VPA: F (1, 14) = 0.028; p = 0.869 VPA: F (0, 14) = 3.0.04; p < 0.0001**** RSV: VPA: 262.1±52.99 VPA: F (1, 14) = 3.0.04; p < 0.0001**** RSV: VPA: 0.013** RSV: VPA: 0.0000*** RSV: 0.780 VPA: 0.0000*** RSV: 0.780 VPA: 0.0866 VPA: F (1, 14) = 1.550; p = 0.2336 VPA: RSV: VPA: 0.0866 VPA: F (1, 14) = 1.550; p = 0.2336 VPA: D.0866 VPA: F (1, 14) = 1.550; p = 0.2336 VPA: D.0866 VPA: F (1, 14) = 1.550; p = 0.2366 VPA: F (1, 14) = 1.550; p = 0.2366 VPA: F (1, 14) = 0.0009*** RSV: VPA: RSV+VPA: 0.0324* RSV: VPA: D.0324* RSV: VPA: D.0324* RSV: F (1, 14) = 0.2064; p = 0.6565 VPA: SRSV+VPA: 0.0324* RSV: VPA: RSV+VPA: 0.0324* RSV: VPA: D.0324* R | (II/III) | RSV+VPA: 249.5±114.8 | RSV. F $(1, 14) = 0.039$, p = 0.845 | RSV vs RSV+VPA: | 0.168 |
| AQP1 CON: 444.4±79.13 | | | | VPA vs RSV+VPA: | 0.999 |
| RSV: 383.0±66.96 | - | | | CON vs RSV: | 0.739 |
| PrL (IV/V) RSV+VPA: 189.3±85.19 | AQP1 | CON: 444.4±79.13 | L. ((| CON vs VPA: | 0.013* |
| Con | | RSV: 383.0±66.96 | | CON vs RSV+VPA: | 0.0007*** |
| AQP1 CON: 458.1±84.61 | PrL | VPA: 262.1±52.99 | | RSV vs VPA: | 0.146 |
| AQP1 CON: 438.1±84.61 RSV: 365.1±95.19 VPA: 205.0±41.25 (IIIIII) RSV+VPA: 238.9±122.0 AQP1 CON: 459.4±80.84 RSV: 418.4±89.54 IL VPA: 247.2±30.44 (IV/V) RSV+VPA: 229.1±138.2 AQP4 CON: 522.8±86.07 RSV: 400.5±94.07 aCC VPA: 298.3±119.1 (II/III) RSV+VPA: 433.3±213.3 AQP4 CON: 555.4±160.3 RSV: F(1, 16) = 0.01204; p = 0.0125* VPA: 247.0±30.84 RSV: F(1, 16) = 0.01204; p = 0.0125* VPA: 247.0±30.84 RSV: F(1, 16) = 0.01204; p = 0.0125* VPA: F(1, 16) = 0.6246; p = 0.0449* VPA vs RSV+VPA: 0.098 CON vs RSV: 0.780 CON vs RSV+VPA: 0.294 VPA vs RSV+VPA: 0.294 VPA vs RSV+VPA: 0.294 VPA vs RSV+VPA: 0.996 CON vs RSV: 0.982 CON vs RSV: 0.068# RSV vs VPA: 0.0036* CON vs RSV: 0.0036* CON vs RSV+VPA: 0.0048* RSV vs VPA: 0.068# RSV vs RSV+VPA: 0.068# RSV vs RSV+VPA: 0.064# CON vs RSV: 0.064# CON vs RSV: 0.064# CON vs RSV: 0.064# CON vs RSV: 0.068# RSV vs RSV+VPA: 0.099 VPA vs RSV+VPA: 0.0769 RSV vs RSV | (IV/V) | RSV+VPA: 189.3±85.19 | | RSV vs RSV+VPA: | 0.0080** |
| AQP1 CON: 438.1±84.61 | 13 | | | VPA vs RSV+VPA: | 0.686 |
| IL VPA: 205.0±41.25 VPA: F (1, 14) = 1.550; p = 0.2336 VPA: F (1, 14) = 17.51; p = 0.0009*** RSV: VPA: 203.0±41.25 RSV: F (1, 14) = 0.2064; p = 0.6565 RSV vs VPA: 0.111 RSV vs RSV+VPA: 0.294 VPA vs RSV+VPA: 0.996 VPA vs RSV+VPA: 0.996 VPA vs RSV+VPA: 0.998 CON vs RSV: 0.982 CON vs RSV: 0.982 CON vs RSV: 0.982 CON vs RSV: 0.0032* RSV: 418.4±89.54 Interaction: F (1, 14) = 0.069; p = 0.796 VPA: F (1, 14) = 0.459; p = 0.0004*** RSV: vs VPA: 0.0489* RSV: vs VPA: 0.0489* RSV: vs VPA: 0.0489* RSV: vs VPA: 0.0489* RSV: vs RSV+VPA: 0.0489* RSV: vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.064# RSV: 400.5±94.07 VPA: F (1, 16) = 2.709; p = 0.1193 RSV: F (1, 16) = 0.01204; p = 0.9140 RSV vs VPA: 0.099 VPA vs RSV+VPA: 0.880 RSV vs VPA: 0.099 VPA vs RSV+VPA: 0.689 CON vs RSV: 0.093 RSV: SSV+VPA: 0.099 VPA vs RSV+VPA: 0.099 VPA vs RSV+VPA: 0.099 VPA vs RSV+VPA: 0.099 VPA vs RSV+VPA: 0.769 RSV vs VPA: 0.717 CON vs RSV: 0.093 VPA: F (1, 16) = 9.964; p = 0.0061** RSV vs VPA: 0.134 RSV vs VPA: 0.134 RSV vs VPA: 0.134 RSV vs RSV+VPA: 0.987 RSV vs VPA: 0.134 RSV vs RSV+VPA: 0.987 RSV vs VPA: 0.134 RSV vs RSV+VPA: 0.987 RSV vs RSV+VPA: 0.987 RSV vs RSV+VPA: 0.987 RSV vs RSV+VPA: 0.987 RSV vs RSV+VPA: 0.134 RSV vs RSV+VPA: 0.134 RSV vs RSV+VPA: 0.987 RSV v | | | | CON vs RSV: | 0.780 |
| IL VPA: 205.041.25 RSV: F (1, 14) = 17.51; p = 0.0009*** RSV vs VPA: 0.111 RSV vs RSV+VPA: 0.294 VPA vs RSV+VPA: 0.294 VPA vs RSV+VPA: 0.294 VPA vs RSV+VPA: 0.996 VPA vs RSV+VPA: 0.0438* VPA vs RSV+VPA: 0.0132* VPA vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.064# RSV: 400.5±94.07 RSV: 400.5±94.07 RSV: 400.5±94.07 RSV: F (1, 16) = 2.709; p = 0.1193 RSV: F (1, 16) = 0.01204; p = 0.9140 RSV vs VPA: 0.999 VPA vs RSV+VPA: 0.064# RSV: S4.4±76.63 RSV: 354.4±76.63 RSV: 354.4±76.63 RSV: 42.98 RSV: F (1, 16) = 0.6246; p = 0.4409 RSV vs VPA: 0.722 CON vs RSV: 0.717 CON vs RSV+VPA: 0.722 CON vs RSV+VPA: 0.722 CON vs RSV+VPA: 0.722 CON vs RSV+VPA: 0.722 CON vs RSV+VPA: 0.064** RSV: S4.4±6±144.4 RSV: F (1, 16) = 9.964; p = 0.0061** RSV vs VPA: 0.0064** RSV vs VPA: 0.0337 RSV: F (1, 16) = 0.1171; p = 0.0367 RSV vs VPA: 0.0337 RSV vs VPA: 0.0337 RSV: SV-VPA: 0.134 RSV vs VPA: 0.0337 RSV: SV-VPA: 0.134 RSV vs VPA: 0.0337 RSV: SV-VPA: 0.0337 RSV: SV-VPA: 0.1717 CON vs VPA: 0.0064** RSV: SV-VPA: 0.1717 RSV vs VPA: 0.0337 RSV: SV-VPA: 0.0337 | AQP1 | CON: 438.1±84.61 | L. ((E /4 . 4.4) 4 . E F O | CON vs VPA: | 0.0108* |
| RSV + VPA: 238.9±122.0 RSV: F (1, 14) = 0.2064; p = 0.6565 RSV vs RSV+VPA: 0.294 VPA vs RSV+VPA: 0.996 VPA vs RSV+VPA: 0.996 CON vs RSV: 0.982 CON vs RSV: 0.982 CON vs RSV: 0.982 CON vs RSV: 0.982 CON vs PA: 0.0236* CON vs PA: 0.0236* RSV vs PA: 0.0236* CON vs PA: 0.0236* CON vs PA: 0.0236* CON vs PA: 0.0236* RSV vs PA: 0.0489* RSV vs PA: 0.0489* RSV vs PA: 0.0489* RSV vs RSV+VPA: 0.0489* RSV vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.064# CON vs RSV: 0.064# CON vs RSV: 0.064# CON vs RSV+VPA: 0.0866 CON vs RSV: 0.064# RSV vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.064# CON vs RSV: 0.064# CON vs RSV: 0.064# CON vs RSV+VPA: 0.880 RSV vs VPA: 0.880 RSV vs RSV+VPA: 0.999 VPA vs RSV+VPA: 0.999 VPA vs RSV+VPA: 0.099 VPA vs RSV+VPA: 0.0058** CON vs RSV: 0.093 CON vs RSV: 0.093 CON vs RSV+VPA: 0.769 RSV vs RSV+VPA: 0.769 RSV vs RSV+VPA: 0.769 RSV vs RSV+VPA: 0.722 CON vs RSV: 0.717 CON vs RSV+VPA: 0.722 CON vs RSV+VPA: 0.722 CON vs RSV+VPA: 0.134 RSV vs RSV+VPA: 0 | | RSV: 365.1±95.19 | | CON vs RSV+VPA: | 0.0324* |
| RSV+VPA: 236.9±122.0 RSV vs RSV+VPA: 0.294 VPA vs RSV+VPA: 0.294 VPA vs RSV+VPA: 0.996 CON vs RSV: 0.982 CON vs RSV: 0.0236* CON vs RSV+VPA: 0.0236* CON vs RSV+VPA: 0.0132* RSV vs RSV+VPA: 0.0132* RSV vs RSV+VPA: 0.0489* RSV vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.066# RSV vs RSV+VPA: 0.064# CON vs RSV: 0.586 CON vs RSV: 0.586 CON vs RSV: 0.064# CON vs RSV: 0.064# RSV vs VPA: 0.999 VPA vs RSV+VPA: 0.0589 CON vs RSV: 0.093 CON vs RSV: 0.068** CON vs RSV+VPA: 0.769 RSV vs VPA: 0.717 RSV vs VPA: 0.717 CON vs RSV+VPA: 0.722 CON vs RSV: 0.0717 CON vs RSV+VPA: 0.337 RSV: SV PA: 0.132 CON vs RSV+VPA: 0.337 RSV vs RSV+VPA: 0.171 CON vs RSV+VPA: 0.172 CON vs RSV+VPA: 0.337 RSV vs RSV+VPA: 0.174 RSV vs RSV+VPA: 0.174 CON vs RSV+VPA: 0 | IL | VPA: 205.0±41.25 | | RSV vs VPA: | 0.111 |
| AQP1 CON: 459.4±80.84 RSV: 418.4±89.54 IL VPA: 247.2±30.44 (IV/V) RSV+VPA: 229.1±138.2 AQP4 CON: 552.8±86.07 RSV: 400.5±94.07 VPA: F (1, 14) = 0.069; p = 0.004** RSV+VPA: 433.3±213.3 AQP4 CON: 555.4±160.3 RSV: F (1, 16) = 0.01204; p = 0.0125* QPA: 254.0±42.98 Interaction: F (1, 16) = 7.910; p = 0.0125* VPA: 254.0±4.098 (IV/V) RSV+VPA: 366.8±167.8 AQP4 CON: 428.0±68.79 RSV: 354.4±90.40 VPA: 213.2±45.47 (II/III) RSV+VPA: 314.6±144.4 AQP4 CON: 428.0±68.79 RSV: 354.1±90.40 VPA: 213.2±45.47 (II/III) RSV+VPA: 314.6±144.4 AQP4 CON: 428.0±68.79 RSV: 354.1±90.40 VPA: 213.2±45.47 (II/III) RSV+VPA: 314.6±144.4 | (II/III) | RSV+VPA: 238.9±122.0 | K3V. F(1, 14) = 0.2004, p = 0.0303 | RSV vs RSV+VPA: | 0.294 |
| CON: 459.4±80.84 RSV: 418.4±89.54 VPA: 247.2±30.44 RSV: 418.4±89.54 VPA: F (1, 14) = 0.069; p = 0.796 VPA: F (1, 14) = 21.20; p = 0.0004*** RSV + VPA: 229.1±138.2 RSV: F (1, 14) = 0.459; p = 0.5089 RSV vs RSV+VPA: 0.086# RSV vs RSV+VPA: 0.0489* RSV: F (1, 14) = 0.459; p = 0.5089 RSV vs RSV+VPA: 0.0489* RSV vs RSV+VPA: 0.0999 RSV: 400.5±94.07 VPA: F (1, 16) = 2.709; p = 0.1193 RSV: F (1, 16) = 2.709; p = 0.1193 RSV: F (1, 16) = 0.01204; p = 0.9140 RSV vs RSV+VPA: 0.880 RSV vs RSV+VPA: 0.999 RSV vs RSV+VPA: 0.999 RSV: 354.4±76.63 RSV: 354.4±76.63 RSV: 354.4±76.63 RSV: F (1, 16) = 6.711; p = 0.0125* VPA: F (1, 16) = 6.711; p = 0.0125* CON vs RSV: 0.093 RSV: SRSV+VPA: 0.099 RSV: F (1, 16) = 0.6246; p = 0.4409 RSV vs RSV+VPA: 0.999 RSV vs RSV+VPA: 0.099 RSV: SRSV+VPA: 0.0722 RSV vs RSV+VPA: 0.099 RSV: SRSV+VPA: 0.099 RSV vs RSV+VPA: 0.099 RSV: SRSV+VPA: 0.0058** RSV: F (1, 16) = 0.6246; p = 0.4409 RSV vs RSV+VPA: 0.099 RSV vs RSV+VPA: 0.0064** RSV: F (1, 16) = 9.964; p = 0.0061** RSV vs RSV+VPA: 0.0337 RSV vs RSV+VPA: 0.337 RSV vs RSV+VPA: 0.997 RSV vs RSV+VPA: | | | | VPA vs RSV+VPA: | 0.996 |
| RSV: 418.4±89.54 VPA: 247.2±30.44 VPA: 247.2±30.44 RSV: F (1, 14) = 21.20; p = 0.0004*** RSV vs VPA: 0.0132* RSV vs VPA: 229.1±138.2 RSV: F (1, 14) = 0.459; p = 0.5089 RSV vs RSV+VPA: 0.086# RSV vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.064# RSV: 400.5±94.07 RSV: 400.5±94.07 RSV: 400.5±94.07 RSV+VPA: 433.3±213.3 RSV: F (1, 16) = 2.709; p = 0.0121* VPA: F (1, 16) = 2.709; p = 0.0121* RSV vs RSV+VPA: 0.880 RSV vs VPA: 0.787 RSV vs RSV+VPA: 0.589 RSV: 354.4±76.63 RSV: 354.4±76.63 RSV: 354.4±76.63 RSV: F (1, 16) = 6.711; p = 0.0197* RSV: F (1, 16) = 0.6246; p = 0.4409 RSV vs RSV+VPA: 0.722 RSV vs RSV+VPA: 0.722 RSV: 354.1±90.40 RSV: 354.1±90.40 RSV: 354.1±90.40 RSV: F (1, 16) = 9.964; p = 0.0061** RSV vs RSV+VPA: 0.337 RSV: F (1, 16) = 9.964; p = 0.0061** RSV vs RSV+VPA: 0.337 RSV: SRSV+VPA: 0.337 RSV: F (1, 16) = 9.964; p = 0.0061** RSV vs RSV+VPA: 0.344 RSV vs RSV+VPA: 0.345 RS | | 1) | | CON vs RSV: | 0.982 |
| IL VPA: 247.2±30.44 RSV: F (1, 14) = 21.20; p = 0.0004*** RSV vs VPA: 0.086# RSV vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.0489* VPA vs RSV+VPA: 0.0999 CON vs RSV: 0.586 CON vs RSV: 0.064# CON vs RSV: 0.064# CON vs RSV+VPA: 0.064# RSV vs RSV+VPA: 0.064# CON vs RSV+VPA: 0.064# CON vs RSV+VPA: 0.064# CON vs RSV+VPA: 0.080 RSV vs RSV+VPA: 0.080 RSV vs RSV+VPA: 0.0999 VPA vs RSV+VPA: 0.0999 VPA vs RSV+VPA: 0.0999 VPA vs RSV+VPA: 0.0058** CON vs RSV: 0.093 CON vs RSV: 0.093 CON vs RSV: 0.093 CON vs RSV: 0.0058** CON vs RSV: 0.0058** CON vs RSV: 0.0058** CON vs RSV: 0.0058** CON vs RSV: 0.0769 RSV vs VPA: 0.0769 RSV vs RSV+VPA: 0.0722 CON vs RSV: 0.0722 CON vs RSV: 0.0722 CON vs RSV: 0.0064** CON vs RSV: 0.0064** CON vs RSV: 0.0064** RSV: 364.1±90.40 VPA: F (1, 16) = 9.964; p = 0.0061** RSV vs VPA: 0.0064** RSV vs VPA: 0.0064** RSV: F (1, 16) = 0.1171; p = 0.7367 RSV vs VPA: 0.0337 RSV vs VPA: 0.0344 RSV vs RSV+VPA: 0.0347 RSV vs RSV+V | AQP1 | CON: 459.4±80.84 | Interaction: F (4, 44) 0,060; p. 0,706 | CON vs VPA: | 0.0236* |
| RSV + VPA : 247.2±30.44 | | | | CON vs RSV+VPA: | 0.0132* |
| RSV+VPA: 229.1±138.2 | | | | RSV vs VPA: | 0.086# |
| AQP4 CON: 522.8±86.07 RSV: 400.5±94.07 VPA: 298.3±119.1 RSV+VPA: 433.3±213.3 AQP4 CON: 555.4±160.3 RSV: 354.4±76.63 RSV: 54.0±42.98 RSV+VPA: 366.8±167.8 AQP4 CON: 428.0±68.79 RSV: 354.1±90.40 PrL VPA: 213.2±45.47 (II/III) RSV+VPA: 314.6±144.4 AQP4 CON: 428.0±68.79 RSV: 354.4±6.4144.4 Interaction: F (1, 16) = 4.879; p = 0.0421* VPA: F (1, 16) = 2.709; p = 0.1193 RSV: F (1, 16) = 0.01204; p = 0.9140 RSV vs RSV+VPA: 0.769 RSV: SSV+VPA: 0.093 CON vs RSV: 0.093 CON vs RSV: 0.093 CON vs RSV: 0.093 CON vs RSV+VPA: 0.769 RSV vs RSV+VPA: 0.769 RSV vs RSV+VPA: 0.769 RSV vs RSV+VPA: 0.722 CON vs RSV: 0.717 CON vs RSV+VPA: 0.722 CON vs RSV: 0.717 RSV vs VPA: 0.0064** RSV vs VPA: 0.0064** RSV vs VPA: 0.0134 RSV vs VPA: 0.134 RSV vs RSV+VPA: 0.987 | (IV/V) | RSV+VPA: 229.1±138.2 | 1.3 v. i (1, 14) = 0.433, p = 0.3003 | RSV vs RSV+VPA: | 0.0489* |
| AQP4 CON: 522.8±86.07 Interaction: F (1, 16) = 4.879; p = 0.0421* CON vs VPA: 0.880 VPA: 298.3±119.1 RSV+VPA: 433.3±213.3 RSV: F (1, 16) = 0.01204; p = 0.9140 RSV vs VPA: 0.787 RSV vs RSV+VPA: 0.999 VPA vs RSV+VPA: 0.589 CON vs RSV+VPA: 0.093 CON vs RSV+VPA: 0.093 CON vs RSV+VPA: 0.093 CON vs RSV: 0.093 CON vs RSV+VPA: 0.0769 RSV vs VPA: 0.0769 RSV vs VPA: 0.769 RSV vs RSV+VPA: 0.722 CON vs RSV+VPA: 0.0064** CON vs RSV+VPA: 0.0064** CON vs RSV+VPA: 0.0064** CON vs RSV+VPA: 0.337 RSV: 354.1±90.40 VPA: F (1, 16) = 9.964; p = 0.0061** RSV vs VPA: 0.0337 RSV vs VPA: 0.134 RSV vs RSV+VPA: 0.987 RSV v | | The same of the sa | | VPA vs RSV+VPA: | >0.999 |
| RSV: 400.5±94.07 | | | | | 0.586 |
| AQP4 CON: 555.4±160.3 RSV+VPA: 366.8±167.8 AQP4 CON: 428.0±68.79 RSV: 354.1±90.40 PrL VPA: 298.3±14.4+76.63 RSV: 354.1±90.40 VPA: F (1, 16) = 2.709; p = 0.1193 RSV: F (1, 16) = 0.01204; p = 0.9140 RSV vs RSV+VPA: 0.722 CON vs RSV: 0.093 CON vs RSV: 0.0058** CON vs RSV+VPA: 0.0589 CON vs RSV: 0.093 CON vs RSV+VPA: 0.0589 CON vs RSV+VPA: 0.0589 CON vs RSV+VPA: 0.0058** CON vs RSV+VPA: 0.172 RSV vs RSV+VPA: 0.769 RSV vs RSV+VPA: 0.769 RSV vs RSV+VPA: 0.722 CON vs RSV+VPA: 0.737 RSV vs RSV+VPA: 0.337 RSV: F (1, 16) = 9.964; p = 0.0061** RSV vs RSV+VPA: 0.337 | AQP4 | CON: 522.8±86.07 | Interaction: E (1, 16) = 4,970; n = 0,0421* | | 0.064# |
| RSV+VPA: 433.3±213.3 RSV: F (1, 16) = 0.01204; p = 0.9140 RSV vs RSV+VPA: 0.787 | | | | | |
| RSV+VPA: 433.3±213.3 RSV vs RSV+VPA: 0.999 VPA vs RSV+VPA: 0.589 VPA vs RSV+VPA: 0.589 CON vs RSV: 0.093 CON vs RSV: 0.093 CON vs VPA: 0.0058** CON vs RSV+VPA: 0.172 RSV vs VPA: 0.172 RSV vs VPA: 0.172 RSV vs RSV+VPA: 0.769 RSV vs RSV+VPA: 0.769 RSV vs RSV+VPA: 0.722 CON vs RSV+VPA: 0.722 CON vs RSV+VPA: 0.722 CON vs RSV vs RSV+VPA: 0.337 RSV: 354.1±90.40 VPA: 213.2±45.47 VPA: 213.2±45.47 RSV+VPA: 314.6±144.4 RSV: F(1, 16) = 0.1171; p = 0.7367 RSV vs RSV+VPA: 0.987 RSV vs RSV+VPA: 0.987 | | | | | 0.787 |
| $ \begin{array}{c ccccccccccccccccccccccccccccccccccc$ | (11/111) | RSV+VPA: 433.3±213.3 | πον. τ (τ, το) = 0.01204, β = 0.0140 | RSV vs RSV+VPA: | 0.999 |
| $ \begin{array}{c ccccccccccccccccccccccccccccccccccc$ | | | | | |
| aCC (IV/V) RSV+VPA: 354.4 ± 76.63 VPA: 254.0 ± 42.98 RSV+VPA: 366.8 ± 167.8 VPA: $F(1, 16) = 6.711$; $p = 0.0197^*$ RSV vs VPA: 90.0099 RSV vs RSV+VPA: 90.0099 RSV vs RSV+VPA: 90.0099 RSV vs RSV+VPA: 90.0099 RSV: 90.0099 | | | | | |
| ACC (IV/V) VPA: 254.0 ± 42.98 RSV+VPA: 366.8 ± 167.8 VPA: $F(1, 16) = 6.711$; $p = 0.0197^*$ RSV: $F(1, 16) = 0.6246$; $p = 0.4409$ RSV vs RSV+VPA: $F(1, 16) = 0.0197^*$ RSV vs RSV+VPA: $F(1, 16) = 0.0117^*$ RSV vs RSV+VP | AQP4 | | Interaction: F (1 16) - 7 910: n - 0 0125* | | |
| RSV + VPA: 366.8±167.8 RSV: F (1, 16) = 0.6246; p = 0.4409 RSV vs RSV + VPA: 0.769 | | | | | |
| RSV vs RSV+VPA: \$0.999 VPA vs RSV+VPA: \$0.999 VPA vs RSV+VPA: \$0.722 | | | | | |
| AQP4 CON: 428.0±68.79 RSV: 354.1±90.40 PrL VPA: 213.2±45.47 (II/III) RSV+VPA: 314.6±144.4 CON vs RSV: 0.717 | (IV/V) | RSV+VPA: 366.8±167.8 | 110 1.1 (1, 10) 0102 10 , p 011 100 | | |
| $ \begin{array}{llllllllllllllllllllllllllllllllllll$ | | | | | |
| RSV: 354.1±90.40 VPA: 213.2±45.47 (II/III) RSV+VPA: 314.6±144.4 Interaction: F (1, 16) = 4.736; p = 0.0449* VPA: F (1, 16) = 9.964; p = 0.0061** RSV: F (1, 16) = 0.1171; p = 0.7367 RSV vs RSV+VPA: 0.134 RSV vs RSV+VPA: 0.987 | | | | | |
| PrL VPA: 213.2±45.47 RSV: F (1, 16) = 9.964; p = 0.0061** RSV vs VPA: 0.134 RSV vs RSV+VPA: 0.987 | AQP4 | | Interaction: F (1, 16) – 4,736: n – 0,0440* | | |
| (II/III) RSV+VPA: 314.6±144.4 RSV: F (1, 16) = 0.1171; p = 0.7367 $\frac{RSV \vee S \vee PA:}{RSV \vee S \wedge RSV + VPA:} \frac{0.134}{0.987}$ | | | | | |
| (IVIII) R5V+VPA: 314.6±144.4 R5V vs R5V+VPA: 0.987 | | | | | |
| VPA vs RSV+VPA: 0.500 | (11/111) | RSV+VPA: 314.6±144.4 | (., .σ, σ , ρ = σσσ. | | |
| | | | | VPA vs RSV+VPA: | 0.500 |

| | | | CON vs RSV: | 0.216 |
|----------|--|---|-----------------|-----------|
| AQP4 | CON: 482.0±107.3 | Internation: F (4, 4C) 7,440; n 0,0407* | CON vs VPA: | 0.0061** |
| | RSV: 339.8±92.58 | Interaction: F (1, 16) = 7.142; p = 0.0167* | CON vs RSV+VPA: | 0.266 |
| PrL | | VPA: F (1, 16) = 7.278; p = 0.0158* RSV: F (1, 16) = 0.1057; p = 0.7493 | RSV vs VPA: | 0.501 |
| (IV/V) | RSV+VPA: 338.6±157.5 | K3V. F(1, 10) = 0.1037, p = 0.7493 | RSV vs RSV+VPA: | >0.999 |
| | VPA vs RSV+VPA: | 0.576 | | |
| 1 | | | CON vs RSV: | 0.331 |
| AQP4 | CON: 466.8±100.4 RSV: 356.7±105.0 VPA: 193.8±27.21 | Interaction: F (1, 16) = 4.930; p = 0.0412* VPA: F (1, 16) = 19.05; p = 0.0005*** RSV: F (1, 16) = 0.1895; p = 0.6692 | CON vs VPA: | 0.0009*** |
| | | | CON vs RSV+VPA: | 0.0237* |
| IL | | | RSV vs VPA: | 0.0736# |
| (11/111) | RSV+VPA: 267.8±111.5 | | RSV vs RSV+VPA: | 0.668 |
| | | | VPA vs RSV+VPA: | 0.817 |
| - 3 | | | CON vs RSV: | 0.184 |
| AQP4 | CON: 446.3±70.78 | Intercetion: E (1, 16) - 7,072: n - 0,0171* | CON vs VPA: | 0.0029** |
| | RSV: 333.9±87.51 | Interaction: F (1, 16) = 7.073; p = 0.0171* | CON vs RSV+VPA: | 0.122 |
| IL | VPA: 235.7±52.18 | VPA: F (1, 16) = 10.13; p = 0.0058** RSV: F (1, 16) = 0.2106; p = 0.6525 | RSV vs VPA: | 0.351 |
| (IV/V) | RSV+VPA: 315.0±108.4 | 1.0 v. 1 (1, 10) = 0.2 100, p = 0.0323 | RSV vs RSV+VPA: | 0.999 |
| | | | VPA vs RSV+VPA: | 0.643 |

II/III: upper cortical layers; **IV/V**: deeper cortical layers; **aCC**: anterior cingulate cortex; **IL**: infralimbic cortex; **PrL**: prelimbic cortex; **SD**: standard deviation. p <0.05 was considered significant. *p<0.05, **p<0.01, ***p<0.001, ***p<0.0001. Statistical analyses: two-way ANOVA parametric test followed by Sidak. Ncon: 5, Nrsv: 6, Nvpa: 6, Nrsv+vpa: 5. "Mean" represents de mean of fluorescence.

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Table 5 – Descriptive statistics of the AQP1 and AQP4 distribution profile in AmR Mean ± SD F (DFn, DFd); p Value Pairwise comparisons CON vs RSV: 0.8603 CON vs VPA: CON: 394.4±13.92 0.0621# Interaction: F(1, 17) = 0.0279; p = 0.869AQP1 CON vs RSV+VPA: 0.0049** RSV: 354.3±46.69 VPA: F(1, 17) = 18.95; p = 0.0004***VPA: 291.2±67.83 RSV vs VPA: 0.3386 RSV: F(1, 17) = 3.226; p = 0.090RSV+VPA: 242.8±67.61 RSV vs RSV+VPA: 0.0249* VPA vs RSV+VPA: 0.6718 CON vs RSV: 0.887 CON: 112.9±15.93 CON vs VPA: 0.991 Interaction: F(1, 16) = 0.2881; p = 0.5988AQP4 CON vs RSV+VPA: RSV: 124.6±11.77 0.939 VPA: F (1, 16) = 0.1118; p = 0.7425 RSV vs VPA: VPA: 119.7±10.60 0.998 RSV: F (1, 16) = 0.9222; p = 0.3512 RSV+VPA: 123.0±26.89 RSV vs RSV+VPA: >0.999 VPA vs RSV+VPA: 0.999

AmR: amygdala region; **SD**: standard deviation. p <0.05 was considered significant. *p<0.05, **p<0.01, ***p<0.001. Statistical analyses: two-way ANOVA parametric test followed by Sidak. Ncon: 5, Nrsv: 6, Nrpa: 6, Nrsv+vpa: 5. "Mean" represents de mean of fluorescence.

Table 6 - Descriptive statistics of the AQP1 and AQP4 distribution profile in

hippocampus.

| | hippocampus. | F/DF DE IV V-L | Daimed | |
|---------|---|--|-----------------|--------|
| | Mean ± SD | F (DFn, DFd); p Value | Pairwise compa | |
| | | | CON vs RSV: | 0.694 |
| AQP1 | CON: 420.2±68.76 | Interaction: F $(1, 18) = 0.049$; p = 0.826 | CON vs VPA: | 0.999 |
| ۸۹. ۱ | RSV: 315.9±121.7 | VPA: F (1, 18) = 0.697; p = 0.415 | CON vs RSV+VPA: | 0.287 |
| DG | VPA: 387.9±154.8 | RSV: F (1, 18) = 4.837; p = 0.041* | RSV vs VPA: | 0.905 |
| | RSV+VPA: 260.1±123.2 | πον. τ (τ, το) = ποστ, ρ = σ.σ.τ | RSV vs RSV+VPA: | 0.976 |
| | | | VPA vs RSV+VPA: | 0.482 |
| | | | CON vs RSV: | 0.607 |
| AQP1 | CON: 418.9±72.71 | Interaction: $F(1, 18) = 0.0057$; $p = 0.940$ | CON vs VPA: | 0.841 |
| AUFI | RSV: 316.8±114.7 | | CON vs RSV+VPA: | 0.136 |
| CA1 | VPA: 341.8±107.7 | VPA: F (1, 18) = 2.413; p = 0.137 RSV: F (1, 18) = 4.340; p = 0.052# | RSV vs VPA: | 0.999 |
| OAI | RSV+VPA: 246.9±136.7 | | RSV vs RSV+VPA: | 0.892 |
| | - | | VPA vs RSV+VPA: | 0.679 |
| | | | CON vs RSV: | 0.768 |
| AOD1 | CON: 424.1±50.26 | Interaction: E (1, 19) - 0.245; n - 0.626 | CON vs VPA: | 0.895 |
| AQP1 | RSV: 340.3±107.3 | Interaction: F (1, 18) = 0.245; p = 0.626 VPA: F (1, 18) = 3.858; p = 0.065 RSV: F (1, 18) = 5.331; p = 0.033* | CON vs RSV+VPA: | 0.057 |
| CA2 | VPA: 356.3±126.8 | | RSV vs VPA: | >0.999 |
| UAZ | RSV+VPA: 226.6±124.3 | $1.3 \text{ v. } \Gamma (1, 10) = 0.331, p = 0.033$ | RSV vs RSV+VPA: | 0.465 |
| 4 | | | VPA vs RSV+VPA: | 0.327 |
| | | | CON vs RSV: | 0.972 |
| A O D 4 | CON: 366.6±54.55 | Literative F (4, 40) 0 504 0 400 | CON vs VPA: | 0.997 |
| AQP1 | RSV: 316.9±105.5 | V: 316.9±105.5 A: 335.5±120.2 VPA: F (1, 18) = 0.564; p = 0.462 - VPA: F (1, 18) = 2.071; p = 0.167 | CON vs RSV+VPA: | 0.212 |
| 040 | VPA: 335.5±120.2 | | RSV vs VPA: | 0.999 |
| CA3 | RSV+VPA: 217.8±124.0 RSV: F (1, 18) = 3.418; p = 0.081# | RSV vs RSV+VPA: | 0.592 | |
| - 1 | | VPA vs RSV+VPA: | 0.404 | |
| | | | CON vs RSV: | 0.967 |
| | CON: 297.6±56.51 | - | CON vs VPA: | 0.718 |
| AQP4 | RSV: 338.4±50.03 | Interaction: F (1, 19) = 4.936; p = 0.0387* | CON vs RSV+VPA: | 0.929 |
| | VPA: 361.9±96.46 | VPA: F (1, 19) = 0.1120; p = 0.7415 | RSV vs VPA: | 0.997 |
| DG | RSV+VPA: 251.4±95.38 | RSV: F (1, 19) = 1.048; p = 0.3187 | RSV vs RSV+VPA: | 0.437 |
| | 1 > | | VPA vs RSV+VPA: | 0.135 |
| | | | CON vs RSV: | 0.935 |
| | CON: 300.7±44.61 | | CON vs VPA: | 0.929 |
| AQP4 | RSV: 346.4±66.26 | Interaction: $F(1, 19) = 3.456$; $p = 0.0786$ # | CON vs RSV+VPA: | 0.982 |
| | VPA: 343.9±107.4 | VPA: F (1, 19) = 0.2992; p = 0.5908 | RSV vs VPA: | >0.999 |
| CA1 | RSV+VPA: 267.2±65.51 | RSV: F (1, 19) = 0,2202; p = 0.6442 | RSV vs RSV+VPA: | 0.505 |
| | | | VPA vs RSV+VPA: | 0.447 |
| | | | CON vs RSV: | 0.901 |
| | CON: 307.7±73.17 | | CON vs VPA: | 0.979 |
| AQP4 | RSV: 351.1±40.68 | Interaction: $F(1, 19) = 2.790$; $p = 0.1113$ | CON vs RSV+VPA: | 0.995 |
| | VPA: 336.3±79.70 | VPA: F (1, 19) = 0.4353; p = 0.5173 | RSV vs VPA: | 0.999 |
| CA2 | RSV+VPA: 285.2±62.61 | RSV: F (1, 19) = 0.01884; p = 0.8923 | RSV vs RSV+VPA: | 0.540 |
| | NOVIVI A. 200.2102.01 | | VPA vs RSV+VPA: | 0.540 |
| | | | CON vs RSV: | |
| | CON. 277 4: 40 00 | | | 0.353 |
| AQP4 | CON: 277.4±48.82 | Interaction: F $(1, 19) = 4.422$; p = 0.0490 * | CON vs VPA: | 0.967 |
| | RSV: 342.3±27.86 | VPA: F (1, 19) = 0.9866; p = 0.3331 | CON vs RSV+VPA: | >0.999 |
| CA3 | VPA: 302.4±68.88 | RSV: F (1, 19) = 0.6090; p = 0.4448 | RSV vs VPA: | 0.770 |
| | RSV+VPA: 272.6±51.07 | , | RSV vs RSV+VPA: | 0.238 |
| | | | VPA vs RSV+VPA: | 0.909 |

DG: dentate gyrus; **SD**: standard deviation. p <0.05 was considered significant. *p<0.05. Statistical analyses: two-way ANOVA parametric test followed by Sidak. Ncon: 5, Nrsv: 6, Nrpa: 6, Nrsv+vpa: 5. "Mean" represents de mean of fluorescence.

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Table 7 – Descriptive statistics of the GFAP fluorescence and number of GFAP+-astrocytes in pSSA

| a | strocytes in pasa | | | |
|----------|--------------------------------------|---|-----------------|---------|
| | Mean ± SD | F (DFn, DFd); p Value | Pairwise compa | arisons |
| | | | CON vs RSV: | 0.983 |
| GFAP | CON: 153.1±38.39 | Interaction: $F(1, 18) = 0.1940$; $p = 0.6649$ | CON vs VPA: | 0.6985 |
| GI AF | RSV: 177.3±49.76 | VPA: F (1, 18) = 5.797; p = 0.0270* | CON vs RSV+VPA: | 0.105 |
| (11/111) | VPA: 201.6±35.43 | RSV: F (1, 18) = 3.797, p = 0.0270 | RSV vs VPA: | 0.978 |
| (11/111) | RSV+VPA: 247.4±93.58 | KSV. F(1, 10) = 2.023, p = 0.1721 | RSV vs RSV+VPA: | 0.306 |
| | | | VPA vs RSV+VPA: | 0.746 |
| | | | CON vs RSV: | 0.822 |
| GFAP | CON: 109.6±45.17 | Interaction: E (1, 10) = 0.2125; n = 0.5021 | CON vs VPA: | 0.991 |
| GFAF | RSV: 146.5±32.95 | Interaction: F (1, 18) = 0.3125; p = 0.5831 VPA: F (1, 18) = 2.004; p = 0.1740 | CON vs RSV+VPA: | 0.133 |
| (IV/V) | VPA: 128.4±8.990 | RSV: F (1, 18) = 5.017; p = 0.0380* | RSV vs VPA: | 0.991 |
| (14/4) | RSV+VPA: 189.9±91.40 | | RSV vs RSV+VPA: | 0.695 |
| | | | VPA vs RSV+VPA: | 0.324 |
| | | | CON vs RSV: | >0.999 |
| Number | CON: 66.40±19.51 | Interaction: E (1.10) 0.7000; p. 0.4106 | CON vs VPA: | 0.459 |
| Number | RSV: 65.00±28.09 | Interaction: F (1, 18) = 0.7098; p = 0.4106 VPA: F (1, 18) = 10.99; p = 0.0038** | CON vs RSV+VPA: | 0.082# |
| (11/111) | VPA: 89.00±22.12 | RSV: F (1, 18) = 10.99, p = 0.0038 | RSV vs VPA: | 0.341 |
| (11/111) | RSV+VPA: 103.0±8.485 | K3V. F(1, 10) = 0.4732, p = 0.4994 | RSV vs RSV+VPA: | 0.051# |
| | | | VPA vs RSV+VPA: | 0.875 |
| 4 | | | CON vs RSV: | 0.373 |
| Number | CON: 39.40±18.42 | Interpotion F (4, 10) 0.4040, p. 0.5000 | CON vs VPA: | 0.067# |
| number | RSV: 63.00±18.45 VPA: 74.50±28.47 | Interaction: F (1, 18) = 0.4240; p = 0.5232 | CON vs RSV+VPA: | 0.0117* |
| (IV/V) | | VPA: F (1, 18) = 11.05; p = 0.0038** RSV: F (1, 18) = 4.086; p = 0.0584# | RSV vs VPA: | 0.922 |
| (14/4) | RSV+VPA: 86.60±11.67 | Nov. $\Gamma(1, 10) = 4.000, p = 0.0004#$ | RSV vs RSV+VPA: | 0.373 |
| | | | VPA vs RSV+VPA: | 0.921 |

II/III: upper cortical layers; **IV/V**: deeper cortical layers; **pSSA**: primary somatosensory area; **SD**: standard deviation. p <0.05 was considered significant. *p<0.05, **p<0.01. Statistical analyses: two-way ANOVA parametric test followed by Sidak. Ncon: 5, Nrsv: 6, Nvpa: 6, Nrsv+vpa: 5.

Table 8 – Descriptive statistics of the GFAP fluorescence and number of GFAP+-astrocytes in mPFC

| a | strocytes in mPFC | F (DF#, DF=1) V=1 | Deimodes second | · · · · · · · · · · · · · · · · · · · |
|-------------|----------------------|---|-----------------|---------------------------------------|
| | Mean ± SD | F (DFn, DFd); p Value | Pairwise compa | |
| 0545 | CON. 240 5 42 22 | | CON vs RSV: | 0.980 |
| GFAP | CON: 316.5±48.96 | Interaction: F (1, 14) = 2.882; p = 0.1117 | CON vs VPA: | 0.0789# |
| 000 | RSV: 366.6±87.41 | VPA: F (1, 14) = 5.252; p = 0.0379* | CON vs RSV+VPA: | 0.869 |
| aCC | VPA: 522.5±157.7 | RSV: F (1, 14) = 0.5305; p = 0.4784 | RSV vs VPA: | 0.269 |
| (II/III) | RSV+VPA: 397.2±130.6 | , , | RSV vs RSV+VPA: | 0.998 |
| | PA | | VPA vs RSV+VPA: | 0.554 |
| | | | CON vs RSV: | 0.999 |
| GFAP | CON: 307.0±30.66 | Interaction: F (1, 14) = 7.310; p = 0.0171* | CON vs VPA: | 0.0348* |
| -00 | RSV: 319.9±44.57 | VPA: F (1, 14) = 3.549; p = 0.0805 | CON vs RSV+VPA: | >0.999 |
| aCC | VPA: 422.3±49.83 | RSV: F (1, 14) = 4.806; p = 0.0458* | RSV vs VPA: | 0.071# |
| (IV/V) | RSV+VPA: 299.3±81.86 | • | RSV vs RSV+VPA: | 0.993 |
| | | | VPA vs RSV+VPA: | 0.0321* |
| | @ | | CON vs RSV: | 0.999 |
| GFAP | CON: 297.9±35.88 | Interaction: F (1, 14) = 1.234; p = 0.2853 | CON vs VPA: | 0.231 |
| | RSV: 313.1±61.58 | VPA: F (1, 14) = 4.171; p = 0.0604# | CON vs RSV+VPA: | 0.923 |
| PrL | VPA: 406.1±79.69 | RSV: F (1, 14) = 0.4471; p = 0.5146 | RSV vs VPA: | 0.377 |
| (II/III) | RSV+VPA: 345.0±106.4 | то то (т, т т) | RSV vs RSV+VPA: | 0.987 |
| | | | VPA vs RSV+VPA: | 0.825 |
| | | | CON vs RSV: | 0.979 |
| GFAP | CON: 312.7±31.30 | Interaction: F (1, 14) = 4.820; p = 0.0455* | CON vs VPA: | 0.061# |
| | RSV: 288.5±44.65 | VPA: F (1, 14) = 4.820 , p = 0.0435 | CON vs RSV+VPA: | 0.960 |
| PrL | VPA: 416.3±48.23 | RSV: F (1, 14) = 3.930, p = 0.0074# RSV: F (1, 14) = 10.05; p = 0.0068** | RSV vs VPA: | 0.0158* |
| (IV/V) | RSV+VPA: 283.2±80.47 | 13 V. Γ (1, 14) = 10.03, β = 0.0000 | RSV vs RSV+VPA: | >0.999 |
| 100 | | | VPA vs RSV+VPA: | 0.0173* |
| 100 | | | CON vs RSV: | 0.955 |
| GFAP | CON: 338.2±49.43 | Interestion: F (4, 4.4) 0.2527; n. 0.5624 | CON vs VPA: | 0.746 |
| | RSV: 297.2±55.84 | Interaction: F (1, 14) = 0.3527; p = 0.5621 | CON vs RSV+VPA: | 0.999 |
| IL | VPA: 405.2±106.1 | VPA: F (1, 14) = 1.659; p = 0.2186 RSV: F (1, 14) = 3.036; p = 0.1034 | RSV vs VPA: | 0.265 |
| (11/111) | RSV+VPA: 322.0±87.58 | N3V. F $(1, 14) = 3.030, p = 0.1034$ | RSV vs RSV+VPA: | 0.997 |
| | | | VPA vs RSV+VPA: | 0.593 |
| | 1 \ | | CON vs RSV: | 0.723 |
| GFAP | CON: 316.5±24.27 | Internation: F (4, 44) 0,450; n 0,4600 | CON vs VPA: | 0.204 |
| | RSV: 270.5±60.38 | Interaction: F (1, 14) = 2.159; p = 0.1638 | CON vs RSV+VPA: | 0.886 |
| IL | VPA: 398.6±70.81 | VPA: F (1, 14) = 3.182; p = 0.0962# | RSV vs VPA: | 0.0176* |
| (IV/V) | RSV+VPA: 278.4±50.56 | RSV: F (1, 14) = 10.84; p = 0.0053** | RSV vs RSV+VPA: | >0.999 |
| | | | VPA vs RSV+VPA: | 0.0383* |
| | | | CON vs RSV: | 0.869 |
| Number | CON: 109.2±25.58 | | CON vs VPA: | 0.240 |
| | RSV: 139.8±61.92 | Interaction: F (1, 15) = 0.04255; p = 0.8393 | CON vs RSV+VPA: | 0.075 |
| aCC | VPA: 170.0±22.97 | VPA: F (1, 15) = 7.828; p = 0.0135* | RSV vs VPA: | 0.876 |
| (11/111) | RSV+VPA: 192.3±54.32 | RSV: F (1, 15) = 1.705; p = 0.2113 | RSV vs RSV+VPA: | 0.451 |
| | P) | | VPA vs RSV+VPA: | 0.975 |
| | | | CON vs RSV: | 0.897 |
| Number | CON: 85.00±21.71 | | CON vs VPA: | 0.0443* |
| | RSV: 113.6±48.03 | Interaction: F (1, 15) = 1.542; p = 0.2334 | CON vs RSV+VPA: | 0.243 |
| aCC | VPA: 170.0±25.50 | VPA: F (1, 15) = 8.959; p = 0.0091** | RSV vs VPA: | 0.303 |
| (IV/V) | RSV+VPA: 148.8±70.04 | RSV: F (1, 15) = 0.033; p = 0.8572 | RSV vs RSV+VPA: | 0.818 |
| ` ' | | | VPA vs RSV+VPA: | 0.979 |
| | | | CON vs RSV: | >0.999 |
| Number | CON: 111.4±45.53 | | CON vs VPA: | 0.756 |
| HUITIDE | RSV: 114.0±54.61 | Interaction: $F(1, 15) = 0.4302$; $p = 0.5218$ | CON vs RSV+VPA: | 0.730 |
| PrL | VPA: 156.2±30.54 | VPA: F (1, 15) = 6.018; p = 0.0269* | RSV vs VPA: | 0.801 |
| (II/III) | RSV+VPA: 191.5±81.46 | RSV: F (1, 15) = 0.5779; p = 0.4589 | | |
| (11/111) | NOVTVFA. 191.0±01.40 | | RSV vs RSV+VPA: | 0.262 |
| | | | VPA vs RSV+VPA: | 0.921 |

| | | | CON vs RSV: | 0.998 |
|----------|----------------------|---|-----------------|---------|
| Number | CON: 93.20±19.65 | Interaction: F (1, 15) 0.201; p. 0.5079 | CON vs VPA: | 0.0120* |
| | RSV: 105.0±39.21 | Interaction: F (1, 15) = 0.291; p = 0.5978 VPA: F (1, 15) = 20.99; p = 0.0004*** | CON vs RSV+VPA: | 0.0342* |
| PrL | VPA: 188.2±21.46 | RSV: F (1, 15) = 20.99, p = 0.0004 RSV: F (1, 15) = 0.010; p = 0.9240 | RSV vs VPA: | 0.0307* |
| (IV/V) | RSV+VPA: 180.0±70.15 | N3V.1 (1, 13) = 0.010, p = 0.9240 | RSV vs RSV+VPA: | 0.081# |
| | | | VPA vs RSV+VPA: | 0.999 |
| | | | CON vs RSV: | >0.999 |
| Number | CON: 103.6±31.94 | Interaction: F (1, 15) 0.2200; p. 0.6452 | CON vs VPA: | 0.678 |
| | RSV: 106.6±79.16 | Interaction: F (1, 15) = 0.2208; p = 0.6452 VPA: F (1, 15) = 5.935; p = 0.0278* | CON vs RSV+VPA: | 0.292 |
| IL | VPA: 158.0±49.46 | RSV: F (1, 15) = 0.3345; p = 0.5716 | RSV vs VPA: | 0.729 |
| (11/111) | RSV+VPA: 187.0±70.94 | | RSV vs RSV+VPA: | 0.328 |
| | | | VPA vs RSV+VPA: | 0.980 |
| | | | CON vs RSV: | >0.999 |
| Number | CON: 82.20±33.26 | Interaction: : F (1, 15) = 4.55e-005; p = | CON vs VPA: | 0.084# |
| - | RSV: 89.00±36.88 | 0.9947 | CON vs RSV+VPA: | 0.074# |
| IL | VPA: 166.4±50.58 | VPA: F (1, 15) = 14.41; p = 0.0018** | RSV vs VPA: | 0.128 |
| (IV/V) | RSV+VPA: 173.5±70.11 | RSV: F (1, 15) = 0.097; p = 0.7587 | RSV vs RSV+VPA: | 0.118 |
| | | | VPA vs RSV+VPA: | >0.999 |

II/III: upper cortical layers; **IV/V**: deeper cortical layers; **aCC**: anterior cingulate cortex; **IL**: infralimbic cortex; **PrL**: prelimbic cortex; **SD**: standard deviation. p <0.05 was considered significant. *p<0.05, **p<0.01, ***p<0.001. Statistical analyses: two-way ANOVA parametric test followed by Sidak. Ncon: 5, Nrsv: 6, Nvpa: 6, Nrsv+vpa: 5.

Accepted

Table 9 – Descriptive statistics of the GFAP fluorescence and number of GFAP*-astrocytes in AmR

| _ | | | | |
|--------|----------------------|---|-----------------|---------|
| | Mean ± SD | F (DFn, DFd); p Value | Pairwise compa | arisons |
| | | | CON vs RSV: | 0.996 |
| | CON: 88.98±30.23 | Interestion: F (4, 40) | CON vs VPA: | >0.999 |
| GFAP | RSV: 75.56±14.83 | Interaction: F (1, 16) = 4.728 ; p = 0.0450 * | CON vs RSV+VPA: | 0.187 |
| | VPA: 82.66±14.10 | VPA: F (1, 16) = 3.357; p= 0.0856 RSV: F (1, 16) = 2.095; p = 0.1670 | RSV vs VPA: | >0.999 |
| RSV+ | RSV+VPA: 149.6±74.12 | K3V. F (1, 10) = 2.095, p = 0.1070 | RSV vs RSV+VPA: | 0.069# |
| | | | VPA vs RSV+VPA: | 0.119 |
| | | | CON vs RSV: | 0.083# |
| | CON: 33.80±7.463 | Interaction: F (1, 16) = 14.26; p = 0.0017** | CON vs VPA: | 0.398 |
| Number | RSV: 11.40±5.225 | | CON vs RSV+VPA: | 0.978 |
| | | VPA: F (1, 16) = 1.307; p = 0.2697 | RSV vs VPA: | 0.948 |
| | RSV+VPA: 39.80±21.74 | RSV: F (1, 16) = 0.01080; p = 0.9185 | RSV vs RSV+VPA: | 0.0185* |
| | 3 | | VPA vs RSV+VPA: | 0.111 |

AmR: amygdala region; **SD**: standard deviation. p <0.05 was considered significant. *p<0.05, **p<0.01. Statistical analyses: two-way ANOVA parametric test followed by Sidak. Ncon: 5, Nrsv: 6, Nvpa: 6, Nrsv+vpa: 5.

Table 10 - Descriptive statistics of the GFAP fluorescence and number of

GFAP⁺-astrocytes in hippocampus.

| | Mean ± SD | F (DFn, DFd); p Value | Pairwise compa | arisons |
|--------|----------------------|---|-------------------|----------|
| | | 7 | CON vs RSV: | 0.170 |
| | CON: 458.6±93.64 | | CON vs VPA: | 0.751 |
| GFAP | RSV: 642.1±62.81 | Interaction: F (1, 19) = 8.292; p = 0.0096** | CON vs RSV+VPA: | 0.999 |
| DO | VPA: 553.7±135.4 | VPA: F (1, 19) = 1.130; p = 0.3012 | RSV vs VPA: | 0.806 |
| DG | RSV+VPA: 435.8±162.8 | RSV: F (1, 19) = 0.3929; p = 0.5382 | RSV vs RSV+VPA: | 0.0753# |
| | | | VPA vs RSV+VPA: | 0.484 |
| | | | CON vs RSV: | 0.262 |
| | CON: 445.0±121.7 | | CON vs VPA: | 0.964 |
| GFAP | RSV: 623.1±140.4 | Interaction: F (1, 19) = 6.811; p = 0.0172* | CON vs RSV+VPA: | 0.988 |
| | VPA: 509.1±168.7 | VPA: F (1, 19) = 2.175; p = 0.1567 RSV: F (1, 19) = 0.2951; p = 0.5933 | RSV vs VPA: | 0.656 |
| CA1 | RSV+VPA: 392.4±81.06 | | RSV vs RSV+VPA: | 0.0611# |
| 7 | 100 | | VPA vs RSV+VPA: | 0.578 |
| | 1 | | CON vs RSV: | 0.0323* |
| | CON: 409.5±170.2 | | CON vs VPA: | 0.987 |
| GFAP | RSV: 635.5±73.25 | Interaction: F (1, 19) = 6.286; p = 0.0214^* | CON vs RSV+VPA: | 0.998 |
| 040 | VPA: 453.4±109.4 | VPA: F (1, 19) = 2.539; p = 0.1276 | RSV vs VPA: | 0.077# |
| CA2 | RSV+VPA: 438.6±86.79 | RSV: F (1, 19) = 4.842; p = 0.0403* | RSV vs RSV+VPA: | 0.059# |
| | | | VPA vs RSV+VPA: | >0.999 |
| - 4 | | | CON vs RSV: | 0.0023** |
| | CON: 427.0±97.55 | | CON vs VPA: | 0.999 |
| GFAP | RSV: 682.6±36.56 | Interaction: F (1, 19) = 9.676; p = 0.0058** | CON vs RSV+VPA: | 0.996 |
| 0.10 | VPA: 446.3±119.8 | VPA: F (1, 19) = 6.882; p = 0.0167* RSV: F (1, 19) = 11.25; p = 0.0033** | RSV vs VPA: | 0.0023** |
| CA3 | RSV+VPA: 455.9±87.03 | | RSV vs RSV+VPA: | 0.0047** |
| | | | VPA vs RSV+VPA: | >0.999 |
| | _ | | CON vs RSV: | 0.069# |
| 2.50 | CON: 80.80±11.73 | | CON vs VPA: | 0.441 |
| Number | RSV: 120.8±28.44 | Interaction: F (1, 19) = 6.965; p = 0.0162* | CON vs RSV+VPA: | 0.928 |
| | VPA: 104.4±22.65 | VPA: F (1, 19) = 0.03032; p = 0.8636 | RSV vs VPA: | 0.798 |
| DG | RSV+VPA: 93.83±24.41 | RSV: F (1, 19) = 2.353; p = 0.1416 | RSV vs RSV+VPA: | 0.333 |
| | | | VPA vs RSV+VPA: | 0.959 |
| | 1 \ | | CON vs RSV: | 0.307 |
| | CON: 71.00±19.25 | | CON vs VPA: | 0.965 |
| Number | RSV: 99.60±29.59 | Interaction: F (1, 19) = 1.825; p = 0.1926 | CON vs RSV+VPA: | 0.908 |
| 044 | VPA: 81.71±19.49 | VPA: F (1, 19) = 0.04939; p = 0.8265 | RSV vs VPA: | 0.720 |
| CA1 | RSV+VPA: 84.67±21.86 | RSV: F (1, 19) = 2.762; p = 0.1129 | RSV vs RSV+VPA: | 0.869 |
| | | | VPA vs RSV+VPA: | >0.999 |
| | | | CON vs RSV: | 0.0093** |
| N1 | CON: 76.40±12.40 | Literative F (4, 40) - 0.4555 | CON vs VPA: | 0.977 |
| Number | RSV: 118.8±16.63 | Interaction: F (1, 19) = 0.4575; p = 0.5069 | CON vs RSV+VPA: | 0.0107* |
| C 4 2 | VPA: 84.29±20.77 | VPA: F (1, 19) = 0.1254; p = 0.7272 | RSV vs VPA: | 0.0253* |
| CA2 | RSV+VPA: 116.3±19.82 | RSV: F (1, 19) = 23.66; p = 0.0001*** | RSV vs RSV+VPA: | >0.999 |
| | F) | | VPA vs RSV+VPA: | 0.0299* |
| | | | CON vs RSV: | 0.0174* |
| | CON: 88.20±17.75 | | CON vs VPA: | 0.999 |
| Number | RSV: 132.4±18.50 | Interaction: F (1, 19) = 4.553; p = 0.0461* | CON vs RSV+VPA: | 0.999 |
| 0.10 | VPA: 84.29±21.88 | VPA: F (1, 19) = 6.695; p = 0.0181* | RSV vs VPA: | 0.0045** |
| CA3 | RSV+VPA: 91.67±22.23 | RSV: F (1, 19) = 8.936; p = 0.0075** | RSV vs RSV+VPA: | 0.0232* |
| | 3 | | VPA vs RSV+VPA: | 0.988 |
| | | | TITLE TO THE TENT | 3.000 |

DG: dentate gyrus; **SD**: standard deviation. p <0.05 was considered significant. *p<0.05, **p<0.01, ***p<0.001. Statistical analyses: two-way ANOVA parametric test followed by Sidak. Ncon: 5, Nrsv: 6, Nrsv+VPA: 5.