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A facile one pot multi component synthesis of alkyl 4-oxo-coumarinyl ethylidene hydrazono-thiazolidin-5-ylidene acetates and their antiviral activity

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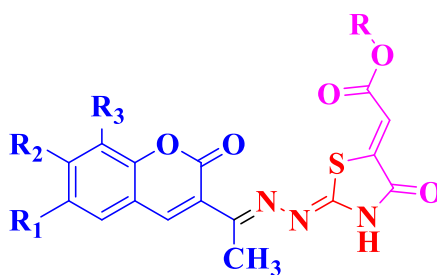
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IV (1-31)

Abstract: An efficient one-pot synthesis of alkyl 4-oxo-coumarinyl ethylidene hydrazono-thiazolidin-5-ylidene acetate derivatives has successfully been achieved via a three component cyclization reaction of various substituted 3-acetyl coumarins, thiosemicarbazide and dialkyl acetylenedicarboxylates, in presence of acetic acid. The isolated products were obtained in pure form with high yields through simple workup. The newly synthesised compounds structure was established on the basis of spectral (IR, ¹H NMR, ¹³C NMR, ESI-mass) elemental analysis and single crystal X-ray data. All synthesised compounds were screened for their antiviral activity against a broad spectrum of human viruses in different cell cultures. Of the novel synthesised compounds, thirteen compounds exerted activity against

Punta Toro virus, including compound **IV-19**, for which an antiviral potency was noted against a broad panel of DNA and RNA viruses as well.

Keywords: One- pot three - component synthesis, 3-acetylcoumarin, thiosemicarbazide, thiazolidinone and Antiviral activity.

1. Introduction

According to the world health organization, infectious diseases caused by the different members of DNA- and RNA-containing viruses are related with significant proportions of human deaths across the globe. Antiviral agents are considered to be very useful weapons for controlling these infectious diseases. However, the treatment of viral infections still remains an important challenge because of the rapid mutability of the virus resulting in escape mutants and drug-resistant virus strains [1-3]. Due to the emergence of anti-viral resistance, there is a continuous interest in developing novel antiviral agents that have less side effects, high therapeutic indices and are active against different circulating resistant viruses.

Multi component reaction (MCR) chemistry is becoming a promising field of chemistry, due to the many positive implications of economic and ecological issues [4,5]. This MCR approach provides an elegant and rapid synthesis platform with various diversified complex molecules in one-pot, high synthetic efficiency, and minimal waste generation by reducing the reaction stages and the reaction work-up like solvent extraction, purification, selectivity and atom economy.

Numerous biologically active molecules contain various hetero atoms like nitrogen, oxygen, sulphur and selenium. Due to their biological importance, enormous interest was given to the synthesis of these heterocyclic scaffolds resulting in one of the most fertile research domains for medicinal chemists all over the world. Besides all the other heterocyclic systems, functionalized thiazolidinone is an N- and S-containing privileged heterocyclic scaffold, with versatile pharmacological properties such as anticancer [6, 7], antiviral [8,9,10], herbicidal [11], antioxidant [12,], antiinflammatory [13, 14], antimicrobial [15-19], antimalarial [20] and ubiquitously present in a large number of clinically used drugs.

In addition, coumarins and their derivatives are an important class of natural and synthetic heterocyclic compounds with vital pharmacological and material application properties including anti-inflammatory [21], antileishmanial [22], steroid hydroxylase [23], MALDI-FT ICR MS detection of hydrophobic compounds [24], stokes shift calculation [25], anti HIV [26, 27], solid state dye lasers [28], fluorescent chemosensor [29], antitumor agents [30, 31], antituberculosis[32, 33]. From this synthetic, medicinal and molecular diversity point of view and in continuation of our research towards one pot multicomponent synthesis of potential heterocyclic systems [34 - 37] constructing a chemical entity having both coumarin and thiazolidinone heterocyclic motifs along with a potential functional group into a single molecular framework, would be of great interest as these two combined nuclei might result in interesting biological properties.

Here we developed an efficient one pot multi component approach for the synthesis of alkyl - 4-oxo-(coumarinyl ethylidene hydrazono)-thiazolidin-5-ylidene acetates and studied their antiviral properties against a broad spectrum of human viruses in different cell culture medium.

2. Experimental:

2.1 Materials and Methods:

The reagents like thiosemicarbazide, dimethyl acetylene dicarboxylate, diethyl acetylene dicarboxylate, di *tert.* butyl acetylene dicarboxylate and the solvents used in this method are procured from the commercial sources and used without any further purification unless until mentioned. The progress of the reaction was monitored through thin layer chromatography technique by using silica gel plates (60 F₂₅₄, Merck) and the spot visualization was done with ultraviolet radiation and Iodine vapour. The melting points were recorded on digital Stuart melting point apparatus (SMP-30, Stuart, Staffordshire, UK). The FT-IR spectra were recorded on Perkin- Elmer spectrophotometer (Spectrum-100S). All the ¹H NMR and ¹³C NMR were recorded on Bruker at 400 MHz and 100 MHz frequencies and the chemical shift values are given with respect to tetramethylsilane (TMS) as an internal standard. The elemental analysis was performed on Carlo- Erba 1108 instrument. The crystallographic data

was collected on Bruker Kappa Apex II diffractometer with graphite monochromated MoK α radiation. The compounds structure was solved by using X-shell software. The single crystal XRD data was deposited in Cambridge Crystallographic Data Centre and the CCDC number was assigned (CCDC: 1418103).

2.2. General procedure for the synthesis of alkyl - 4-oxo-(coumarinyl ethylidene hydrazono)-thiazolidin-5-ylidene acetates (IV 1-31):

A mixture substituted 3-acetyl coumarin (1 mmol) and thiosemicarbazide (1mmol) were taken in the round bottom flask having 5 mL of acetic acid solvent. The reaction mixture was stirred at room temperature for about 15 minutes. To this reaction mixture dialkylacetylene dicarboxylate (1 mmol) was added. The reaction mixture was stirred and then the temperature of the reaction mixture was raised to 60-65 °C. After completion of the reaction (monitored by the TLC) the reaction mixture was cooled, the solid separated was filtered and recrystallized from a methanol solvent.

2.2.1. (Z)-methyl 2-((Z)-4-oxo-2-((E)-(1-(2-oxo-2H-chromen-3-yl)ethylidene) hydrazono) thiazolidin-5-ylidene)acetate (IV-1) :

Yield : (82%); m.p.: 247 – 249 °C; IR (KBr, ν cm⁻¹): 3431(-NH), 1723(lactone -C=O), 1643 (ester -C=O); 1517 (-C=N); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 2.38 (s, 3H, -CH₃); 3.76 (s, 3H, -OCH₃); 6.68 (s, 1H, -C=H); 7.40 (t, *J* = 7.6 Hz, 1H, -Ar-H); 7.43 – 7.47 (m, 1H, -Ar-H); 7.66 – 7.71 (m, 1H, -Ar-H); 7.91 – 7.93 (m, 1H, -Ar-H); 8.27(s, 1H, -C₄-H of coumarin); 12.98 (s, 1H, -NH); ¹³C NMR (100 MHz, DMSO-*d*₆, ppm): δ 22.5; 57.7; 119.7; 121.2; 123.9; 130.0; 131.3; 134.7; 138.0; 147.2; 147.3; 148.0; 158.8; 164.1; 165.7; 167.4; 170.9; *Anal.* Calcd for C₁₇H₁₃N₃O₅S: C, 54.98; H, 3.53; N, 11.31; S, 8.63; Found: C, 54.92; H, 3.58; N, 11.26; S, 8.56; HRMS (ESI) *m/z* calculated for C₁₇H₁₃N₃NaO₅S [M + Na]⁺: 394.0474, found. 394.0472.

2.2.2. (Z)-methyl 2-((Z)-2-((E)-(1-(6-chloro-2-oxo-2H-chromen-3-yl)ethylidene) hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-2) :

Yield: (85%); m.p.: 267 – 269 °C; IR (KBr, ν cm⁻¹): 3152 (-NH), 1722 (lactone – C=O), 1623 (ester – C=O), 1505 (-C=N); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 2.36 (s, 1H, -CH₃); 3.79 (s, 1H, -OCH₃); 6.67 (s, 1H, -C=H); 7.50 (d, *J* = 8.8 Hz, 1H, -Ar-H) 7.70 – 7.72 (m, 1H, -Ar-H); 8.08 (d, *J* = 2.4 Hz, 1H, -Ar-H); 8.23 (s, 1H, -C₄-H of coumarin); 12.98 (s, 1H, -NH); *Anal.* Calcd for C₁₇H₁₂ClN₃O₅S: C, 50.31; H, 2.98; N, 10.35; s, 7.90; Found: C, 50.37; H,

2.93; N, 10.31; S, 7.84; HRMS (ESI) m/z calculated for $C_{17}H_{12}ClN_3NaO_5S$ $[M + Na]^+$: 428.0084, found: 428.0083.

2.2.3. (Z)-methyl 2-((Z)-2-((E)-(1-(6,8-dichloro-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-3):

Yield: (86%); m.p.: 265 – 267 °C; IR (KBr, ν cm^{-1}): 3152 (-NH), 1725(lactone -C=O), 1614 (ester -C=O); 1H NMR (400MHz, DMSO- d_6 , δ ppm): 2.36 (s, 3H, -CH₃); 3.76 (s, 3H, -OCH₃), 6.66 (s, 1H, -C=H); 7.70 (t, J = 6.4 Hz, 1H, -Ar-H); 8.08 (s, 1H, -Ar-H); 8.23 (s, 1H, -C₄-H of coumarin); 12.99 (s, 1H, -NH); *Anal.* Calcd for $C_{17}H_{11}Cl_2N_3O_5S$: C, 46.38; H, 2.52; N, 9.54; S, 7.28; Found: C, 46.32; H, 2.58; N, 9.59; S, 7.22; MS (ES m/z): 441 $[M + H]^+$.

2.2.4. (Z)-methyl 2-((Z)-2-((E)-(1-(6-bromo-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-4):

Yield: (92%); m.p.: 276 – 278 °C; IR (KBr, ν cm^{-1}): 3132 (-NH), 1708 (lactone-C=O), 1708 (ester -C=O), 1602 (amide-C=O); 1H NMR (400 MHz, DMSO- d_6 , δ ppm): 2.35 (s, 3H, -CH₃); 3.77 (s, 3H, -OCH₃); 6.93 (s, 1H, -C=H); 7.42 – 7.45 (m, 1H, -Ar-H); 7.81 – 7.84 (m, 1H, -Ar-H); 8.16 (d, J = 2.4Hz, 1H, -Ar-H); 8.23 (s, 1H, -C₄-H of coumarin); 12.97 (s, 1H, -NH); ^{13}C NMR (100 MHz, DMSO- d_6 , ppm): δ 17.1; 52.4; 116.3; 118.3; 120.6; 122.5; 131.3; 135.0; 140.7; 152.6; 171.9; *Anal.* Calcd for $C_{17}H_{12}BrN_3O_5S$: C, 45.35; H, 2.69; N, 9.33; S, 7.12; Found: C, 45.39; H, 2.74; N, 9.38; S, 7.16; HRMS (ESI) m/z calculated for $C_{17}H_{12}BrN_3NaO_5S$ $[M + Na]^+$: 471.9579, found: 471.9575.

2.2.5. (Z)-methyl 2-((Z)-2-((E)-(1-(6,8-dibromo-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-5):

Yield: (90%); m.p.: 248 – 250 °C; IR (KBr, ν cm^{-1}): 3150 (-NH), 1728 (lactone -C=O), 1614 (amide -C=O); 1H NMR (400 MHz, DMSO- d_6 , δ ppm): 2.36 (s, 3H, -CH₃); 3.77 (s, 3H, -OCH₃); 6.68 (s, 1H, -C=H); 7.45 (s, 1H, -Ar-H); 7.83 (s, 1H, -Ar-H); 8.23 (s, 1H, -C₄-H of coumarin); 12.99 (s, 1H, -NH); *Anal.* Calcd for $C_{17}H_{11}Br_2N_3O_5S$: C, 38.59; H, 2.10; N, 7.94; S, 6.06; Found: C, 38.63; H, 2.15; N, 7.90; S, 6.17; MS (ESI m/z): 530 $[M + H]^+$.

2.2.6. (Z)-methyl 2-((Z)-4-oxo-2-((E)-(1-(3-oxo-3H-benzo[f]chromen-2-yl) ethylidene)hydrazono)thiazolidin-5-ylidene)acetate (IV-6):

Yield: (91%); m.p.: 312 - 314 °C; IR (KBr, ν cm^{-1}): 3138 (-NH), 1717 (lactone -C=O), 1617 (amide -C=O), 1570 (-C=N); 1H NMR (400 MHz, DMSO- d_6 , δ ppm): 2.31(s, 3H, -CH₃);

3.71 (s, 3H, -OCH₃); 7.65 – 7.67 (m, 2H, -Ar-H); 7.77 (t, *J* = 7.6 Hz, 2H, -Ar-H); 8.08 (d, *J* = 8.8 Hz, 1H, -Ar-H); 8.23 (d, *J* = 9.2 Hz, 1H, -Ar-H); 8.43 (s, 1H, -Ar-H); 9.11 (s, 1H-C₄-H of coumarin); 10.43 (s, 1H, -NH); *Anal.* Calcd. for C₂₁H₁₅N₃O₅S: C, 59.85; H, 3.59; N, 9.97; S, 7.61; Found: C, 59.80; H, 3.64; N, 9.91; S, 7.65; MS (ES *m/z*): 422 [M+H]⁺.

2.2.7. (Z)-methyl 2-((Z)-2-((E)-(1-(8-methoxy-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-7):

Yield: (79%); m.p.: 260 – 262 °C; IR (KBr, ν cm⁻¹): 3155 (-NH), 1705 (lactone -C=O), 1611 (amide -C=O), 1523 (-C=N); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 2.37 (s, 3H, -CH₃); 3.76 (s, 3H, ester-OCH₃); 3.93 (s, 3H, -OCH₃); 6.68 (s, 1H, =CH); 7.31 – 7.38 (m, 2H, -Ar-H); 7.44 – 7.46 (m, 1H, -Ar-H); 8.24 (s, 1H, C₄-H of coumarin); 12.97 (s, 1H, -NH); ¹³C NMR (100 MHz, DMSO-*d*₆, ppm): δ 17.1; 52.4; 56.1; 114.5; 114.9; 119.2; 120.4; 122.4; 124.7; 126.1; 142.2; 142.9; 146.3; 158.5; 165.8; *Anal.* Calcd for C₁₈H₁₅N₃O₆S: C, 53.86; H, 3.77; N, 10.47; S, 7.99; Found: C, 53.81; H, 3.72; N, 10.52; S, 7.93; HRMS (ESI) *m/z* calculated for C₁₈H₁₅N₃O₆S [M + H]⁺ : 402.0760, found: 402.0763.

2.2.8. (Z)-methyl 2-((Z)-2-((E)-(1-(6-bromo-8-methoxy-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-8):

Yield: (90%); m.p.: 267 -269 °C; IR (KBr, ν cm⁻¹): 3420 (-NH), 1717 (lactone -C=O), 1634 (ester -C=O), 1587 (-C=N); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 2.36 (s, 3H, -CH₃); 3.77 (s, 3H, - ester -OCH₃); 3.96 (s, 3H, -OCH₃); 6.68 (s, 1H, -C=H); 7.52 (d, *J* = 2Hz, 1H, -Ar-H); 7.74 (d, *J* = 2 Hz, 1H, -Ar-H); 8.20 (s, 1H, -C₄-H of coumarin); 12.98 (s, 1H, -NH); *Anal.* Calcd for C₁₈H₁₄BrN₃O₆S: C, 45.01; H, 2.94; N, 8.75; S, 6.68; Found: C, 45.10; H, 3.00; N, 8.79; S, 6.63; HRMS (ESI) *m/z* calculated for C₁₈H₁₅BrN₃O₆S [M+H]⁺ : 479.9865, found: 479.9870.

2.2.9. (Z)-methyl 2-((Z)-2-((E)-(1-(8-ethoxy-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-9):

Yield: (82%); m.p.: 255 – 257 °C; IR (KBr, ν cm⁻¹): 3150 (-NH), 1711 (lactone -C=O), 1605 (amide -C=O); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 1.42 (t, *J* = 6.8 Hz, 3H, -CH₂CH₃); 2.37 (s, 3H, CH₃); 3.76 (s, 3H, -OCH₃); 4.17 – 4.23 (m, 2H, CH₂); 6.68 (s, 1H, =CH); 7.29 – 7.37 (m, 2H, -Ar-H); 7.43 – 7.45 (m, 1H, -Ar-H); 8.23 (s, 1H, -C₄-H of coumarin); 12.97 (s, 1H, -NH); *Anal.* Calcd for C₁₉H₁₇N₃O₆S: C, 54.93; H, 4.12; N, 10.12; S, 7.72; Found: C, 54.98; H, 4.18; N, 10.16; S, 7.78; MS (ESI *m/z*): 416 [M+H]⁺.

2.2.10. (Z)-methyl 2-((Z)-2-((E)-1-(6-bromo-8-ethoxy-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-10):

Yield: (88%); m.p.: 257 – 259 °C; IR (KBr, ν cm^{-1}): 3141 (-NH), 1740 (lactone -C=O), 1702 (ester -C=O), 1605 (amide -C=O); ^1H NMR (400 MHz, $\text{DMSO}-d_6$, δ ppm): 1.40 (t, J = 6.8 Hz, 3H, -CH₃); 2.36 (s, 3H, -CH₃); 3.77 (s, 3H, -OCH₃); 4.20 – 4.25 (m, 2H, -CH₂); 6.68 (s, 1H, =CH); 7.50 (d, J = 2 Hz, 1H, -Ar-H); 7.73 (d, J = 2 Hz, 1H, -Ar-H); 8.19 (s, 1H, -C₄-H of coumarin); 12.98 (s, 1H, -NH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$, δ ppm): δ 14.4; 17.2; 52.4; 65.1; 114.6; 116.2; 118.2; 120.5; 122.4; 127.2; 140.9; 142.4; 142.6; 146.4; 161.9; 165.8; Anal. Calcd for $\text{C}_{19}\text{H}_{16}\text{BrN}_3\text{O}_6\text{S}$: C, 46.17; H, 3.26; N, 8.50; S, 6.49; Found: C, 46.11; H, 3.21; N, 8.40; S, 6.54; MS (ES m/z): 496 $[\text{M}+2\text{H}]^+$.

2.2.11. Ethyl (Z)-2-((E)-4-oxo-2-(((Z)-1-(2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)thiazolidin-5-ylidene)acetate (IV-11):

Yield: (82%); m.p.: 236 - 238 °C; IR (KBr, ν cm^{-1}): 3247 (-NH); 1738 (lactone -C=O); 1713 (ester -C=O); 1688 (amide -C=O); 1614 (-C=N); 1233 (C-O-C); ^1H NMR (400 MHz, $\text{DMSO}-d_6$, δ ppm): 1.25 (t, J = 6.8 Hz, 3H, -CH₂-CH₃); 2.38 (s, 3H, -CH₃); 4.20 – 4.25 (m, 2H, -CH₂); 6.65 (s, 1H, =CH); 7.39 – 7.48 (m, 2H, -Ar-H); 7.69 (t, J = 7.2 Hz, 1H, -Ar-H); 7.92 (d, J = 6.8 Hz, 1H, -Ar-H); 8.27 (s, 1H, -C₄-H of coumarin); 12.96 (s, 1H, NH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$, ppm): δ 13.9, 17.1, 61.2, 114.6, 115.8, 116.0, 118.5, 123.9, 124.7, 126.1, 129.4, 131.7, 132.7, 141.1, 141.9, 153.5, 158.8, 171.9; Anal. Calcd for $\text{C}_{18}\text{H}_{15}\text{N}_3\text{O}_5\text{S}$: C, 56.10; H, 3.92; N, 10.90; S, 8.32; Found: C, 56.17; H, 3.99; N, 10.95; S, 8.38; HRMS (ESI) m/z calculated for $\text{C}_{18}\text{H}_{15}\text{N}_3\text{NaO}_5\text{S}$ $[\text{M} + \text{Na}]^+$: 408.0630, found: 408.0637.

2.2.12. Ethyl (Z)-2-((E)-2-(((Z)-1-(6-chloro-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-12):

Yield: (86%); m.p.: 256 - 258 °C; IR (KBr, ν cm^{-1}): 3245 (-NH); 1715 (lactone-C=O); 1630 (ester -C=O); 1603 (amide -C=O); 1572 (-C=N); 1286 (-C-O-C); ^1H NMR (400 MHz, $\text{DMSO}-d_6$, δ ppm): 1.26 (t, J = 6.8 Hz, 3H, -CH₂-CH₃); 2.33 (s, 3H, -CH₃); 4.22 – 4.27 (m, 2H, -CH₂); 6.62 (s, 1H, =CH); 7.51 (d, J = 8.8 Hz, 1H, Ar-H); 7.70 – 7.73 (m, 1H, Ar-H); 7.86 (d, J = 2.8 Hz, 1H, Ar-H); 8.24 (s, 1H, C₄-H of coumarin); 12.64 (s, 1H, -NH); ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$, ppm): δ 14.0; 23.0; 61.3; 114.7; 118.3; 119.6; 127.0; 127.7; 128.6; 132.1; 140.4; 142.5; 151.7; 156.7; 159.9; 165.4; Anal. Calcd. for $\text{C}_{18}\text{H}_{14}\text{ClN}_3\text{O}_5\text{S}$: C,

51.49; H, 3.36; N, 10.01; S, 7.64; Found: C, 51.54; H, 3.44; N, 9.95; S, 7.69; MS (ESI m/z): 420 [M + H]⁺ (100%).

2.2.13. Ethyl (Z)-2-((E)-2-(((Z)-1-(6,8-dichloro-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-13):

Yield: (85%); m.p.: 254 - 256 °C; IR (KBr, ν cm⁻¹): 3251 (-NH); 1741 (lactone -C=O); 1697 (ester -C=O); 1607 (amide -C=O); 1567 (-C=N); 1240 (-C-O-C); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 1.25 (t, *J* = 6.4 Hz, 3H, -CH₂CH₃); 2.36 (s, 3H, -CH₃); 4.20 - 4.24 (m, 2H, -CH₂); 6.65 (s, 1H, =CH); 7.86 (d, *J* = 2.4 Hz, 1H, Ar-H); 8.06 (t, *J* = 2.8 Hz, 1H, Ar-H); 8.23 (s, 1H, -C₄-H of coumarin); 12.69 (s, 1H, NH); ¹³C NMR (100 MHz, DMSO-*d*₆, ppm): δ 13.9; 23.0; 61.3; 114.7; 118.2; 119.6; 126.9; 127.7; 128.3; 128.6; 132.0; 140.3; 142.5; 151.7; 156.7; 165.3; *Anal.* Calcd for C₁₈H₁₃Cl₂N₃O₅S: C, 47.59; H, 2.88; N, 9.25; S, 7.06; Found: C, 47.51; H, 2.92; N, 9.19; S, 7.13; MS (ESI m/z): 454 [M]⁺ (100%); 456[M+2H]⁺.

2.2.14. Ethyl (Z)-2-((E)-2-(((Z)-1-(6-bromo-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-14):

Yield: (88%); m.p.: 258 - 260 °C; IR (KBr, ν cm⁻¹): 3165 (-NH); 1728 (lactone-C=O); 1697 (ester -C=O); 1635 (amide -C=O); 1614 (-C=N); 1246.46 (C-O-C); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 1.26 (t, *J* = 6.8 Hz, 3H, -CH₂CH₃); 2.36 (s, 3H, CH₃); 4.18 - 4.27 (m, 2H, -CH₂); 6.63 (s, 1H, =CH); 7.44 (d, *J* = 8.8 Hz, 1H, Ar-H); 7.82 (d, *J* = 9.2 Hz, 1H, Ar-H); 8.00 (t, *J* = 2 Hz, 1H, Ar-H); 8.21 (s, 1H, -C₄-H of coumarin); 12.96 (s, 1H, -NH); *Anal.* Calcd for C₁₈H₁₄BrN₃O₅S: C, 46.56; H, 3.04; N, 9.05; S, 6.91; Found: C, 46.50; H, 3.10; N, 9.12; S, 6.85; MS (ESI m/z): 464 [M]⁺ (100%).

2.2.15. Ethyl (Z)-2-((E)-2-(((Z)-1-(6,8-dibromo-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-15):

Yield: (91%); m.p.: 246 - 248 °C; IR (KBr, ν cm⁻¹): 3249 (-NH); 1741 (lactone-C=O); 1697 (ester -C=O); 1634 (amide -C=O); 1614 (-C=N); 1241 (-C-O-C); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 1.24 (t, *J* = 4.4 Hz, 3H, -CH₃); 2.36 (s, 3H, -CH₃); 4.18 - 4.23 (m, 2H, -CH₂); 6.92 (s, 1H, =CH); 7.44 (d, *J* = 8.8 Hz, 1H, -Ar-H); 7.81- 7.84 (m, 1H, -Ar-H); 8.58 (s, 1H, -C₄-H of coumarin); 12.99 (s, 1H, -NH); *Anal.* Calcd for C₁₈H₁₃Br₂N₃O₅S: C, 39.80; H, 2.41; N, 7.74; S, 5.90; Found: C, 39.74; H, 2.46; N, 7.70; S, 5.95; MS (ESI m/z): 544 [M + H]⁺.

2.2.16. Ethyl (Z)-2-((Z)-4-oxo-2-(((E)-1-(3-oxo-3H-benzo[f]chromen-2-yl)ethylidene)hydrazono)thiazolidin-5-ylidene)acetate (IV-16):

Yield: (88%); m.p.: 278 - 280 °C; IR (KBr, ν cm⁻¹): 3154 (-NH); 1711 (lactone-C=O); 1632 (ester -C=O); 1607 (-amide -C=O); 1565 (-C=N); 1238 (-C-O-C); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 1.24 (t, *J* = 6.8 Hz, 3H, -CH₂CH₃); 2.45 (s, 3H, -CH₃); 4.18 – 4.23 (m, 2H, -CH₂); 6.61 (s, 1H, =CH); 7.63 – 7.68 (m, 2H, -Ar-H); 7.79 (t, *J* = 8 Hz, 1H, -Ar-H); 8.10 (d, *J* = 8 Hz, 1H, -Ar-H); 8.27 (d, *J* = 8.8 Hz, 1H, -Ar-H); 8.55 (d, *J* = 8 Hz, 1H, -Ar-H); 9.00 (s, 1H, -C₄-H of coumarin); 12.97 (s, 1H, -NH); *Anal.* Calcd for C₂₂H₁₇N₃O₅S: C, 60.68; H, 3.93; N, 9.65; S, 7.36; Found: C, 60.62; H, 3.97; N, 9.61; S, 7.40; MS (ESI *m/z*): 436 [M + H]⁺.

2.2.17. Ethyl (Z)-2-((E)-2-(((Z)-1-(8-methoxy-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-17):

Yield: (80%); m.p.: 252 -254 °C; IR (KBr, ν cm⁻¹): 3436 (-NH); 1708 (lactone-C=O); 1643 (ester -C=O); 1614 (amide -C=O); 1578 (-C=N); 1235 (-C-O-C); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 1.24 (t, *J* = 6.8 Hz, 3H, -CH₂CH₃); 2.37 (s, 3H, -CH₃); 3.93 (s, 3H, -OCH₃); 4.20 – 4.26 (m, 2H, -CH₂); 6.65 (s, 1H, =CH); 7.26 – 7.28 (m, 1H, Ar-H); 7.33 – 7.36 (m, 1H, Ar-H); 7.44 – 7.46 (m, 1H, Ar-H); 8.24 (s, 1H, -C₄-H of coumarin); 12.95 (s, 1H, -NH); ¹³C NMR (100 MHz, DMSO-*d*₆, δ ppm): 13.9; 17.2; 56.2; 61.3; 114.7; 115.0; 118.8; 119.2; 119.9; 120.5; 124.7; 124.8; 126.0; 126.2; 142.2; 142.4; 146.3; 158.5; 165.3; *Anal.* Calcd for C₁₉H₁₇N₃O₆S: C, 54.93; H, 4.12; N, 10.12; S, 7.72; Found: C, 54.91; H, 4.16; N, 10.18; S, 7.75; MS (ESI *m/z*): 416 [M + H]⁺ (100%).

2.2.18. Ethyl (Z)-2-((E)-2-(((Z)-1-(6-bromo-8-methoxy-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-18):

Yield: (83%); m.p.: 264 - 266 °C; IR (KBr, ν cm⁻¹): 3081 (-NH); 1724 (lactone-C=O); 1632 (ester -C=O); 1618 (amide -C=O); 1600 (-C=N); 1235 (-C-O-C); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 1.25 (t, *J* = 6.8 Hz, 3H, -CH₃); 2.36 (s, 3H, -CH₃); 3.96 (s, 3H, -OCH₃); 4.20 – 4.25 (m, 2H, -CH₂); 6.65 (s, 1H, =CH); 7.52 (d, *J* = 2.4 Hz, 1H, -Ar-H); 7.74 (d, *J* = 2 Hz, 1H, -Ar-H); 8.20 (s, 1H, -C₄-H of coumarin); 12.97 (s, 1H, -NH); *Anal.* Calcd for C₁₉H₁₆BrN₃O₆S: C, 46.17; H, 3.26; N, 8.50; S, 6.49; Found: C, 46.13; H, 3.21; N, 8.59, S, 6.42; MS (ESI *m/z*): 496 [M + 2H]⁺.

**2.2.19. Ethyl (Z)-2-((E)-2-(((Z)-1-(8-ethoxy-2-oxo-2H-chromen-3-yl)ethylidene)
hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-19):**

Yield: (82%); m.p.: 244 - 246 °C; IR (KBr, ν cm⁻¹): 3426 (-NH); 1735 (lactone-C=O); 1708 (ester -C=O); 1696 (amide -C=O); 1607 (-C=N); 1240 (-C-O-C); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 1.25 (t, *J* = 7.2 Hz, 3H, -CH₂-CH₃); 1.42 (t, *J* = 7.2 Hz, 3H, -CH₂CH₃); 2.37 (s, 3H, -CH₃); 4.17 – 4.25 (m, 4H, -CH₂), 6.65 (s, 1H, =CH); 7.29 – 7.35 (m, 2H, -Ar-H); 7.37 – 7.45 (m, 1H, Ar-H); 8.23 (s, 1H, -C4-H of coumarin); 12.96 (s, 1H, -NH); ¹³C NMR (100 MHz, DMSO-*d*₆, δ ppm): 13.9; 14.6; 17.2; 61.3; 64.5; 114.7; 115.9; 119.3; 120.5; 124.8; 126.2; 142.2; 142.7; 143.0; 145.5; 158.6; 165.4; *Anal.* Calcd for C₂₀H₁₉N₃O₆S: C, 55.94; H, 4.46; N, 9.78; S, 7.47; Found: C, 55.90; H, 4.49; N, 9.71; S, 7.41; MS (ESI *m/z*): 452 [M + Na]⁺(100%).

**2.2.20. Ethyl (Z)-2-((E)-2-(((Z)-1-(6-bromo-8-ethoxy-2-oxo-2H-chromen-3-yl)ethylidene)
hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-20):**

Yield: (86%); m.p.: 236 - 238 °C; IR (KBr, ν cm⁻¹): 3251 (-NH); 1726 (lactone -C=O); 1709 (ester-C=O); 1613 (amide -C=O); 1567 (-C=N); 1237 (-C-O-C); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 1.25 (t, *J* = 7.2 Hz, 3H, -CH₂-CH₃); 1.40 (t, *J* = 6.8 Hz, 3H, -CH₂-CH₃); 2.36 (s, 3H, -CH₃); 4.00 – 4.03 (m, 2H, -O-CH₂); 4.21 – 4.25 (m, 2H, -O-CH₂); 6.69 (s, 1H, -C=H); 7.50 (d, *J* = 2 Hz, 1H, -Ar-H); 7.73 (d, *J* = 2 Hz, 1H, -Ar-H); 8.19 (s, 1H, -C₄-H of coumarin); 12.97 (s, 1H, -NH); *Anal.* Calcd for C₂₀H₁₈BrN₃O₆S: C, 47.25; H, 3.57; N, 8.27; S, 6.31; Found: C, 47.21; H, 3.51; N, 8.31; S, 6.38; MS (ESI *m/z*): 510 [M + 2H]⁺.

**2.2.21. Ethyl (Z)-2-((E)-2-(((Z)-1-(7-hydroxy-2-oxo-2H-chromen-3-yl)ethylidene)
hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-21):**

Yield: (82%); m.p.: 260 – 262 °C; IR (KBr, ν cm⁻¹): 3212 (-OH); 3162 (-NH); 1728 (lactone-C=O); 1693 (ester -C=O); 1609 (amide -C=O); 1216 (C-O-C); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 1.25 (t, *J* = 7.2 Hz, 3H, -CH₂CH₃); 2.36 (s, 3H, -CH₃); 4.20 – 4.25 (m, 2H, -CH₂); 6.64 (s, 1H, =CH); 6.76 (d, *J* = 2.4 Hz, 1H, -Ar-H); 6.82 – 6.85 (m, 1H, -Ar-H); 7.73 (d, *J* = 8.8 Hz, 1H, -Ar-H); 8.18 (s, 1H, -C₄-H of coumarin); 10.83 (s, 1H, -OH); 12.91 (s, 1H, -NH); ¹³C NMR (100 MHz, DMSO-*d*₆, ppm): 14.4; 17.7; 61.8; 110.9; 111.6; 113.1; 115.0; 120.9; 128.2; 143.2; 143.5; 154.2; 159.8; 160.2; 160.7; 162.8; 165.8; 166.1; *Anal.* Calcd for C₁₈H₁₅N₃O₆S: C, 53.86; H, 3.77; N, 10.47; S, 7.99; Found: C, 53.81; H, 3.72; N, 10.51; S, 7.93; MS (ESI *m/z*): 402 [M + H]⁺.

2.2.22. Ethyl (Z)-2-((E)-2-(((Z)-1-(7-hydroxy-8-methyl-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-22):

Yield: (77%); m.p.: 273 – 275 °C; IR (KBr, ν cm^{-1}): 3315 (-OH); 3175 (-NH); 1730 (lactone -C=O); 1678 (ester -C=O); 1633 (amide -C=O); 1605 (-C=N); 1230 (-C-O-C); ^1H NMR (400 MHz, $\text{DMSO-}d_6$, δ ppm): 1.25 (t, J = 6.8 Hz, 3H, -CH₂-CH₃); 2.18 (s, 3H, -Ar-CH₃); 2.37 (s, 3H, -CH₃); 4.20 – 4.25 (m, 2H, -CH₂); 6.64 (s, 1H, =CH); 6.89 (d, J = 8.4 Hz, 1H, -Ar-H); 7.57 (d, J = 8.4 Hz, 1H, -Ar-H); 8.16 (s, 1H, -C₄-H of coumarin); 10.75 (s, 1H, -OH); 12.92 (s, 1H, -NH); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$, ppm): δ 7.8; 13.9; 17.2; 61.3; 110.5; 111.1; 112.6; 114.5; 120.5; 127.8; 142.8; 142.9; 153.7; 159.3; 159.7; 160.2; 162.3; 165.3; 165.7; Anal. Calcd for C₁₉H₁₇N₃O₆S: C, 54.93; H, 4.12; N, 10.12; S, 7.72; Found: C, 54.98; H, 4.16; N, 10.08; S, 7.71; MS (ESI m/z): 416 [M+ H]⁺.

2.2.23. tert-butyl (Z)-2-((E)-4-oxo-2-(((Z)-1-(2-oxo-2H-chromen-3-yl) ethylidene)hydrazono) thiazolidin -5-ylidene)acetate (IV-23):

Yield: (78%); m.p.: 240 - 242 °C; IR (KBr, ν cm^{-1}): 3122 (-NH); 1721 (lactone -C=O); 1685 (ester -C=O); 1637 (amide -C=O); 1602 (-C=N); ^1H NMR (400 MHz, $\text{DMSO-}d_6$, δ ppm): 1.47 (s, 9H, *tert.* butyl); 2.38 (s, 3H, -CH₃); 6.54 (s, 1H, =CH); 7.39 – 7.43 (m, 1H, -Ar-H); 7.47 (d, J = 8.4 Hz, 1H, -Ar-H); 7.66 – 7.71 (m, 1H, -Ar-H); 7.90 – 7.93 (m, 1H, -Ar-H); 8.28 (s, 1H, C₄-H of coumarin); 12.91 (s, 1H, -NH); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$, δ ppm): 17.1; 27.6; 82.1; 116.0; 116.5; 118.6; 124.8; 126.0; 129.5; 132.8; 141.7; 142.0; 153.6; 158.8; 160.7; 161.8; 164.6; 165.7; Anal. Calcd for C₂₀H₁₉N₃O₅S: C, 58.10; H, 4.63; N, 10.16; S, 7.76; Found: C, 58.16; H, 4.60; N, 10.11; S, 7.71; HRMS (ESI) m/z calculated for C₂₀H₁₉N₃NaO₅S[M + Na]⁺: 436.0943, found: 436.0942.

2.2.24. tert-butyl (Z)-2-((E)-2-(((Z)-1-(6-chloro-2-oxo-2H-chromen-3-yl) ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-24):

Yield: (85%); m.p.: 272 - 274 °C; IR (KBr, ν cm^{-1}): 3232 (-NH), 1728 (lactone -C=O), 1605 (-amide -C=O); ^1H NMR (400 MHz, $\text{DMSO-}d_6$, ppm): 1.47 (s, 9H, *tert.* butyl); 2.36 (s, 3H, -CH₃); 6.55 (s, 1H, =CH); 7.50 (d, J = 8.8 Hz, 1H, -Ar- H); 7.70 – 7.72 (m, 1H, -Ar-H); 8.07 (d, J = 2.4 Hz, 1H, -Ar-H); 8.24 (s, 1H, C₄-H of coumarin); 12.92 (s, 1H, -NH); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$, ppm): δ 17.1; 27.6; 82.1; 116.4; 118.0; 120.0; 127.2; 128.4; 128.4; 132.2; 140.7; 152.2; 158.4; 161.6; 164.6; Anal. Calcd for C₂₀H₁₈ClN₃O₅S: C, 53.63; H, 4.05; N, 9.38; S, 7.16; found: C, 53.70; H, 4.14; N, 9.30; S, 7.20; MS (ESI m/z): 470 [M + Na]⁺ (100%).

2.2.25. *tert*-butyl (Z)-2-((E)-2-(((Z)-1-(6,8-dichloro-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-25):

Yield: (85%); m.p.: 269 - 270 °C; IR (KBr, ν cm^{-1}): 3141 (-NH); 1732 (lactone -C=O); 1675 (ester -C=O); 1621 (amide -C=O); 1601 (-C=N); ^1H NMR (400 MHz, DMSO- d_6 , δ ppm): 1.47 (s, 9H, *tert.* butyl); 2.36 (s, 3H, -CH₃); 6.55 (s, 1H, =CH); 7.50 (d, J = Hz, 1H, - Ar-H); 7.70 – 7.73 (m, 1H, -Ar-H); 8.07 (d, J = 2.4Hz, 1H, -Ar-H); 8.24 (s, 1H, C₄-H of coumarin); 8.29 (s, 1H, -Ar-H); 12.92 (s, 1H, -NH). *Anal.* Calcd for C₂₀H₁₇Cl₂N₃O₅S: C, 49.80; H, 3.55; N, 8.71; S, 6.65; Found: C, 49.86; H, 3.59; N, 8.77; S, 6.54; MS (ES m/z): 483 [M+H]⁺.

2.2.26. *tert*-butyl (Z)-2-((E)-2-(((Z)-1-(6-bromo-2-oxo-2H-chromen-3-yl) ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-26):

Yield: (84%); m.p.: 274 – 276 °C; IR (KBr, ν cm^{-1}): 3202 (-NH); 1722 (lactone -C=O); 1605 (-C=N); 1241 (-C-O-C); ^1H NMR (400 MHz, DMSO- d_6 , δ ppm): 1.47 (s, 9H, *tert.* butyl); 2.36 (s, 3H, -CH₃); 6.55 (s, 1H, =CH); 7.44 (d, J = 8.8 Hz, 1H, -Ar-H); 7.81 – 7.84 (m, 1H, - Ar-H); 8.20 (d, J = 2.4 Hz, 1H); 8.24 (s, 1H, C₄-H of coumarin); 12.92 (s, 1H, -NH); ^{13}C NMR (100 MHz, DMSO- d_6 , ppm): δ 17.1; 27.6; 82.1; 115.9; 116.4; 124.7; 125.9; 129.4; 132.7; 141.6; 141.9; 153.5; 158.7; 160.7; 161.8; 164.5; 165.6; *Anal.* Calcd for C₂₀H₁₈BrN₃O₅S: C, 48.79; H, 3.69; N, 8.53; S, 6.51; Found: C, 48.84; H, 3.61; N, 8.59; S, 6.54; MS (ESI m/z): 494 [M + 2H]⁺.

2.2.27. *tert*-butyl (Z)-2-((E)-2-(((Z)-1-(6,8-dibromo-2-oxo-2H-chromen-3-yl)ethylidene)hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-27):

Yield : (88%); m.p.: 242 - 244 °C; IR (KBr, ν cm^{-1}): 3064 (-NH); 1725 (lactone -C=O); 1617 (amide -C=O); 1235 (-C-O-C); ^1H NMR (400 MHz, DMSO- d_6 , ppm): 1.47 (s, 9H, - *tert.* butyl); 2.36 (s, 3H, -CH₃); 6.54 (s, 1H, =CH); 7.44 (d, J = 8.8 Hz, 1H, -Ar-H); 7.81 – 7.83 (m, 1H, -Ar-H); 8.20 (s, 1H, -C₄-H of coumarin); 12.92 (s, 1H, -NH); *Anal.* Calcd for C₂₀H₁₇Br₂N₃O₅S: C, 42.05; H, 3.00; N, 7.36; S, 5.61; Found: C, 42.10; H, 2.96; N, 7.32; S, 5.66; MS (ES m/z): 572 [M+H]⁺.

2.2.28. *tert*-butyl (Z)-2-((E)-4-oxo-2-(((Z)-1-(3-oxo-3H-benzol[f]chromen-2-yl)ethylidene)hydrazono)thiazolidin-5-ylidene)acetate (IV-28):

Yield: (80%); m.p.: 318 – 320 °C; IR (KBr, ν cm^{-1}): 3085 (-NH); 1720 (lactone -C=O); 1626 (ester -C=O); 1612 (amide -C=O); 1238 (-C-O-C); ^1H NMR (400 MHz, DMSO- d_6 , δ ppm): 1.46 (s, 9H, *tert.* butyl- H); 2.45 (s, 3H, -CH₃); 6.54 (s, 1H, =CH); 7.64 – 7.69 (m, 2H, -Ar-

H); 7.78 (t, $J = 8$ Hz, 1H, -Ar-H); 8.11 (d, $J = 8.4$ Hz, 1H, -Ar-H); 8.28 (d, $J = 8.8$ Hz, 1H, -Ar-H); 8.56 (d, $J = 8.4$ Hz, 1H, -Ar-H); 9.01 (s, 1H, C₄-H of coumarin); 12.95 (s, 1H, =NH); *Anal.* Calcd for C₂₄H₂₁N₃O₅S: C, 62.19; H, 4.57; N, 9.07; S, 6.92; Found: C, 62.14; H, 4.53; N, 8.97; S, 6.96; MS (ES m/z): 464 [M+H]⁺.

2.2.29. *tert-butyl(Z)-2-((E)-2-(((Z)-1-(8-methoxy-2-oxo-2H-chromen-3-yl) ethylidene) hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-29):*

Yield: (84%); m.p.: 138 - 140 °C; IR (KBr, ν cm⁻¹): 3215(-NH); 1722 (-lactone – C=O); 1614 (amide – C=O); 1247 (-C-O-C); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 1.47 (s, 9H, *tert.* butyl - H); 2.37 (s, 3H, -CH₃); 3.93 (s, -3H, -OCH₃); 6.54 (s, 1H, =CH); 7.31 – 7.38 (m, 2H, -Ar-H); 7.43 – 7.45 (m, 1H, -Ar-H); 8.29 (s, 1H, C₄-H of coumarin); 12.91 (s, 1H, -NH); ¹³C NMR (100 MHz, DMSO-*d*₆, ppm): 17.7; 28.1; 56.7; 82.7; 115.5; 117.0; 119.7; 121.1; 125.3; 126.7; 142.2; 142.3; 143.5; 146.8; 159.1; 162.3; 165.1; 166.3; *Anal.* Calcd for C₂₁H₂₁N₃O₆S: C, 56.87; H, 4.77; N, 9.48, S, 7.23; Found: C, 56.82; H, 4.79; N, 9.42; S, 7.26; MS (ESI m/z): 444 [M + H]⁺.

2.2.30. *tert-butyl(Z)-2-((E)-2-(((Z)-1-(6-bromo-8-methoxy-2-oxo-2H-chromen-3-yl) ethylidene) hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-30):*

Yield: (85%); m.p.: 249 – 250 °C; IR (KBr, ν cm⁻¹): 3196 (-NH); 1734 (lactone-C=O); 1685 (ester –C=O); 1627 (amide –C=O); 1603 (-C=N); ¹H NMR (400 MHz, DMSO-*d*₆, δ ppm): 1.47 (s, 9H, *tert.*butyl-H), 2.35 (s, 3H, -CH₃); 3.95 (s, 3H, -OCH₃); 7.52 (d, $J = 2$ Hz, 1H, -Ar-H); 7.74 (d, $J = 2$ Hz, 1H, -Ar-H); 8.21 (s, 1H, -C₄-H of coumarin); 12.91 (s, 1H, -NH); ¹³C NMR (100 MHz, DMSO-*d*₆, ppm): δ 17.1; 27.6; 56.7; 82.1; 116.1; 116.5; 117.5; 120.4; 122.4; 127.2; 140.9; 141.7; 142.3; 147.1; 158.0; 161.0; 161.5; 164.6; 165.7; *Anal.* Calcd for C₂₁H₂₀BrN₃O₆S: C, 48.28; H, 3.86; N, 8.04; S, 6.14; Found: C, 48.32; H, 3.81; N, 8.12; S, 6.18; MS (ES m/z): 523 [M + H]⁺.

2.2.31. *tert-butyl (Z)-2-((E)-2-(((Z)-1-(8-ethoxy-2-oxo-2H-chromen-3-yl) ethylidene) hydrazono)-4-oxothiazolidin-5-ylidene)acetate (IV-31):*

Yield: (81%); m.p.: 225 - 227 °C; IR (KBr, ν cm⁻¹): 3147 (-NH); 1717 (lactone –C=O); 1608 (-C=N); 1250 (-C-O-C); ¹H NMR (400 MHz, DMSO- *d*₆, δ ppm): 1.42 (t, $J = 6.8$ Hz, 3H, -CH₃); 1.47 (s, 9H, *tert.* butyl –H); 2.37 (s, 3H, -CH₃); 4.17 – 4.22 (m, 2H, -O-CH₂); 6.54 (s, 1H, =CH); 7.29 – 7.37 (m, 2H, -Ar-H); 7.43 (d, $J = 7.2$ Hz, 1H, -Ar-H); 8.24 (s, 1H, C₄-H of coumarin); 12.91 (s, 1H,=NH); ¹³C NMR (100 MHz, DMSO-*d*₆, ppm): δ 14.6; 17.2; 27.6;

64.5; 82.2; 115.9; 116.5; 119.3; 120.5; 124.8; 126.1; 141.7; 142.3; 143.0; 145.5; 158.6; 160.8; 161.9; 164.6; 165.7; *Anal.* Calcd for C₂₂H₂₃N₃O₆S: C, 57.76; H, 5.07; N, 9.18; S, 7.01; Found: C, 57.79; H, 4.98; N, 9.24; S, 6.97; HRMS (ESI m/z calculated for C₂₂H₂₃N₃NaO₆S [M + Na]⁺ : 480.1205, found: 480.1204.

2.3. Antiviral activity evaluation:

The antiviral activity of the new compounds was determined in different cell types. (i) HEL cells were seeded in 96-well plates and incubated for 6 days at 37°C until confluency was reached. Medium was aspirated and replaced by serial dilutions of the test compounds (100 µl per well). One hundred microliters of the virus (*Herpes simplex virus type 1*, *Herpes simplex virus type 2*, *Herpes simplex virus type 1 TK- ACV*, *Vaccinia virus*, *Adeno virus-2* or *Vesicular stomatitis virus*), diluted in medium to obtain a virus input of 100 CCID₅₀ (1 CCID₅₀ being the virus dose that is able to infect 50% of the cell cultures), was added to each well. Mock-treated cell cultures receiving solely the test compounds were included, to determine the cytotoxicity. After 4 days of incubation at 37°C, microscopical analysis was performed to score the virus-induced cytopathicity. (ii) Vero cells were seeded in 96-well plates at 30,000 cells per well and incubated for 1 day at 37°C until confluency was reached. Medium was aspirated and replaced by serial dilutions of the test compounds (100 µl per well). One hundred microliters of the virus (*Coxsackie virus B4*, *Sindbis virus*, *Parainfluenza virus 3*, *Punta Toro virus* or *Reovirus*) diluted in medium to obtain a virus input of 100 CCID₅₀ was added to each well. Mock-treated cell cultures receiving solely the test compounds were included, to determine the cytotoxicity. After 3 days (for Coxsackie virus B4 and Sindbis virus) or 6 days (for Parainfluenza virus 3, Punta Toro virus and Reovirus) of incubation at 37°C microscopical analysis was performed to score the virus-induced cytopathicity. (iii) HeLa cells were seeded in 96-well plates at 15,000 cells per well and incubated for 1 day at 37°C until confluency was reached. Medium was aspirated and replaced by serial dilutions of the test compounds (100 µl per well). One hundred microliters of the virus (*Coxsackie virus B4*, *Vesicular stomatitis virus* or *Respiratory syncytial virus*) diluted in medium to obtain a virus input of 100 CCID₅₀ was added to each well. Mock-treated cell cultures receiving solely the test compounds were included, to determine the cytotoxicity. After 3 days (for Coxsackie virus B4 and Vesicular stomatitis virus) or 6 days (for Respiratory syncytial virus) of incubation at 37°C microscopical analysis was performed to score the virus-induced cytopathicity. (iv) MDCK cells were seeded in 96-well plates at 7,500 cells per well and incubated for 1 day at 35°C. Medium was aspirated and replaced by

serial dilutions of the test compounds (100 µl per well). One hundred microliters of the virus (*Influenza A/H1N1 A/Ned/378/05*, *Influenza A/H3N2 A/HK/7/87* or *Influenza B B/Ned/537/05*) diluted in medium to obtain a virus input of 100 CCID₅₀ was added to each well. Mock-treated cell cultures receiving solely the test compounds were included, to determine the cytotoxicity. After 4 days of incubation at 35°C the virus-induced cytopathicity was determined by visual scoring of the CPE, as well as by measuring the cell viability with the colorimetric formazan-based MTS assay. (v) CRFK cells were seeded in 96-well plates at 30,000 cells per well and incubated for 1 day at 37°C until confluency was reached. Medium was aspirated and replaced by serial dilutions of the test compounds (100 µl per well). One hundred microliters of the virus (*Feline herpes virus* or *Feline corona virus*) diluted in medium to obtain a virus input of 100 CCID₅₀ was added to each well. Mock-treated cell cultures receiving solely the test compounds were included, to determine the cytotoxicity. After 4 days of incubation at 37°C the virus-induced cytopathicity was determined by measuring the cell viability with the colorimetric formazan-based MTS assay.

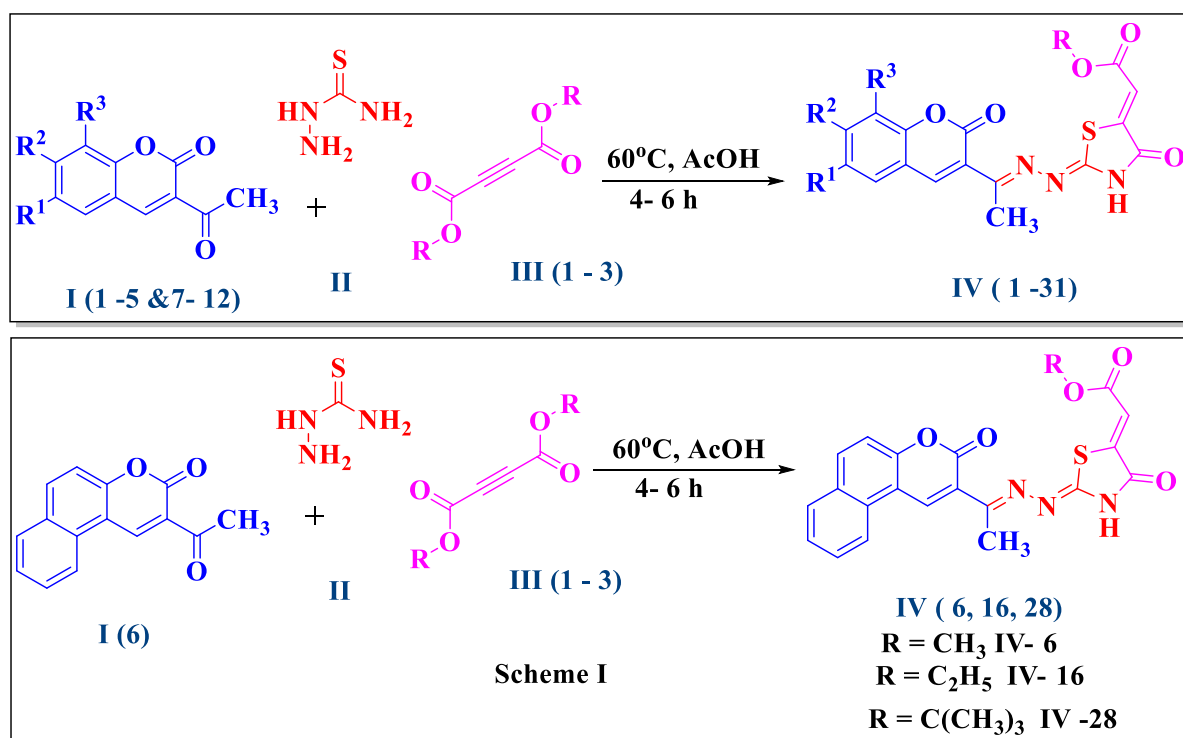
3. Results and discussion

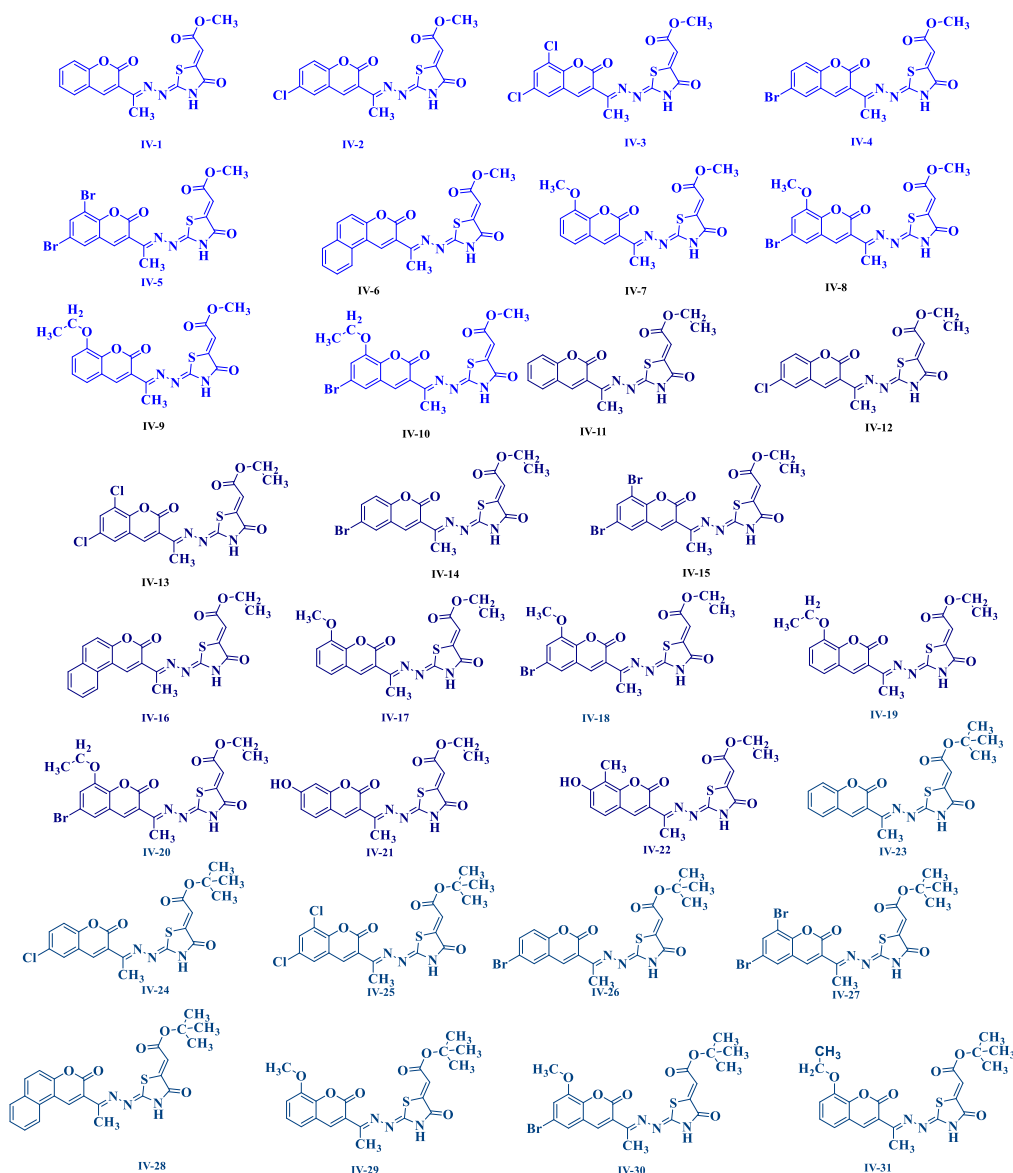
3.1. Chemistry

A proficient and novel synthesis of alky 4-oxo-coumarinyl ethylidene hydrazono- thiazolidin-5-ylidene acetate derivatives were obtained via a one pot multicomponent approach. The schematic representation of the target molecules is depicted in **Scheme 1**. The starting materials of substituted 3-acetyl coumarins were synthesised using various substituted salicylaldehydes and ethylacetoacetate by Knoevenegel condensation method [38, 39]. Initially we performed the reaction in presence of ethanol with substituted 3-acetylcoumarin, thiosemicarbazide and dialkyl acetylene dicarboxylate. The reaction took place at longer time intervals to obtain the desired product, whereas in the presence of acetic acid the reaction was completed within 3-4 h with good yields.

All the synthesised compounds were well characterised by physical and analytical spectral data (IR, ¹H NMR, ¹³C NMR, ESI - Mass). The analytical data of the novel substituted 4-oxo-coumarinyl ethylidene hydrazono- thiazolidin-5-ylidene acetate were presented in the experimental section. The IR spectrum of the compound IV-14 exhibited characteristic absorption bands at 3165, 1728, 1697, 1635 and 1614 cm⁻¹ with respect to the –NH, coumarin lactone, ester carbonyl, amide carbonyl and imine (–C=N) functional groups.

Further support was obtained by using ^1H NMR, ^{13}C NMR, mass spectral data and single crystal X-ray data. The ^1H NMR of compound IV-8 exhibited characteristic singlet peaks at δ 2.36, 3.77, 3.96, 6.68, 8.20 and 12.98 ppm corresponding to $-\text{CH}_3$, $-\text{OCH}_3$ of ester, OCH_3 of coumarin ring, vinylic proton ($=\text{C}-\text{H}$), C_4-H of coumarin, and $-\text{NH}$ proton respectively. The ^{13}C NMR exhibited characteristic peaks at δ 13.98, 120.52, 158.72, 162.32, 165.66 ppm for methyl, vinylic, lactone carbonyl, imino and ester carbons respectively. The LC-MS of the compound IV-12 indicated the molecular ion peaks as $[\text{M}+1]$ and $[\text{M}+3]$ at m/z 420.16 and 422.18 a.m.u.





Scheme 1. The synthetic strategy to make the alkyl 4-oxo-coumarinyl ethylidene hydrazono-thiazolidin-5-ylidene acetate derivatives.

Single crystal X-ray data of the compound IV-31 further confirms the structure (**Figure 2a**). In the title compound, the 2*H*-chromene ring system is essentially planar [r.m.s deviation = 0.003 Å] and are twisted with a little angle of 2.70 (10) to the 1,3-thiazolidine ring. The C8---C3---O1---C2, C3---O1---C2---C1, C9---C10---C12---N1, C10---C12---N1---N2, S1---C14---N2---N1, C14---N2---N1---C12, S1---C16---C17---C18 and C16---C17---C18---O2 torsion angles are 8.5(4), - 174.6(3), 5.1(4), -179.9(2), -0.4(4), -172.8(2), 0.9(4) and 166.7(2) respectively. Selected geometric parameters are given in **Table 1**.

In the crystal, there exist two intramolecular C—H...O hydrogen bonds. N—H...O hydrogen bonds link the molecules to each other, forming centro symmetric R^2_2 (8) dimers (**Table 2**, **Figure. 2b**). Furthermore, C---H... π and π - π interactions are not observed.

Table 1: Single crystal X-ray diffraction data of compound IV- 31

Chemical formula	C ₂₂ H ₂₃ N ₃ O ₆ S
M_r	457.49
Crystal system, space group	Monoclinic, $P2_1/c$
Temperature (K)	296
a, b, c (Å)	5.6858 (3), 28.274 (2), 13.7996 (10)
β (°)	97.095 (3)
V (Å ³)	2201.5 (3)
Z	4
$F(000)$	960
D_x (Mg m ⁻³)	1.380
Radiation type	Mo $K\alpha$
No. of reflections for cell measurement	3354
θ range (°) for cell measurement	5.2–45.5
μ (mm ⁻¹)	0.19
Crystal shape	Block
Colour	Yellow
Crystal size (mm)	0.60 × 0.30 × 0.20
Diffractometer	Bruker axs kappa apex2 CCD Diffractometer
Radiation source	fine-focus sealed tube
Monochromator	Graphite
Scan method	ω and ϕ scan
Absorption correction	Multi-scan SADABS (Bruker, 1999)
$T_{\min}, T_{\max}$	0.894, 0.963
No. of measured, independent and observed [$I > 2\sigma(I)$] reflections	17534, 5392, 3321
R_{int}	0.039
θ values (°)	$\theta_{\max} = 28.4, \theta_{\min} = 1.4$
$(\sin \theta/\lambda)_{\max}$ (Å ⁻¹)	0.670
Range of h, k, l	$h = -7 \rightarrow 7, k = -35 \rightarrow 37, l = -18 \rightarrow 18$
Refinement on	F^2
$R[F^2 > 2\sigma(F^2)], wR(F^2), S$	0.054, 0.177, 1.07
No. of reflections	5392
No. of parameters	294
No. of restraints	0
H-atom treatment	H-atom parameters constrained
Weighting scheme	$w = 1/[\sigma^2(F_o^2) + (0.091P)^2 + 0.0395P]$ where $P = (F_o^2 + 2F_c^2)/3$
$(\Delta/\sigma)_{\max}$	< 0.001
$\Delta_{\max}, \Delta_{\min}$ (e Å ⁻³)	0.40, -0.34

Table 2: Hydrogen Bonds (Å)

D—H...A	D—H	H...A	D...A	D—H...A
N3---H3N...O6	0.86	1.98	2.828(3)	170
C21-H21C...O3	0.96	2.55	3.104 (3)	117
C22---H22B...O3	0.96	2.36	2.908 (3)	115

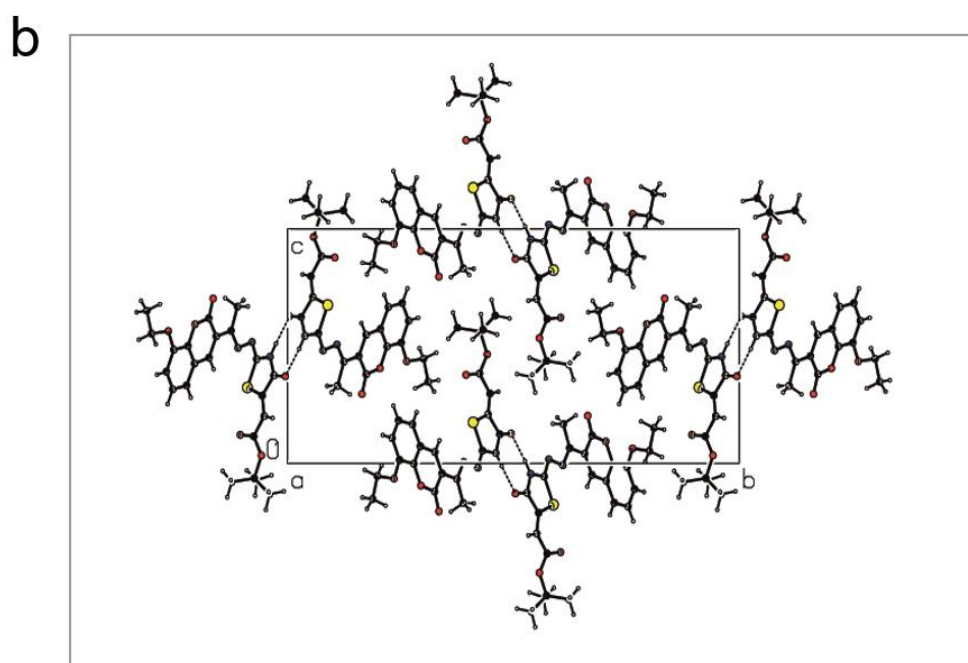
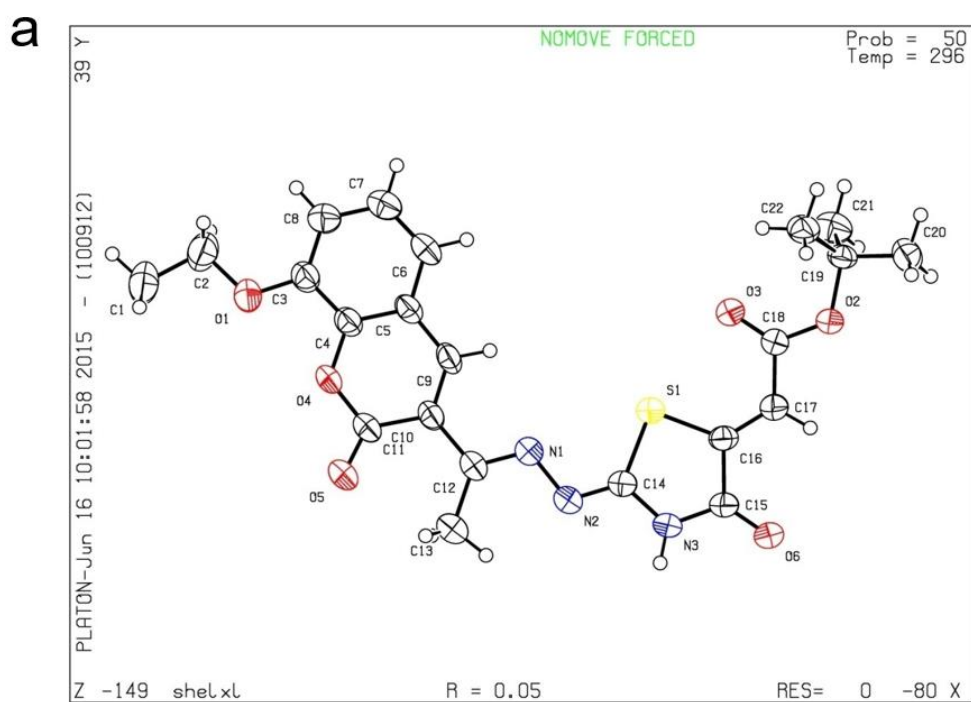


Figure 2. a) The molecular structure of compound IV-31 with the atom labelling (CCDC 418103). b) View of the dimers formed by the pairs of N-H...O hydrogen bonds with R²₂ (8) motifs, down the axis.

3.2. Antiviral activity

The compounds alkyl 4-oxo-(coumarinyl ethylidene hydrazono)-thiazolidin-5-ylidene acetates (IV-1 – IV-31) were evaluated against a wide range of human RNA and DNA viruses in different types of cell cultures. The HEL cell culture was used to evaluate the compounds against herpes simplex virus type I (KOS) [HSV-1 KOS], herpes simplex virus type 2 (G) [HSV-2 G], vaccinia virus [VV], vesicular stomatitis virus (VSV), thymidine kinase-deficient herpes simplex virus type 1 (HSV-1 TK⁻ KOS ACV^r) and adeno virus type 2 (AV-2). Based on the microscopical analysis of the virus-induced cytopathicity (CPE) the antiviral potency of the compounds was determined and compared to different reference antiviral drugs (brivudin, cidofovir, acyclovir, ganciclovir, zalcitabine and alovudine). As summarized in **Table 3**, only one compound (IV-19) showed moderate activity against the tested herpes simplex viruses, with EC₅₀ values of about 10 µM.

Vesicular stomatitis virus (VSV) was also tested in the HeLa cell culture, together with the RNA viruses coxsackie virus B4 (CV B4) and respiratory syncytial virus (RSV). Here, dextran sulphate (DS-10000) and ribavirin were included as reference compounds. **Table 4** shows that compound IV-19, in contrast to HEL cells, inhibited VSV cytopathogenicity of HeLa cells with an EC₅₀ of 9.5 µM. Both for compounds IV-24 and IV-25 antiviral potency was recorded against VSV and RSV in the lower micromolar range (**Table 4**).

Next, Vero cells were utilized to evaluate the compounds against parainfluenza type 3 virus (PI-3V), reovirus type 1 (RV-1), sindbis virus (SV), coxsackie virus B4 (CV B4) and punto toro virus (PTV), together the reference compounds DS-10000 and ribavirin. As depicted in **Table 5**, compound IV-2 had the broadest antiviral spectrum in Vero cell cultures. It was active against 4 of the 5 tested viruses (PI-3V, SV, CV B4 and PTV). Comparable activity was noted for compound IV-3, except for CV B4. Again, compound IV-19 showed antiviral activity against several of the tested viruses in these Vero cells: PI-3V, CV B4 and PTV. A remarkable observation was the activity of the compounds against punto toro virus. For 13 out of the 31 synthesised compounds, a clear inhibition of PTV cytopathogenicity was measured, with EC₅₀ values ranging between 1.9 and 9.5 µM.

Table 5: Cytotoxicity and antiviral activity of compounds in Vero cell cultures.

Compound	Minimum cytotoxic concentration ^a (μ M)	EC ₅₀ ^b (μ M)				
		Para- influenza-3 virus	Reovirus-1	Sindbis virus	Coxsackie virus B4	Punta Toro virus
IV 1	100	>20	>20	>20	>20	>20
IV 2	$\geq$ 100	33	>100	39	39	9.5
IV 3	$\geq$ 100	45	>100	46	>100	8.9
IV 4	100	>20	>20	>20	>20	9.5
IV 5	100	10	>20	>20	>20	>20
IV 6	100	>20	>20	>20	>20	>20
IV 7	$\geq$ 100	>100	>100	>100	>100	>100
IV 8	100	>20	>20	>20	>20	3.2
IV 9	$\geq$ 20	>20	>20	>20	>20	>20
IV 10	100	>20	>20	>20	>20	3.0
IV 11	100	>20	>20	>20	>20	>20
IV 12	100	>20	>20	>20	>20	8.9
IV 13	100	>20	>20	>20	>20	8.4
IV 14	100	>20	>20	>20	>20	7.0
IV 15	100	>20	>20	>20	>20	>20
IV 16	100	>20	>20	>20	>20	>20
IV 17	100	>20	>20	>20	7.0	1.9
IV 18	$\geq$ 20	>20	>20	>20	>20	1.9
IV 19	100	10	>20	>20	7.0	8.3
IV 20	$\geq$ 20	>20	>20	>20	>20	6.2
IV 21	$\geq$ 20	>20	>20	>20	>20	>20
IV 22	$\geq$ 20	>20	>20	>20	>20	>20
IV 23	$\geq$ 20	>20	>20	>20	8.9	>20
IV 24	$\geq$ 20	>20	>20	>20	>20	>20
IV 25	$\geq$ 20	>20	>20	>20	>20	>20
IV 26	>100	>100	>100	>100	>100	>100
IV 27	20	>4	>4	>4	>4	>4
IV 28	$\geq$ 20	>20	>20	>20	>20	>20
IV 29	$\geq$ 20	>20	>20	>20	>20	8.9
IV 30	20	>4	>4	>4	>4	>4
IV 31	$\geq$ 20	>20	9.5	>20	>20	>20
DS-10000 (μ g/ml)	>100	>100	>100	50	8.9	20
Ribavirin	>250	126	>250	250	>250	112

^a Required to cause a microscopically detectable alteration of normal cell morphology.

^b Required to reduce virus-induced cytopathogenicity by 50 %.

Finally, the same compounds were tested for their potency against influenza virus (IV). The MDCK cell line was used to infect with influenza A virus type H1N1 (strain A/Ned/378/05) and type H3N2 (strain A/HK/7/87), and influenza B virus (strain B/Ned/537/05). As reference compounds, zanamivir, ribavirin, amantadine and rimantadine were selected. As

summarized in **Table 6**, only the three compounds IV-7, IV-9 and IV-21 exerted activity against influenza virus with EC₅₀ values in the lower micromolar range, both by determination of the CPE as by measuring the cell viability with the colorimetric formazan-based MTS assay^{40,41}.

Table 6: Cytotoxicity and antiviral activity of compounds in MDCK cell cultures.

Compound	Cytotoxicity (μM)		Antiviral EC ₅₀ ^c (μM)					
	Minimum cytotoxic concentration ^a	CC ₅₀ ^b	Influenza A/H1N1 A/Ned/378/05		Influenza A/H3N2 A/HK/7/87		Influenza B B/Ned/537/05	
			CPE	MTS	CPE	MTS	CPE	MTS
IV 1	100	>100	>20	>20	>20	>20	>20	>20
IV 2	20	8.6	>4	>4	>4	>4	>4	>4
IV 3	20	9.5	>4	>4	>4	>4	>4	>4
IV 4	≥100	>100	>100	>100	>100	>100	>100	>100
IV 5	20	37.7	>4	>4	>4	>4	>4	>4
IV 6	100	>100	>20	>20	>20	>20	>20	>20
IV 7	100	>100	11.0	7.4	6.5	4.1	27.0	13.8
IV 8	20	72.6	>4	>4	>4	>4	>4	>4
IV 9	≥20	39.0	10.5	6.3	7.9	4.2	>20	>20
IV 10	≥20	38.0	>20	>20	>20	>20	>20	>20
IV 11	≥20	40.8	>20	>20	>20	>20	>20	>20
IV 12	100	>100	>20	>20	>20	>20	>20	>20
IV 13	≥20	>100	>20	>20	>20	>20	>20	>20
IV 14	100	57.2	>20	>20	>20	>20	>20	>20
IV 15	20	12.4	>4	>4	>4	>4	>4	>4
IV 16	20	38.8	>4	>4	>4	>4	>4	>4
IV 17	20	43.7	>4	>4	>4	>4	>4	>4
IV 18	20	48.4	>4	>4	>4	>4	>4	>4
IV 19	20	10.1	>4	>4	>4	>4	>4	>4
IV 20	≥4	27.2	>4	>4	>4	>4	>4	>4
IV 21	≥100	>100	20.9	9.1	34	8.7	29.5	24.9
IV 22	100	>100	>20	>20	>20	>20	>20	>20
IV 23	≥4	44.7	>4	>4	>4	>4	>4	>4
IV 24	4	2.1	>0.8	>0.8	>0.8	>0.8	>0.8	>0.8
IV 25	4	8.5	>0.8	>0.8	>0.8	>0.8	>0.8	>0.8
IV 26	>100	>100	>100	>100	>100	>100	>100	>100
IV 27	0.8	0.6	>0.16	>0.16	>0.16	>0.16	>0.16	>0.16
IV 28	0.8	1.8	>0.16	>0.16	>0.16	>0.16	>0.16	>0.16
IV 29	≥4	9.0	>4	>4	>4	>4	>4	>4
IV 30	0.8	1.9	>0.16	>0.16	>0.16	>0.16	>0.16	>0.16
IV 31	20	9.4	>4	>4	>4	>4	>4	>4
Zanamivir	>100	>100	0.2	0.3	0.5	0.6	0.4	0.4
Ribavirin	≥100	>100	8.9	8.2	8.9	5.3	6.8	8.4
Amantadine	>200	>200	8.0	19.1	0.8	1.6	>200	>200
Rimantadine	>200	>200	200	>200	0.3	0.1	>200	>200

^a Minimum compound concentration that causes a microscopically detectable alteration of normal cell morphology.

^b 50% Cytotoxic concentration, as determined by measuring the cell viability with the colorimetric formazan-based MTS assay.

^c 50% Effective concentration, or concentration producing 50% inhibition of virus-induced cytopathic effect, as determined by visual scoring of the CPE, or by measuring the cell viability with the colorimetric formazan-based MTS assay.

Evaluation of these alkyl 4-oxo-coumarinyl ethylidene hydrazono- thiazolidin -5-ylidene acetate derivatives against feline corona virus and feline herpes virus was less successful as none of the compounds showed any detectable activity against these feline viruses (**Table 7**).

The structure activity relationship studies of alkyl 4-oxo-coumarinyl ethylidene hydrazono-thiazolidin -5-ylidene acetate derivatives clearly exhibiting that the presence of substituents on coumarin at 6th and 8th positions and ethoxy ester on thiazolidinone ring are crucial for antiviral activity. Out of twelve ethoxy ester having thiazolidinones, seven compounds exhibited excellent activity against *Punta Toro* virus. The compounds IV-17, IV-18, IV-19 and IV-20 possessed 8-methoxy, 6-bromo 8-methoxy, 8-ethoxy and 6-bromo 8-ethoxy exhibited excellent EC₅₀ values of 1.9, 1.9, 8.3 and 6.2 μM concentrations. It is noteworthy to mention that coumarin having bromo at 6th position and methoxy / ethoxy functional group at 8th position is exhibiting broad spectrum of antiviral activity. Similarly the compounds having halogenated electron withdrawing groups (Cl and Br) were found to be active against Para influenza, Sindbis, Coxsackie, and Punta Toro Viruses.

4. Conclusion

In the search for novel potentially antiviral agents, a one pot synthesis was used to generate alkyl -4-oxo- (coumarinyl ethylidene hydrazono)-thiazolidin-5-ylidene acetate (IV- 1-31) derivatives. Three groups of analogs have been synthesised and evaluated against a broad panel of viruses in different cell lines. It was found that among the tested compounds: (i) IV-19 was the compound that showed antiviral activity against most viruses, both DNA as RNA viruses, (ii) about 1/3 of the analogs showed potency against punta toro virus, and (iii) three compounds were highly selective for influenza virus.

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Conflicts of interest: We declare no conflict of competing or financial interest regarding this manuscript.

Supplementary material associated with this article having analytical spectral data like ¹H NMR, ¹³CNMR and Mass spectral data can be found online....

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Highlights

- One Pot multicomponent synthesis of 4-oxo-coumarinyl ethylidene hydrazono-thiazolidin-5-ylidene acetates.
- Single crystal X-ray data and NMR for structural analysis.
- Antiviral activity against a wide range of human RNA and DNA viruses in different types of cell cultures.
- Compound (IV-19) showed antiviral activity against most viruses, both DNA as RNA viruses, about 1/3 of the analogs showed potency against punta toro virus and three compounds were highly selective for influenza virus.

Table 3: Cytotoxicity and antiviral activity of compounds in HEL cell cultures.

Compound	Minimum cytotoxic concentration ^a (μ M)	EC ₅₀ ^b (μ M)					
		Herpes simplex virus-1 (KOS)	Herpes simplex virus-2 (G)	Vaccinia virus	Vesicular stomatitis virus	Herpes simplex virus-1 TK ⁻ KOS ACV ^r	Adeno virus-2
IV 1	100	>20	>20	>20	>20	>20	>20
IV 2	$\geq$ 100	>100	>100	>100	>100	>100	>100
IV 3	100	>20	>20	>20	>20	>20	>20
IV 4	100	>20	>20	>20	>20	>20	>20
IV 5	100	>20	>20	>20	>20	>20	>20
IV 6	100	>20	>20	>20	>20	>20	>20
IV 7	100	>20	>20	>20	>20	>20	>20
IV 8	100	>20	>20	>20	>20	>20	>20
IV 9	$\geq$ 20	>20	>20	>20	>20	>20	>20
IV 10	20	>4	>4	>4	>4	>4	>4
IV 11	100	>20	>20	>20	>20	>20	>20
IV 12	100	>20	>20	>20	>20	>20	>20
IV 13	100	>20	>20	>20	>20	>20	>20
IV 14	100	>20	>20	>20	>20	>20	>20
IV 15	20	>4	>4	>4	>4	>4	>4
IV 16	100	>20	>20	>20	>20	>20	>20
IV 17	100	>20	>20	>20	>20	>20	>20
IV 18	100	>20	>20	>20	>20	>20	>20
IV 19	100	11	10	>20	>20	10	>20
IV 20	100	>20	>20	>20	>20	>20	>20
IV 21	100	>20	>20	>20	>20	>20	>20
IV 22	$\geq$ 20	>20	>20	>20	>20	>20	>20
IV 23	>100	>100	>100	>100	>100	>100	>100
IV 24	20	>4	>4	>4	>4	>4	>4
IV 25	20	>4	>4	>4	>4	>4	>4
IV 26	>100	>100	>100	>100	>100	>100	>100
IV 27	20	>4	>4	>4	>4	>4	>4
IV 28	$\geq$ 4	>4	>4	>4	>4	>4	>4
IV 29	20	>4	>4	>4	>4	>4	>4
IV 30	$\geq$ 4	>4	>4	>4	>4	>4	>4
IV 31	20	>4	>4	>4	>4	>4	>4
Brivudin	>250	0.05	181	20	>250	150	-
Cidofovir	>250	1.6	1.0	22	>250	1.6	7.9
Acyclovir	>250	0.4	0.2	>250	>250	98	-
Ganciclovir	>100	0.06	0.06	>100	>100	2.9	-
Zalcitabine	>250	-	-	-	-	-	7.5
Alovudine	>250	-	-	-	-	-	16

^a Required to cause a microscopically detectable alteration of normal cell morphology.^b Required to reduce virus-induced cytopathogenicity by 50 %.

Table 4: Cytotoxicity and antiviral activity of compounds in HeLa cell cultures

Compound	Minimum cytotoxic concentration ^a (μ M)	EC ₅₀ ^b (μ M)		
		Vesicular stomatitis virus	Coxsackie virus B4	Respiratory syncytial virus
IV 1	100	>20	>20	>20
IV 2	100	>20	>20	>20
IV 3	100	>20	>20	>20
IV 4	100	>20	>20	>20
IV 5	100	>20	>20	>20
IV 6	100	>20	>20	>20
IV 7	100	>20	>20	>20
IV 8	100	>20	>20	>20
IV 9	100	>20	>20	>20
IV 10	100	>20	>20	>20
IV 11	100	>20	>20	>20
IV 12	>100	>100	>100	>100
IV 13	100	>20	>20	>20
IV 14	$\geq$ 100	>100	>100	>100
IV 15	>100	>100	>100	>100
IV 16	$\geq$ 100	>100	>100	>100
IV 17	100	>20	>20	>20
IV 18	100	>20	>20	>20
IV 19	100	9.5	>20	>20
IV 20	100	>20	>20	>20
IV 21	>100	>100	>100	>100
IV 22	>100	>100	>100	>100
IV 23	100	>20	>20	>20
IV 24	100	20	>20	11
IV 25	100	9.5	>20	10
IV 26	>100	>100	>100	>100
IV 27	20	>4	>4	>4
IV 28	100	>20	>20	>20
IV 29	20	>4	>4	>4
IV 30	20	>4	>4	>4
IV 31	20	>4	>4	>4
DS-10000 (μ g/ml)	>100	1.8	27	3.0
Ribavirin	>250	22	81	7.5

^a Required to cause a microscopically detectable alteration of normal cell morphology.^b Required to reduce virus-induced cytopathogenicity by 50 %.

Table 7: Anti-Feline Corona Virus (FIPV) and anti-Feline Herpes Virus activity and cytotoxicity in CRFK cell cultures.

Compound	CC ₅₀ ^a (μM)	EC ₅₀ ^b (μM)	
		Feline Corona Virus (FIPV)	Feline Herpes Virus
(IV 1)	62.5	>20	>20
(IV 2)	>100	>100	>100
(IV 3)	>100	>100	>100
(IV 4)	47.0	>20	>20
(IV 5)	73.4	>20	>20
(IV 6)	>100	>100	>100
(IV 7)	>100	>100	>100
(IV 8)	55.4	>20	>20
(IV 9)	>100	>100	>100
(IV 10)	46.7	>20	>20
(IV 11)	50.0	>20	>20
(IV 12)	>100	>100	>100
(IV 13)	>100	>100	>100
(IV 14)	>100	>100	>100
(IV 15)	>100	>100	>100
(IV 16)	>100	>100	>100
(IV 17)	85.0	>20	>20
(IV 18)	>100	>100	>100
(IV 19)	52.2	>20	>20
(IV 20)	48.8	>20	>20
(IV 21)	>100	>100	>100
(IV 22)	>100	>100	>100
(IV 23)	50.0	>20	>20
(IV 24)	20.3	>20	>20
(IV 25)	37.6	>20	>20
(IV 26)	>100	>100	>100
(IV 27)	9.5	>4	>4
(IV 28)	23.1	>20	>20
(IV 29)	13.7	>4	>4
(IV 30)	9.7	>4	>4
(IV 31)	39.1	>20	>20
HHA (μg/ml)	>100	24.3	15.1
UDA (μg/ml)	>100	7.3	4.3
Ganciclovir	>100	>100	8.6

^a 50% Cytotoxic concentration, as determined by measuring the cell viability with the colorimetric formazan-based MTS assay.

^b 50% Effective concentration, or concentration producing 50% inhibition of virus-induced cytopathic effect, as determined by measuring the cell viability with the colorimetric formazan-based MTS assay

A facile one pot multi component synthesis of alkyl 4-oxo-coumarinyl ethylidene hydrazono-thiazolidin-5-ylidene acetates and their antiviral activity

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Keywords: One- pot three - component reaction, 3-acetylcoumarin, thiosemicarbazide, thiazolidinone and antiviral activity.

Content

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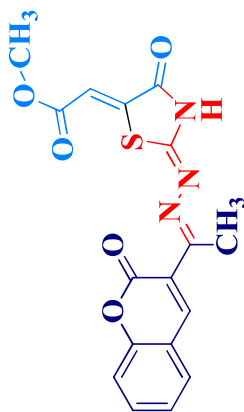
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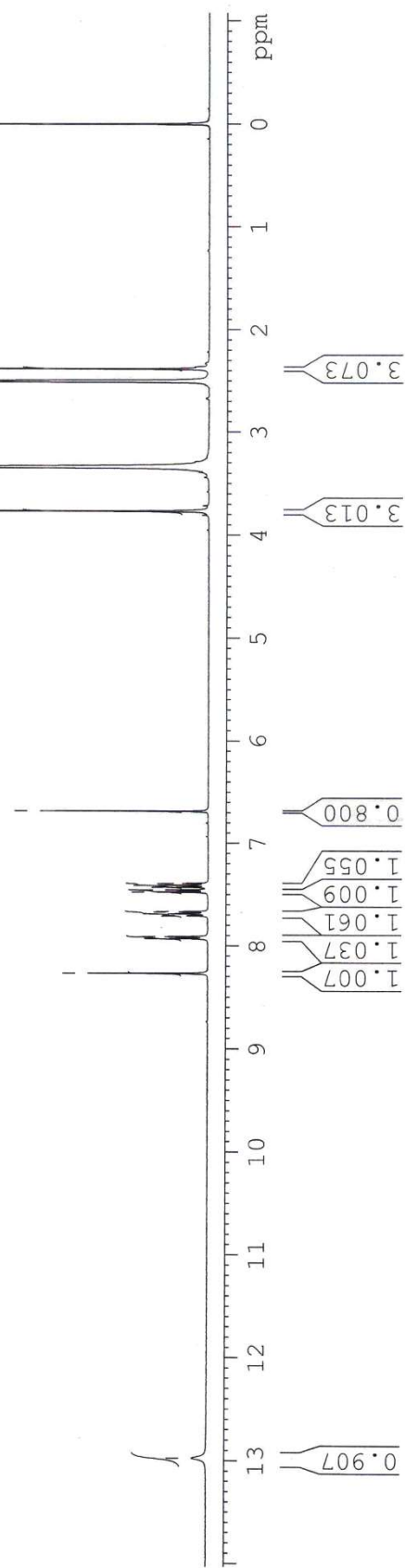
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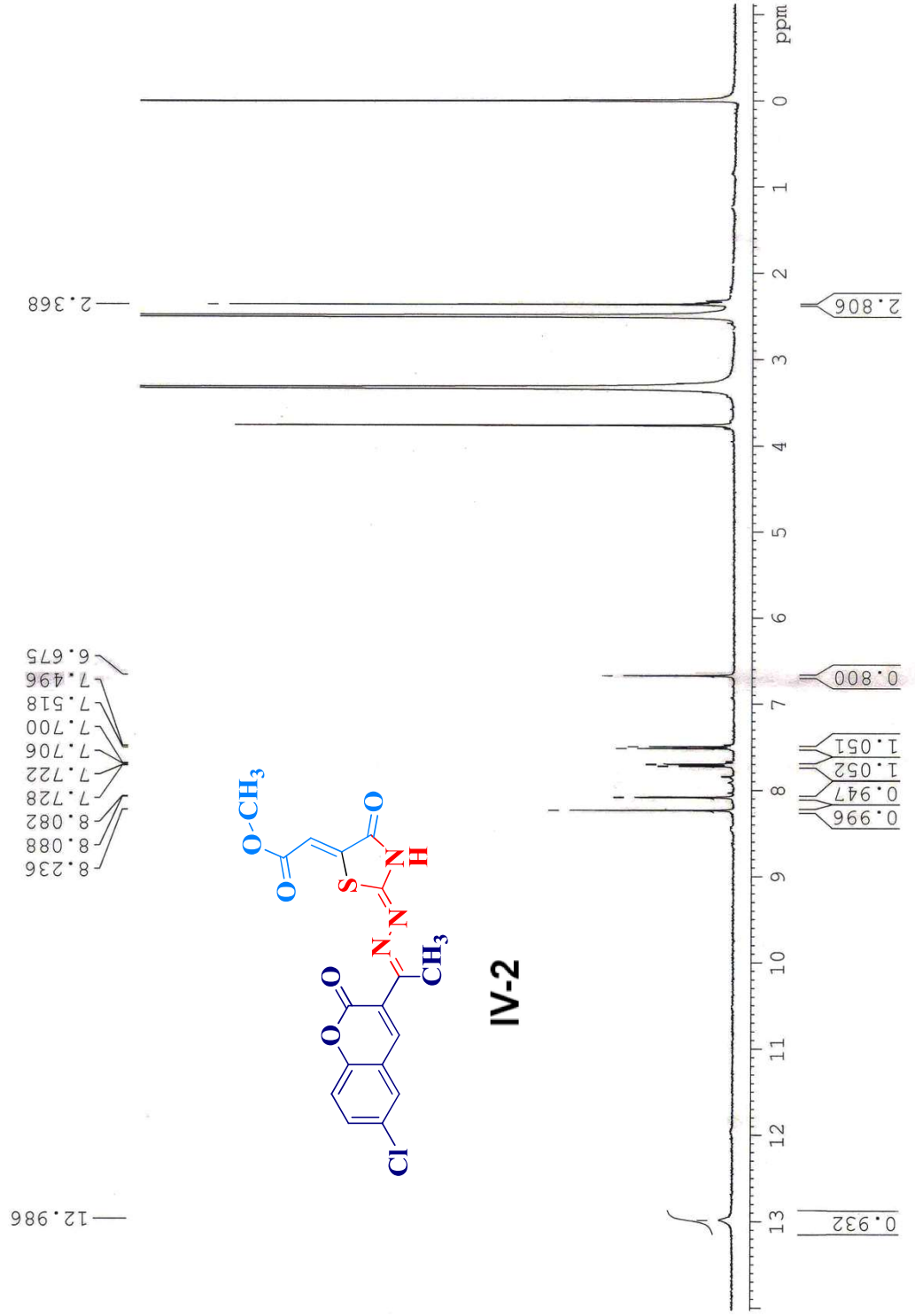
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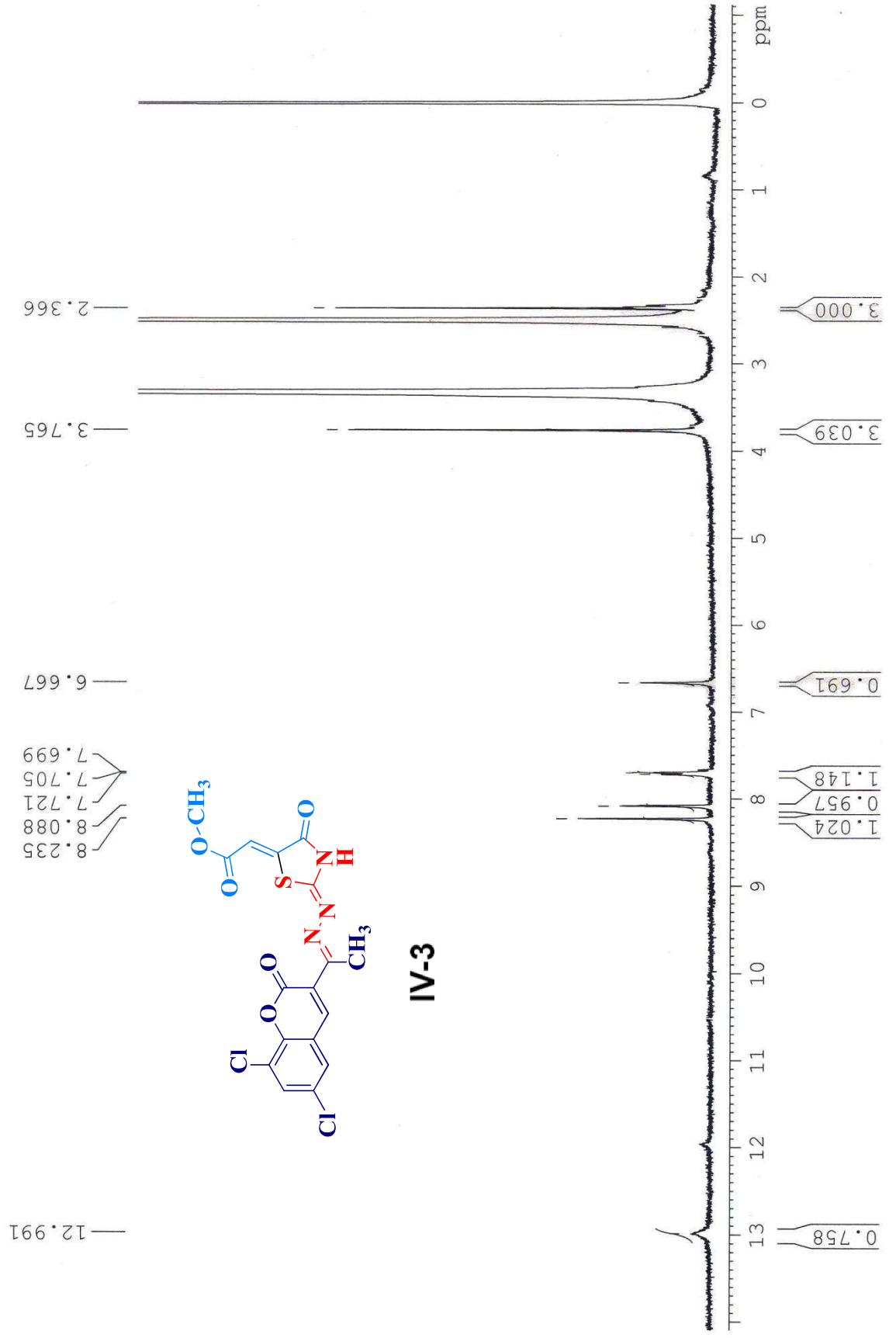
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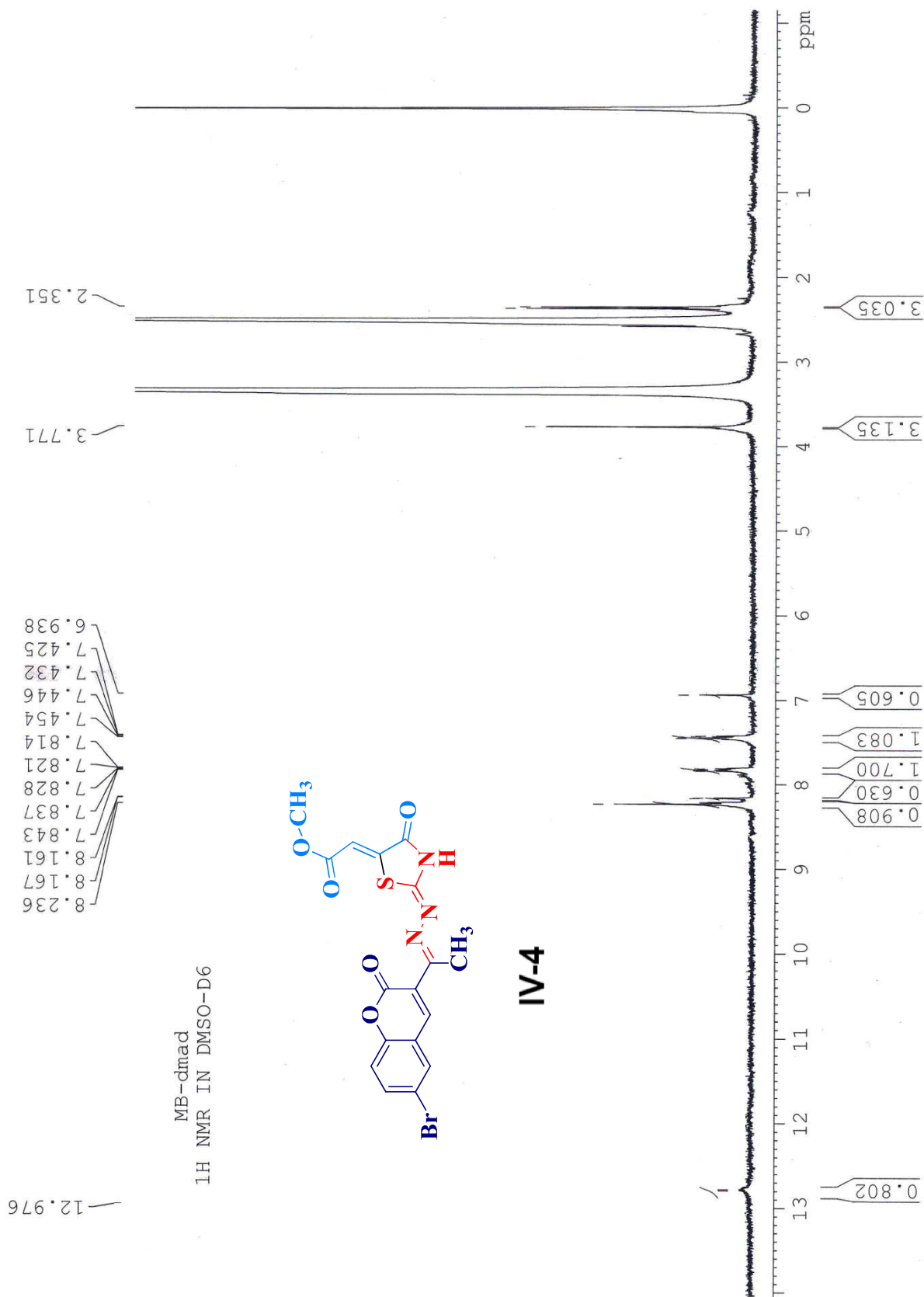


MC+DMAD
1H NMR IN DMSO-D6

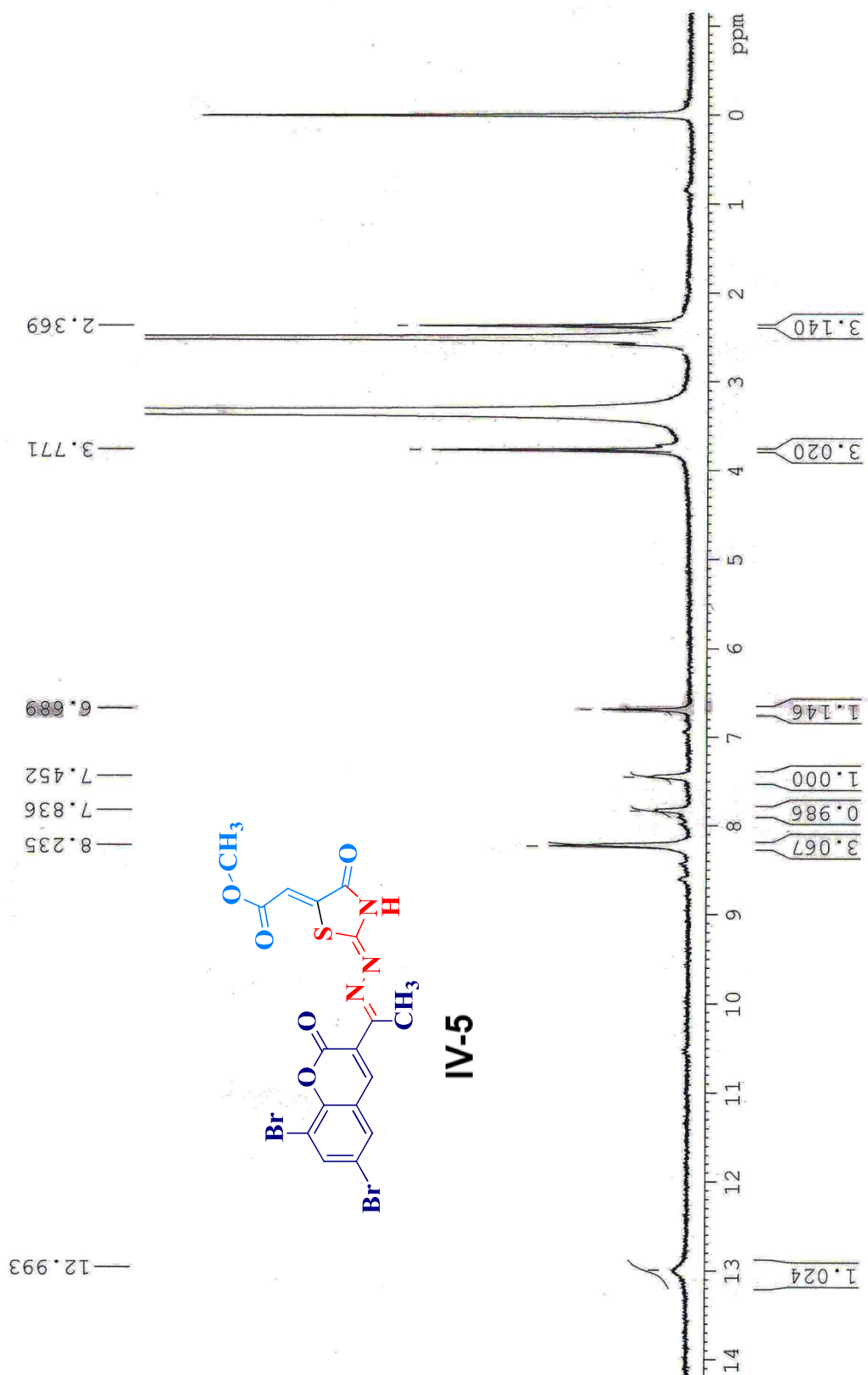


DC+DMAD
1H NMR IN DMSO-D6

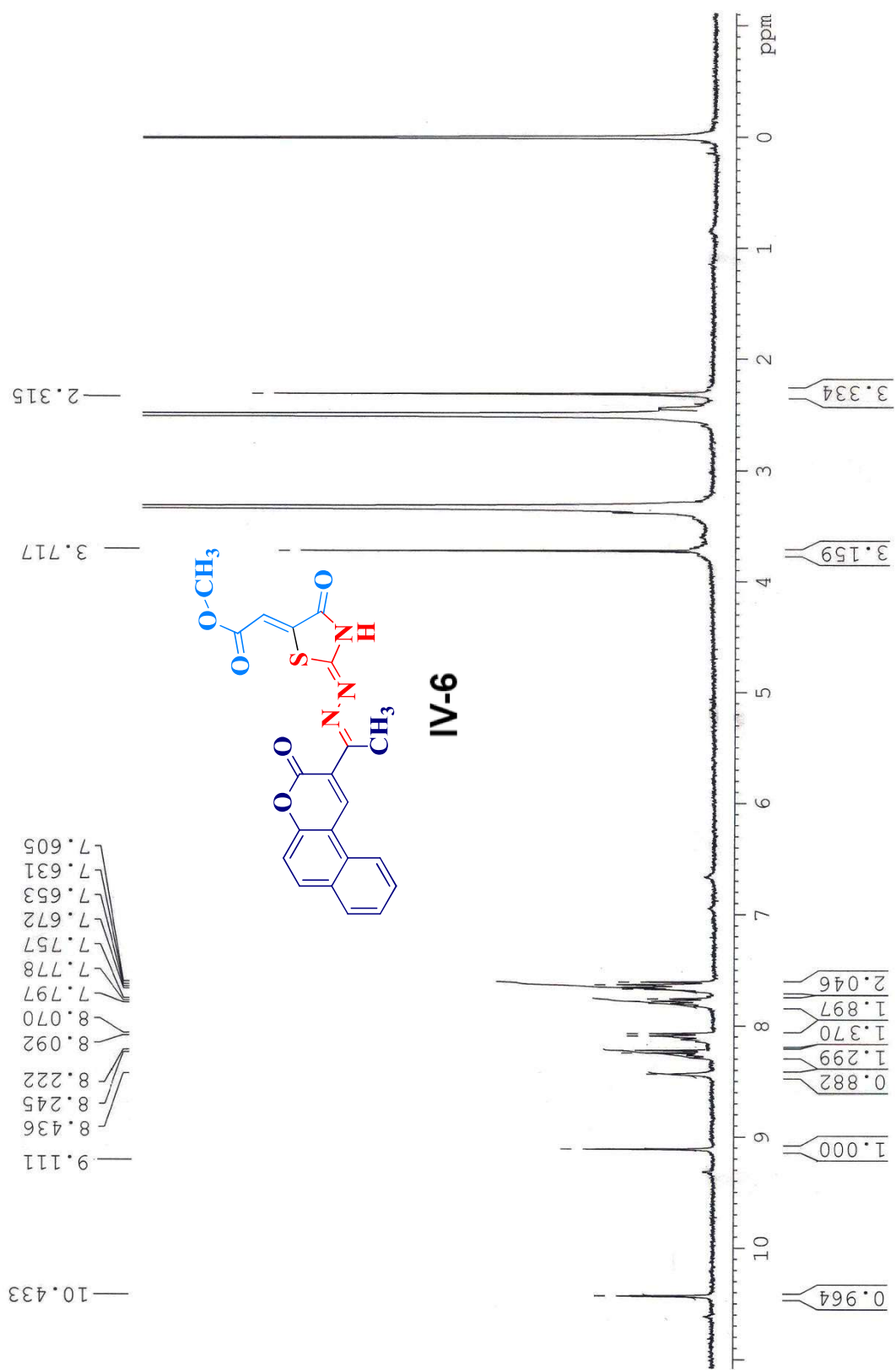




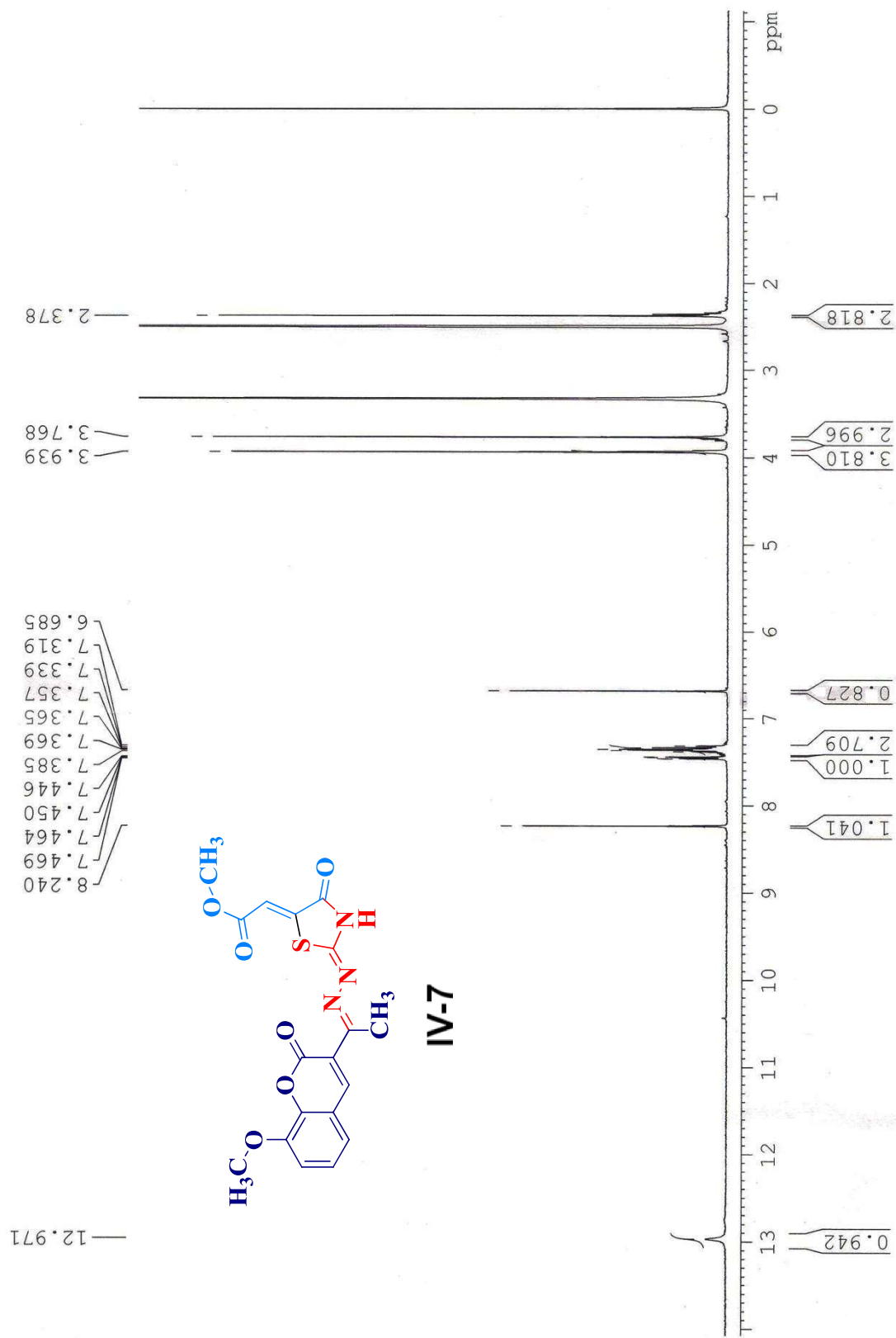
DB-dmad
1H NMR IN DMSO-D6



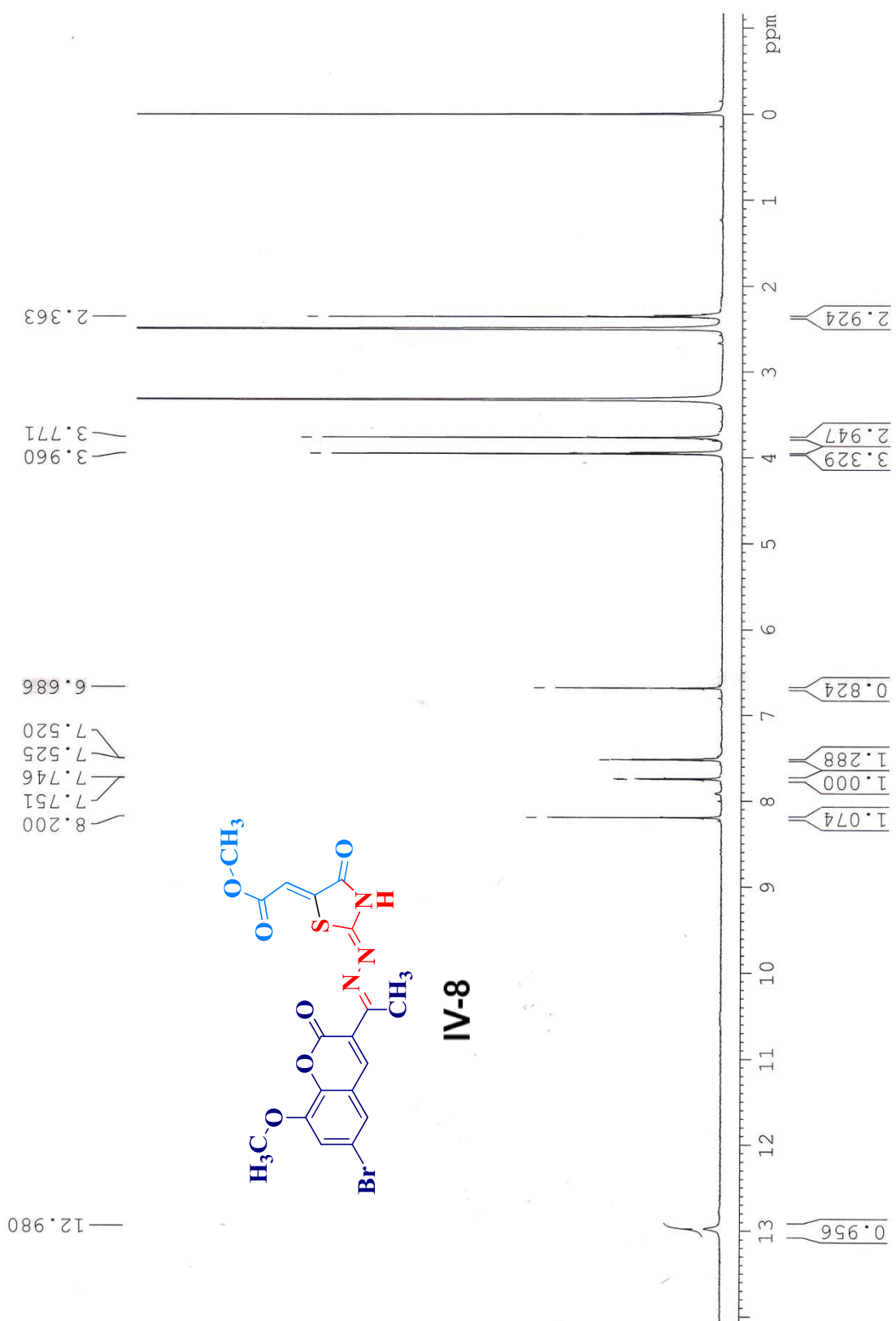
NAP+DMAD
1H NMR IN DMSO-D6



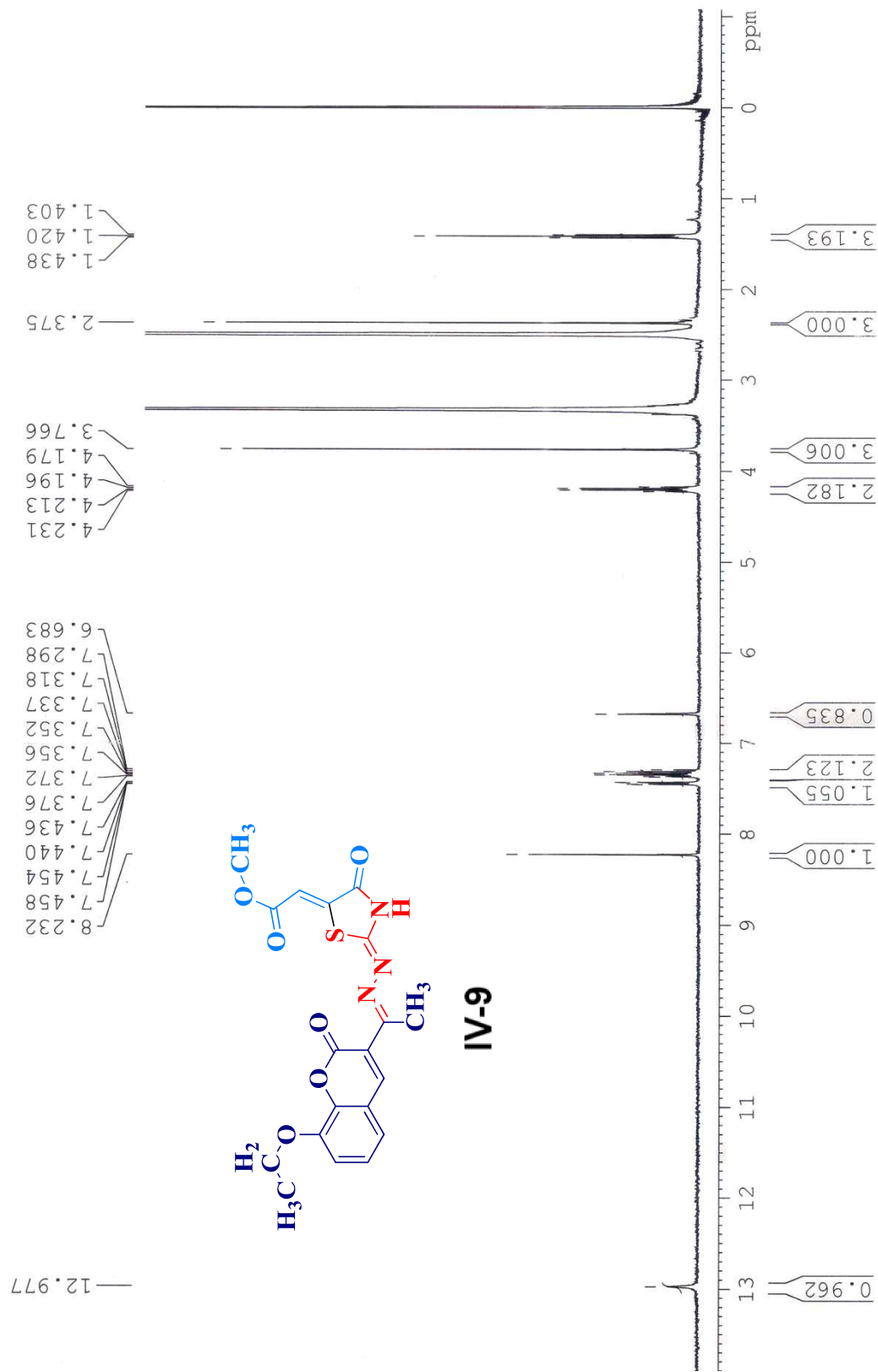
OV+DMAD
1H NMR IN DMSO-D6



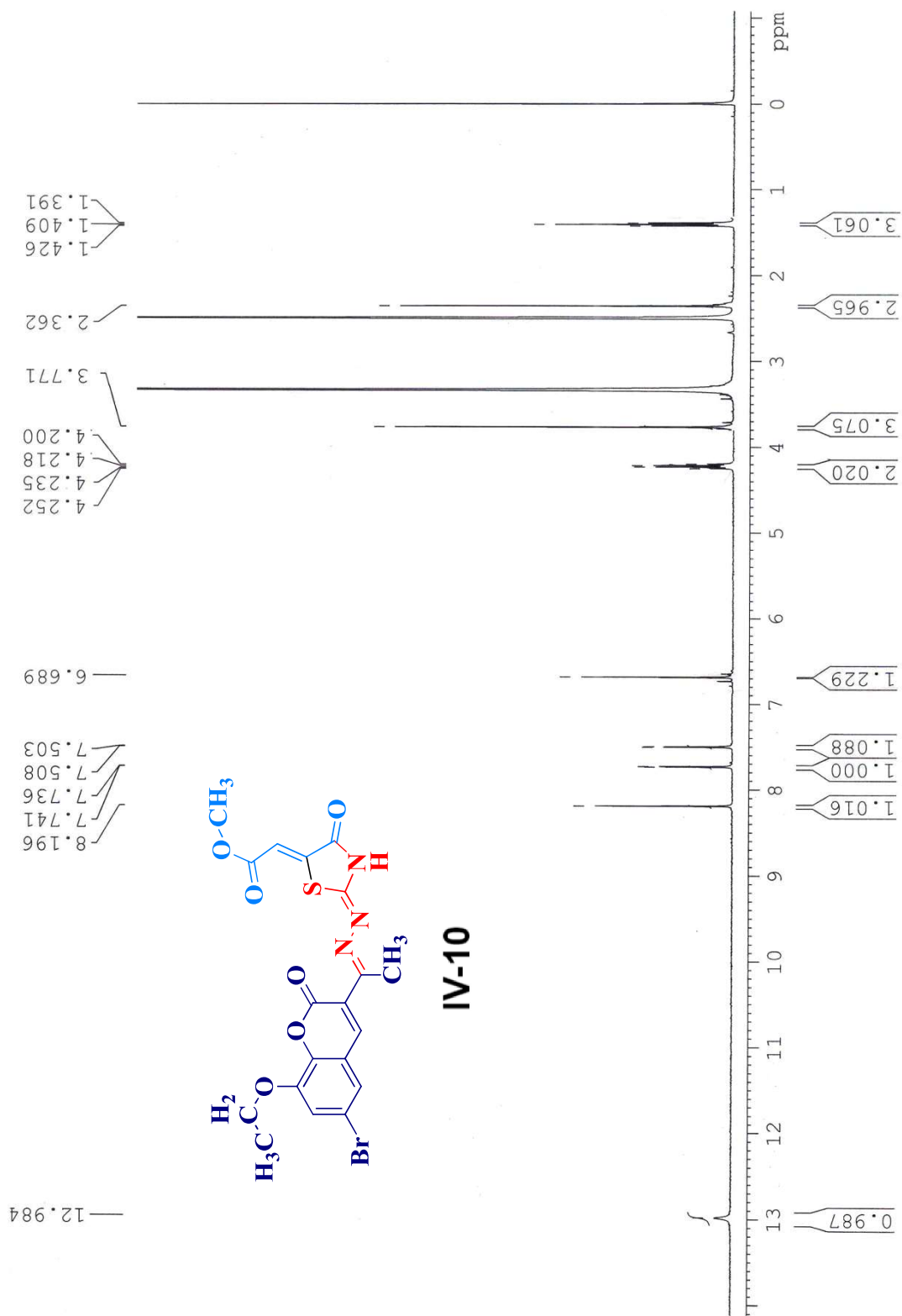
BrOV-dmad
1H NMR IN DMSO-D6



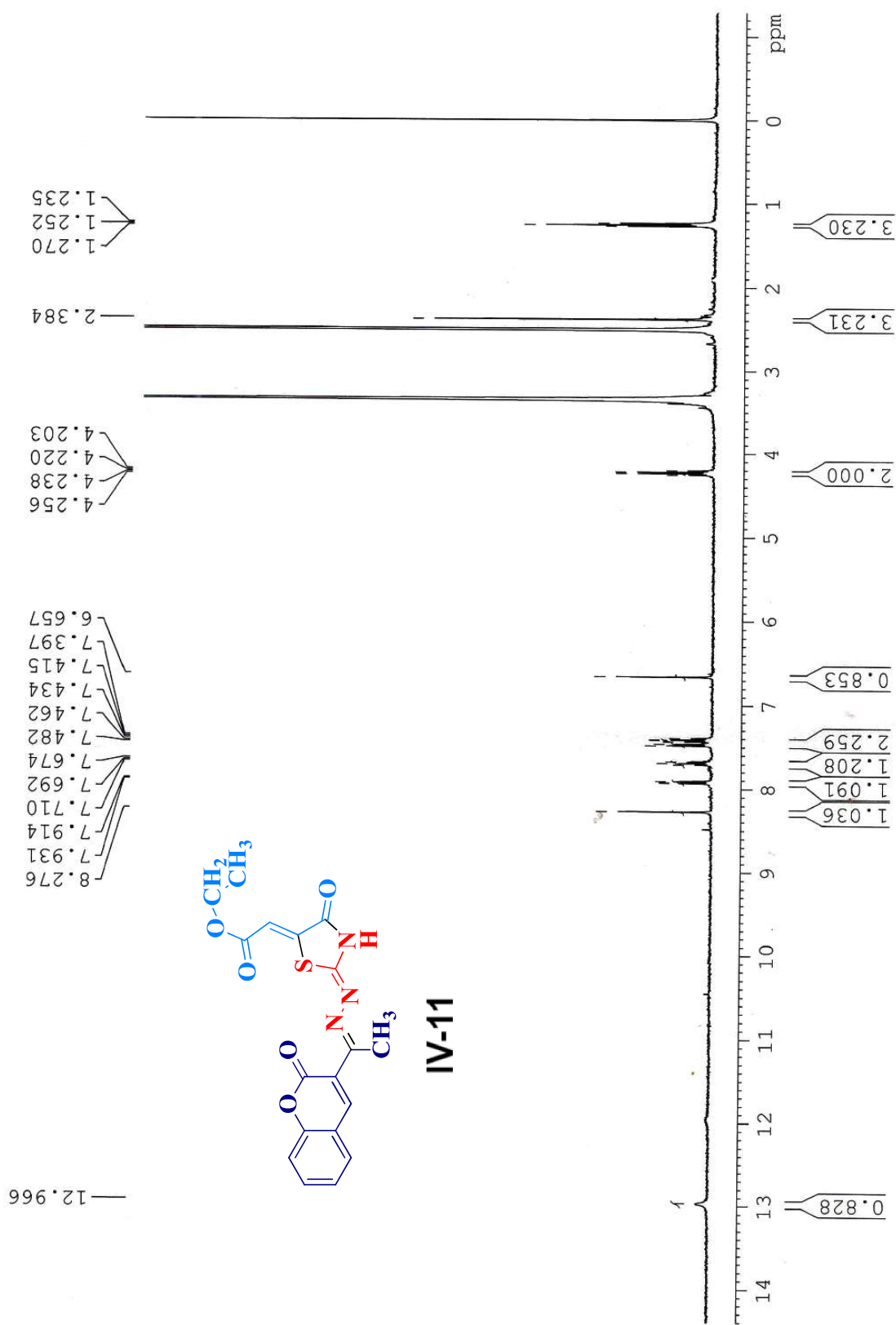
OEE+DMAD
¹H NMR IN DMSO-D6



BroEt-dmad
¹H NMR IN DMSO-D6

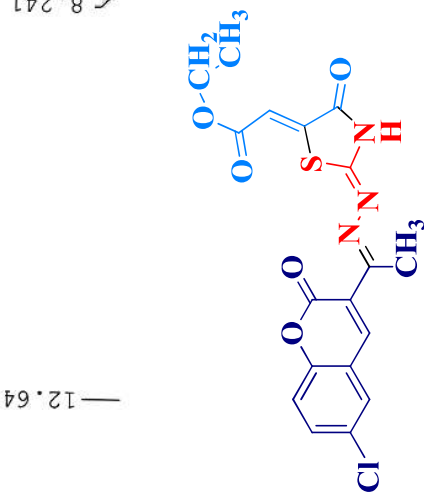


DEAD
¹H NMR IN DMSO-D₆

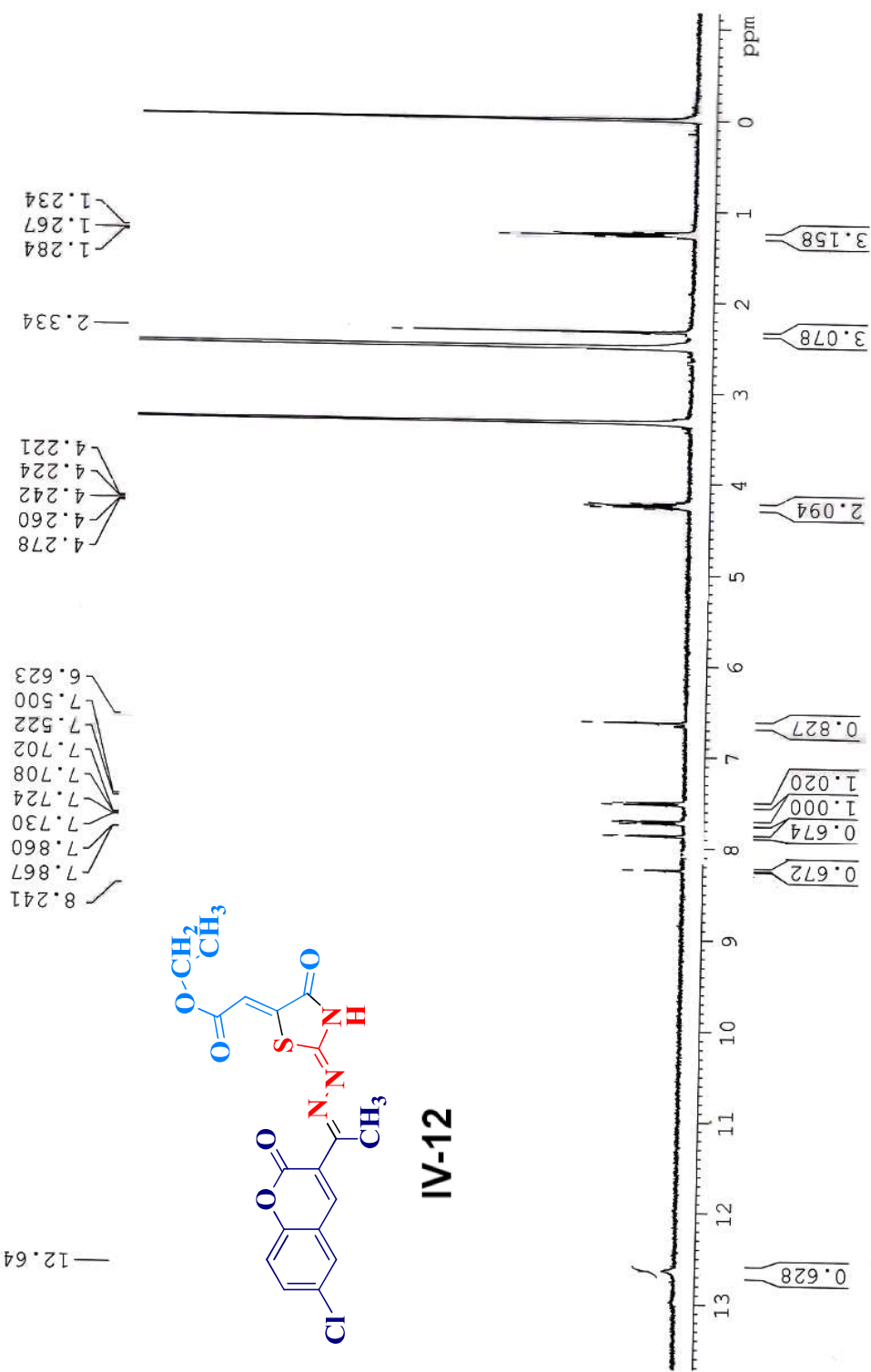


MC
¹H NMR IN DMSO-D6

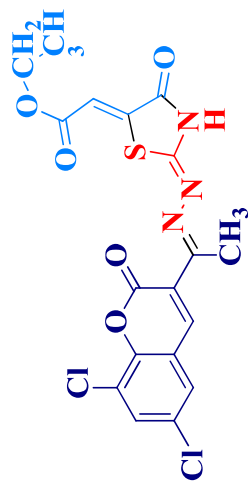
— 12.641



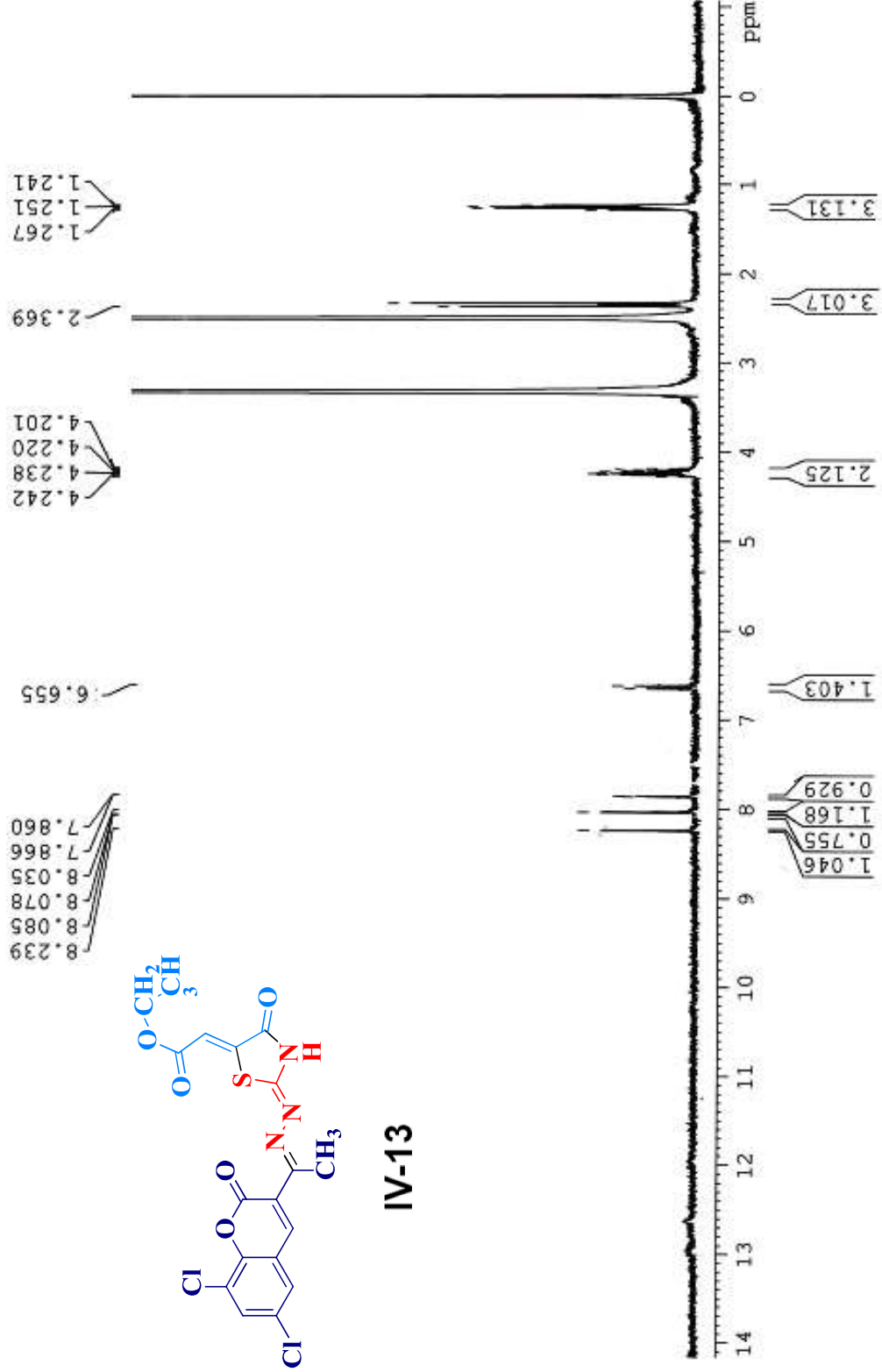
IV-12



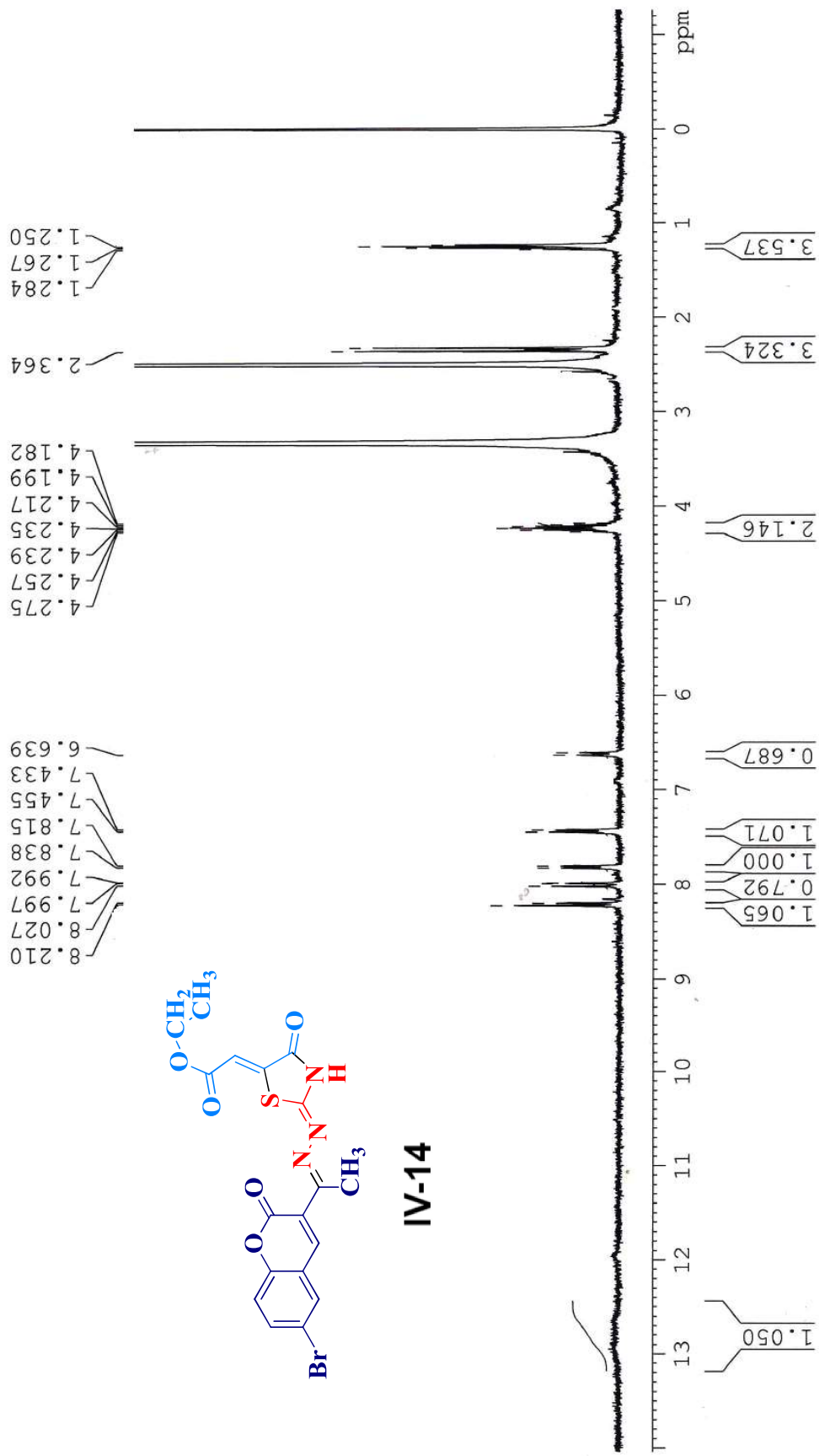
DCD
¹H NMR IN DMSO-D₆



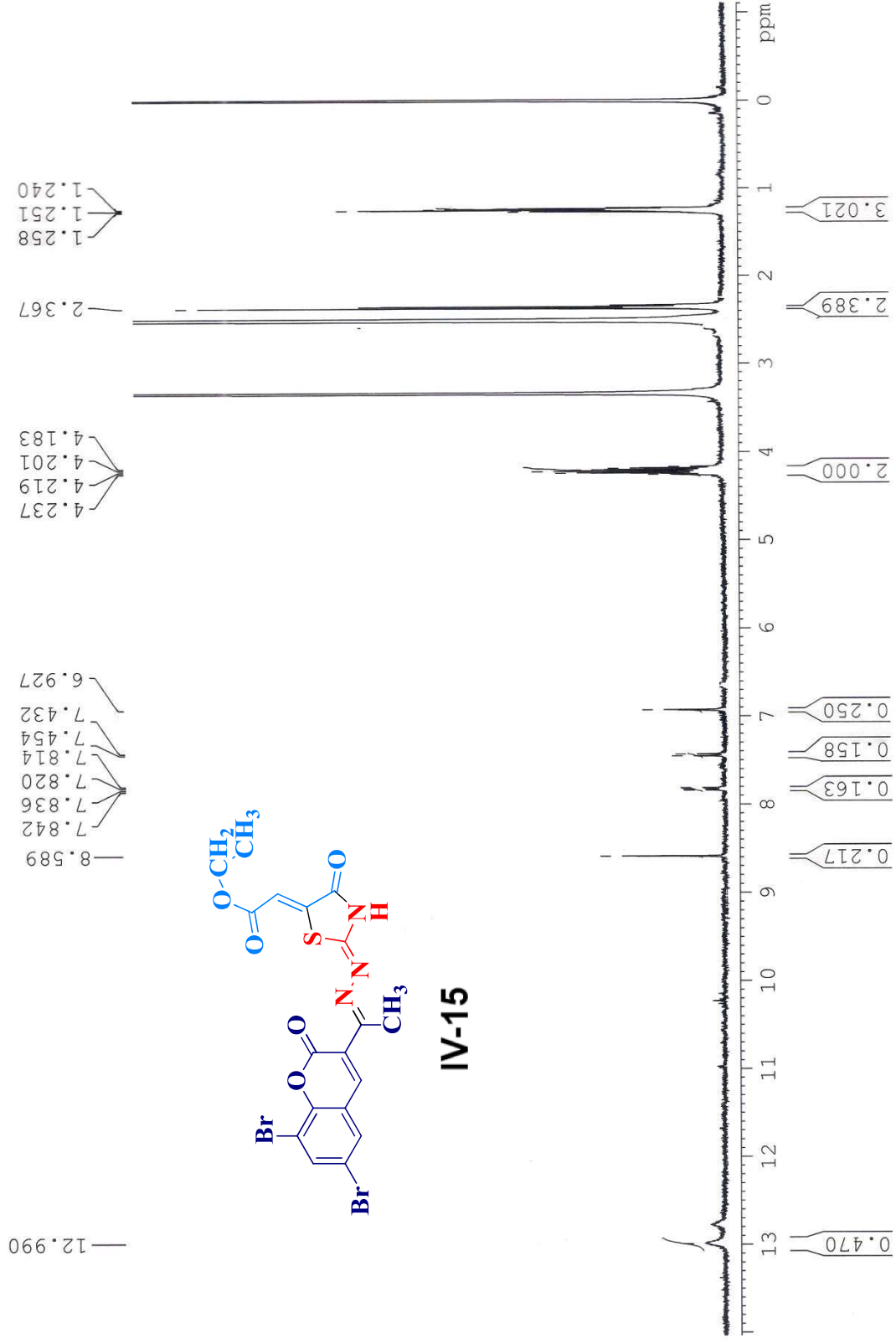
IV-13



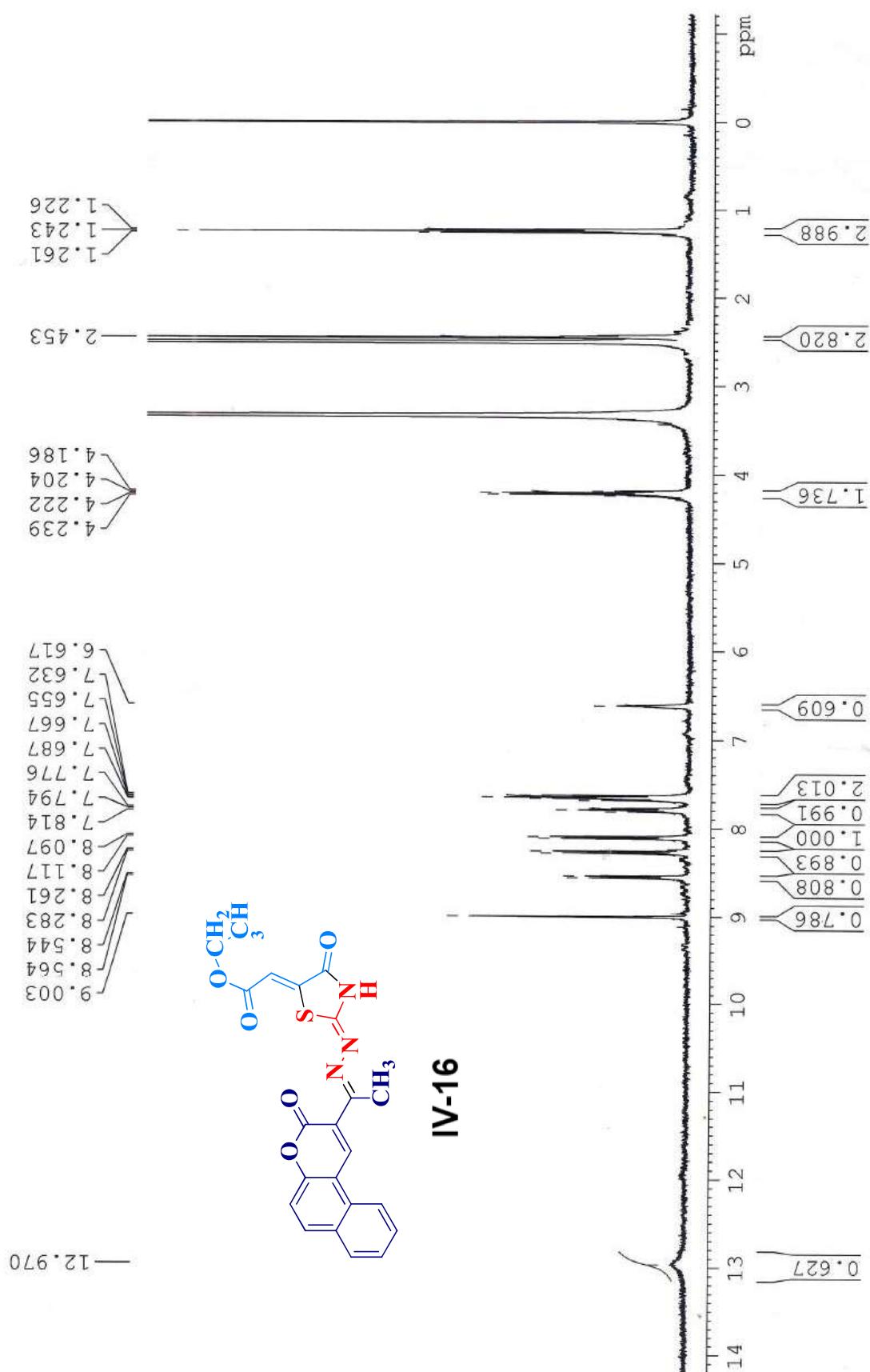
MBD
¹H NMR IN DMSO-D₆



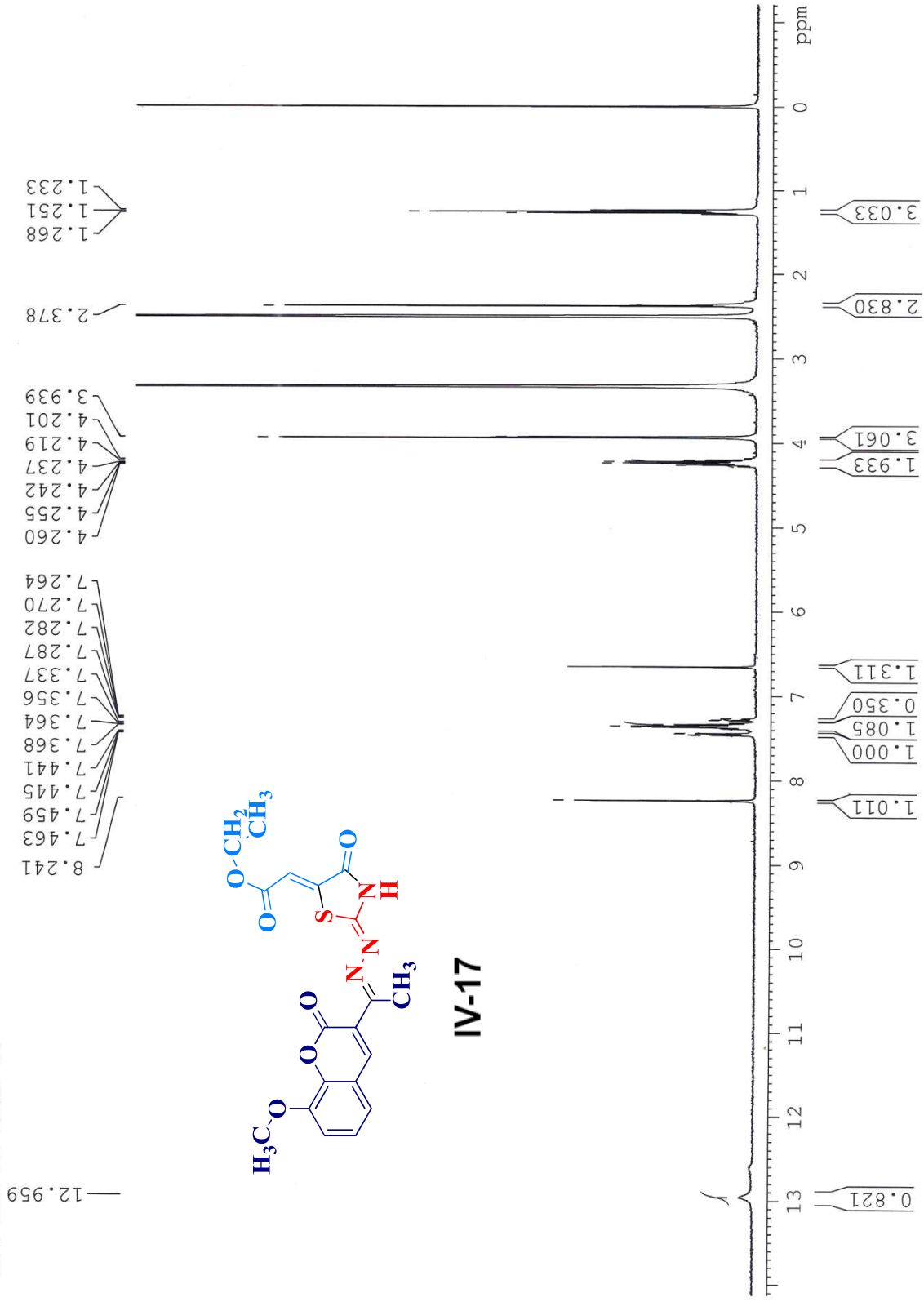
DBD
¹H NMR IN DMSO-D₆



NaPD
¹H NMR IN DMSO-D6

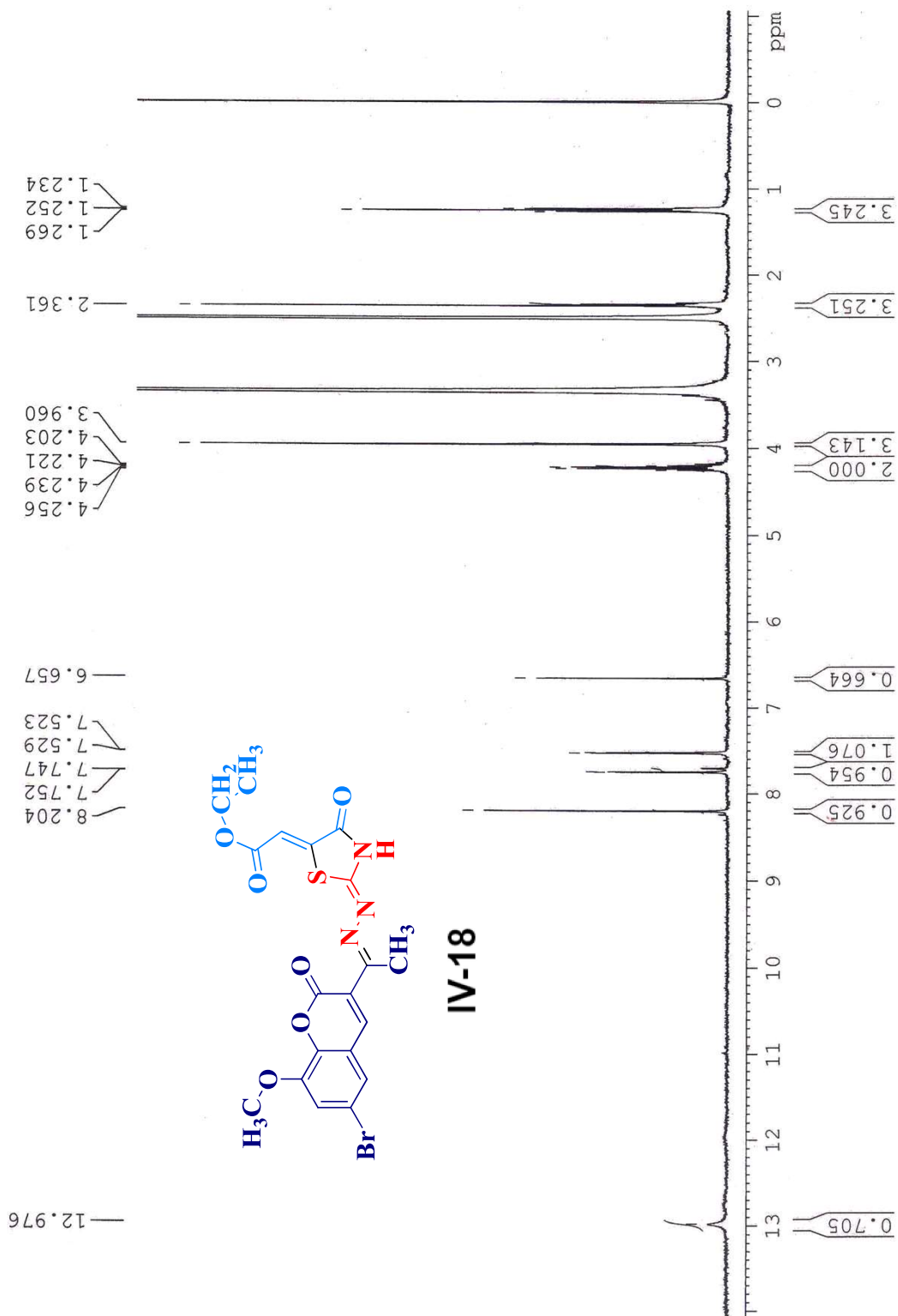


OVD
¹³C NMR IN DMSO-D6

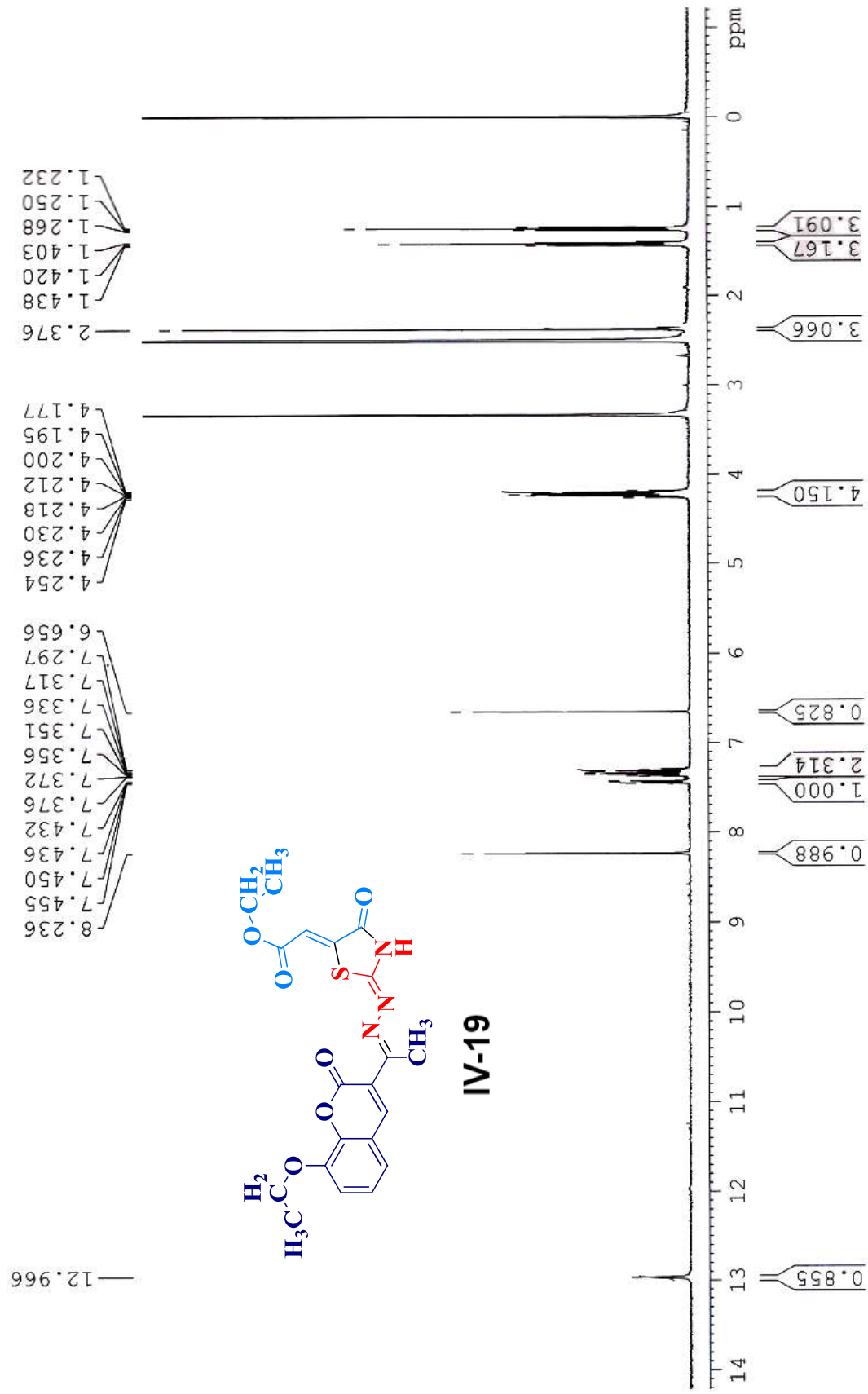


IV-17

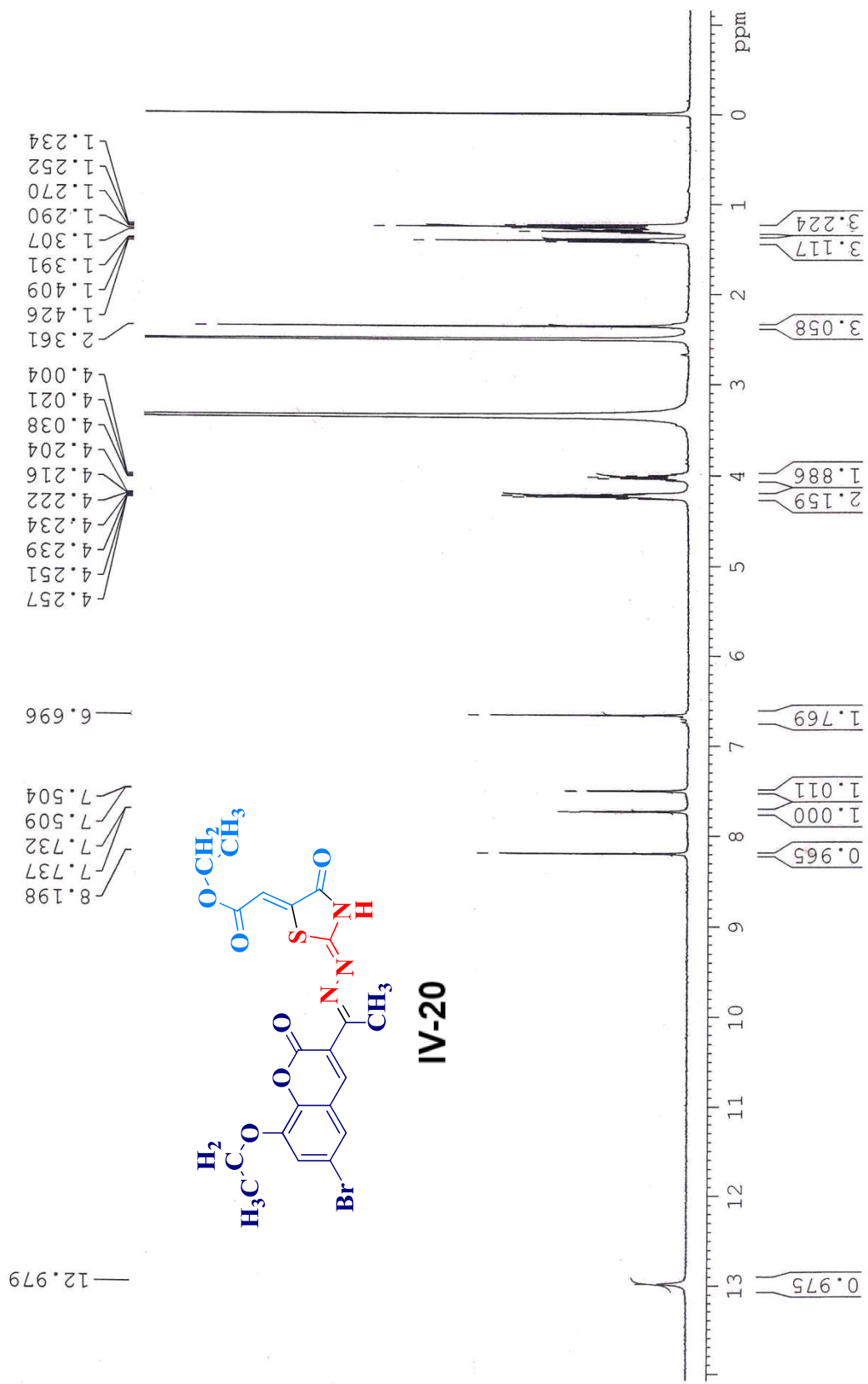
BrOV
¹H NMR IN DMSO-D₆



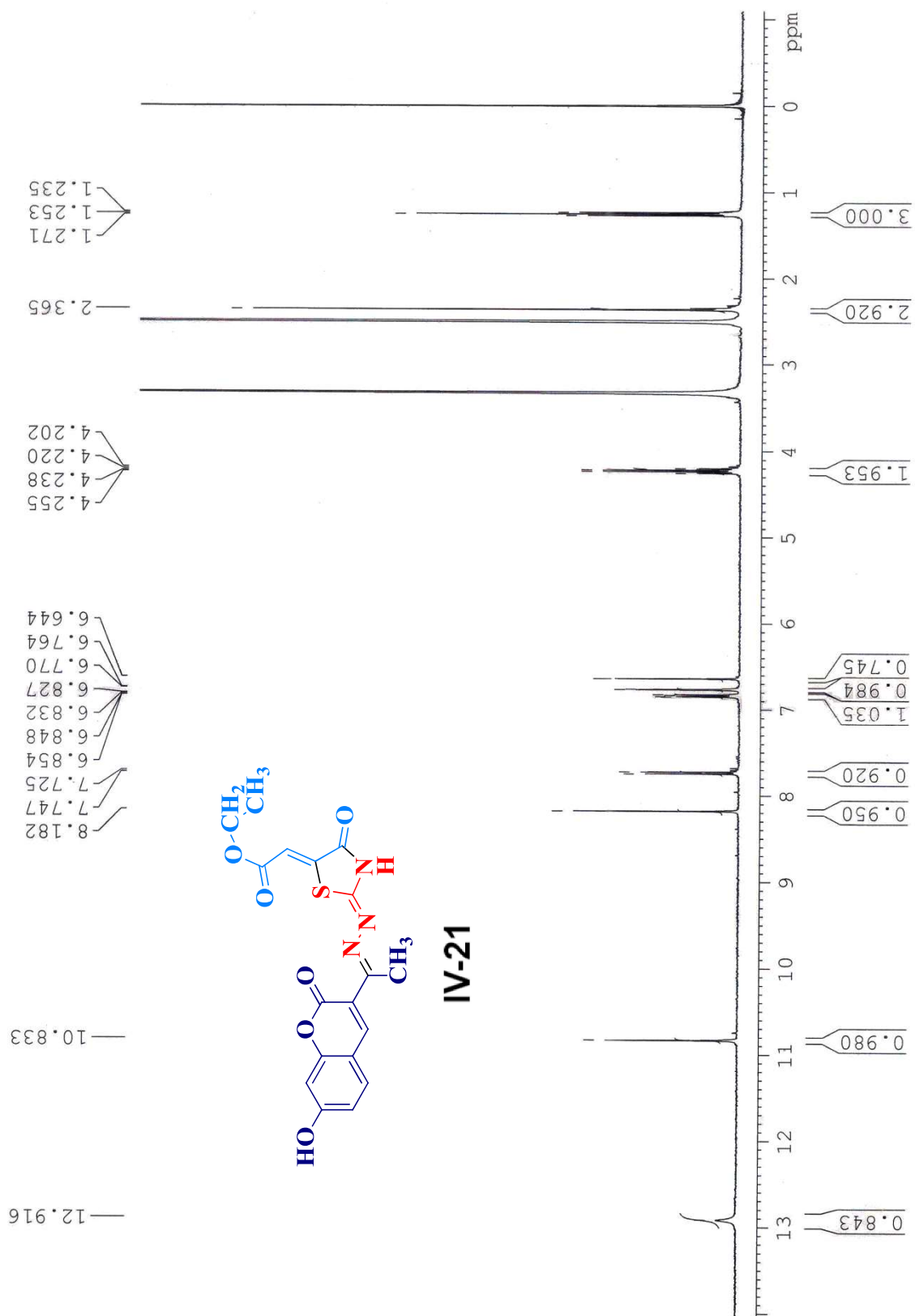
O-Et
¹H NMR IN DMSO-D₆



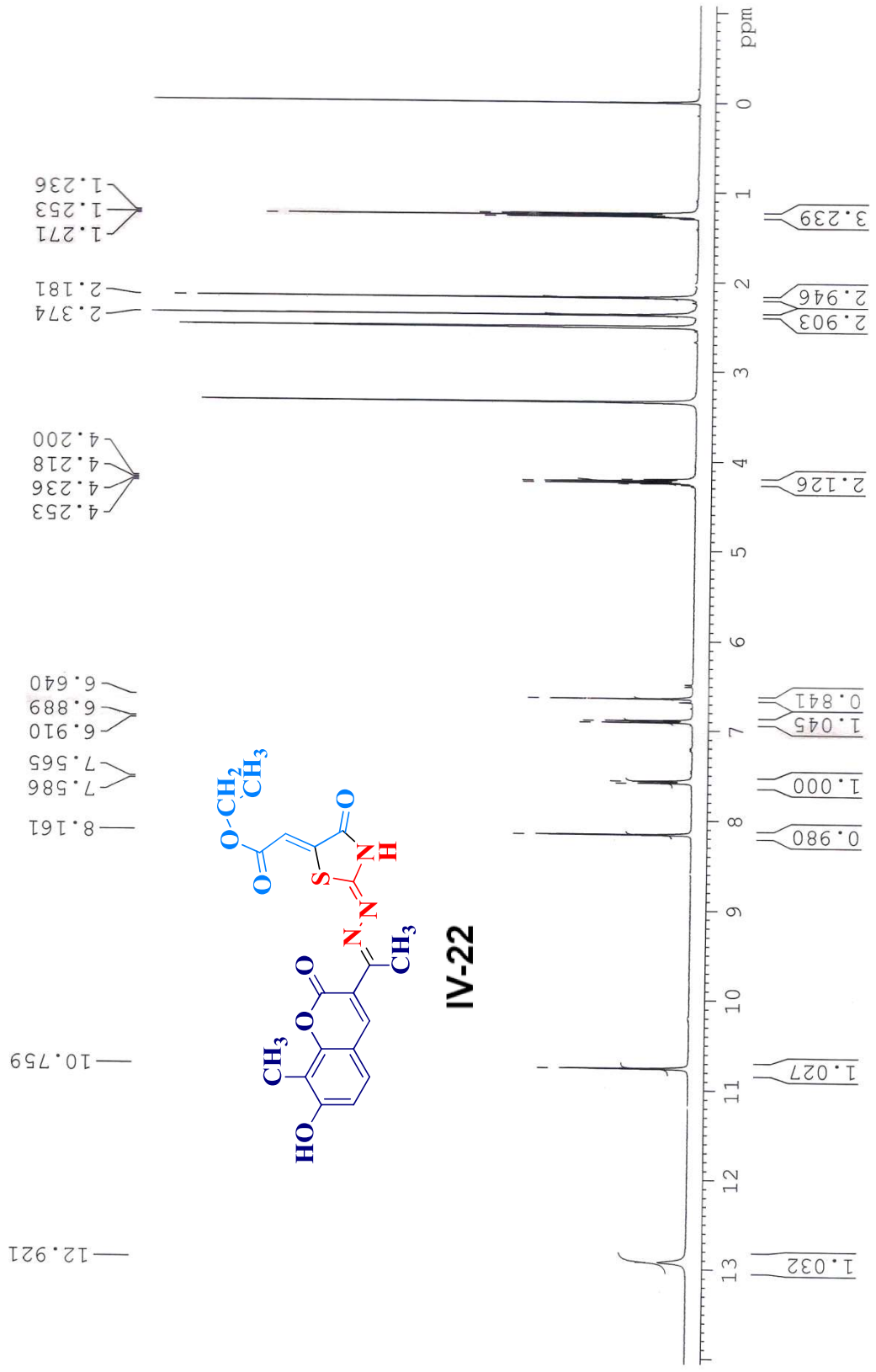
Br-OET
¹H NMR IN DMSO-D6



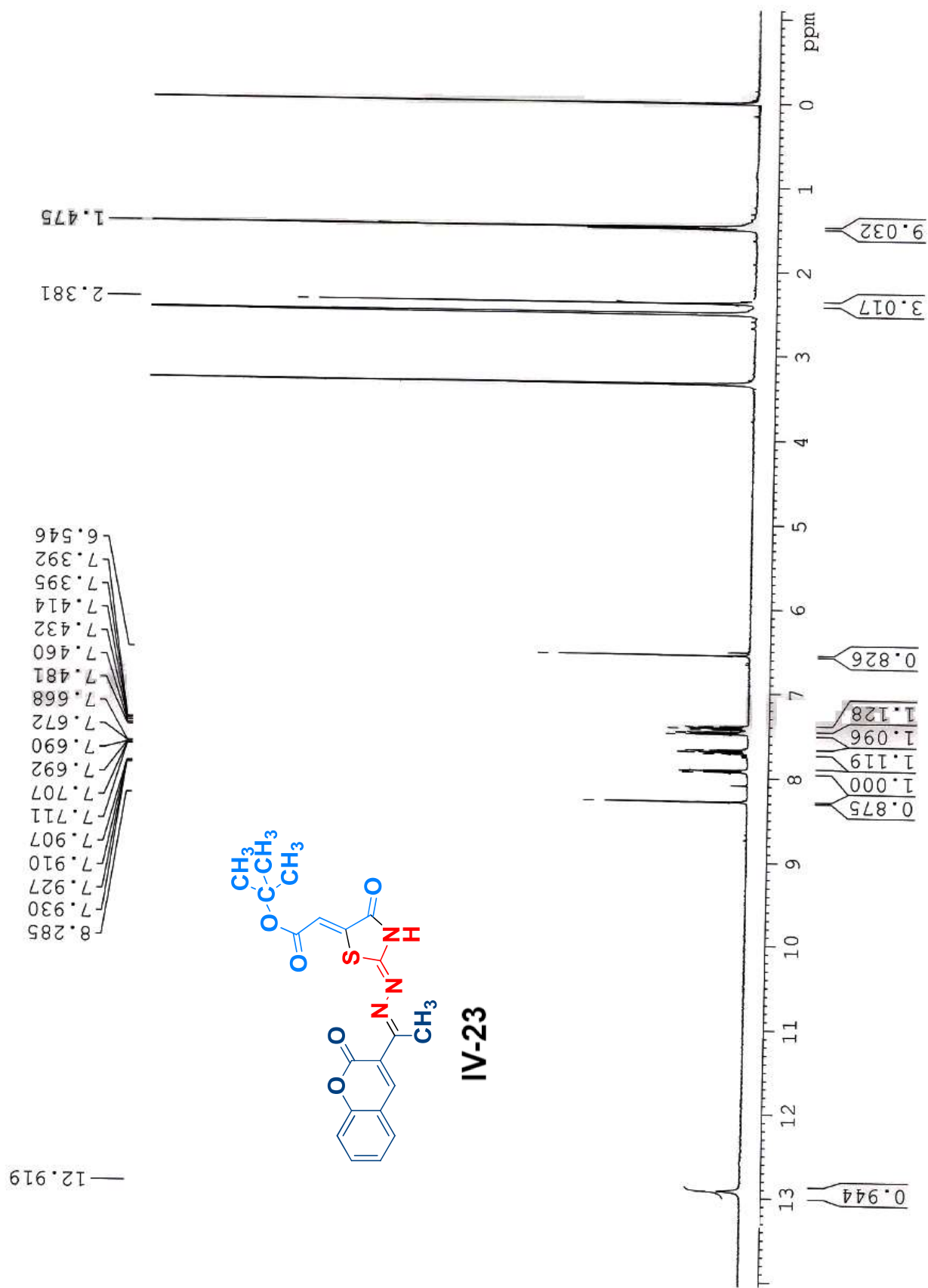
OH+DEAD
NMR IN DMSO-D6



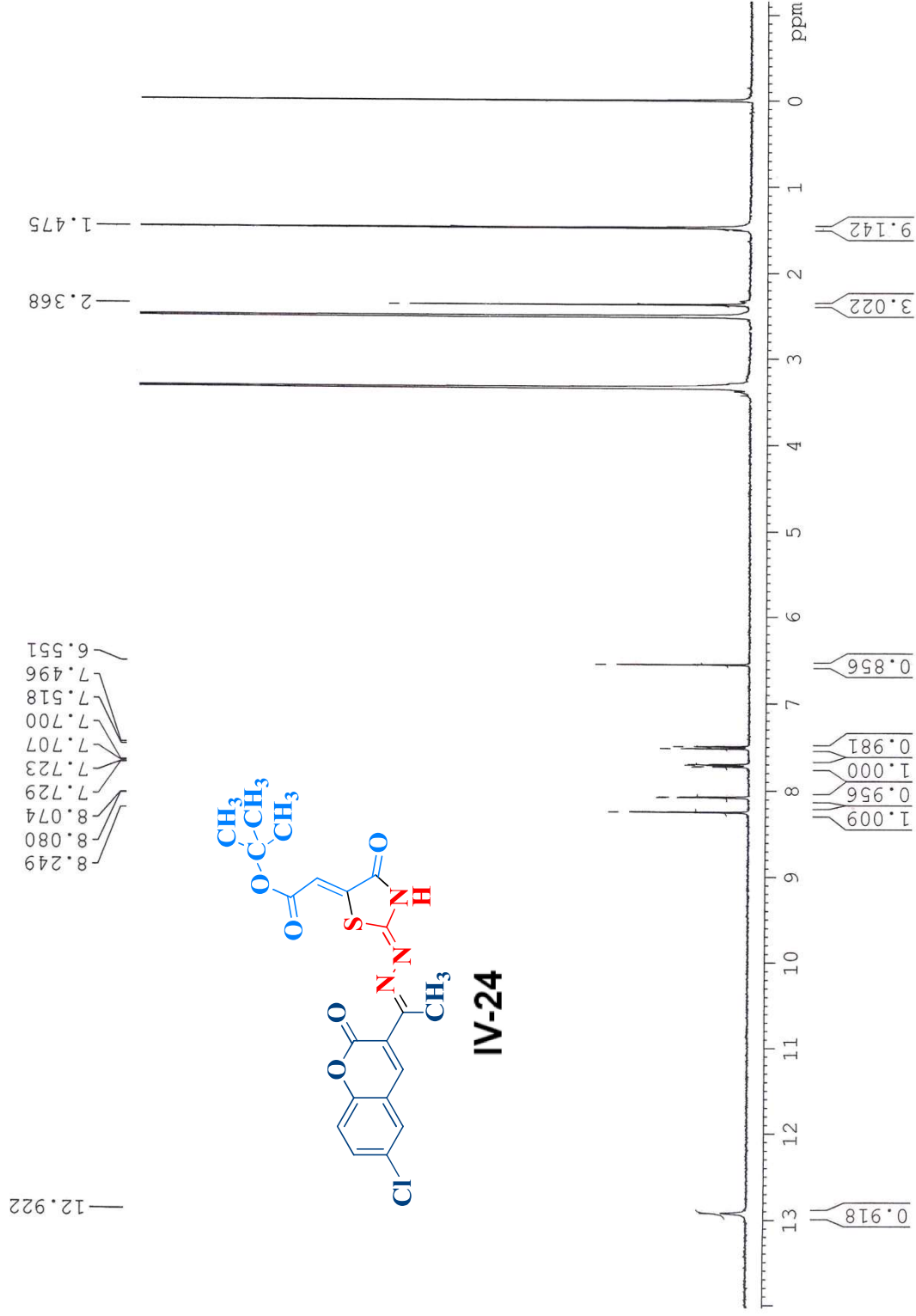
MER
¹H NMR IN DMSO-D6



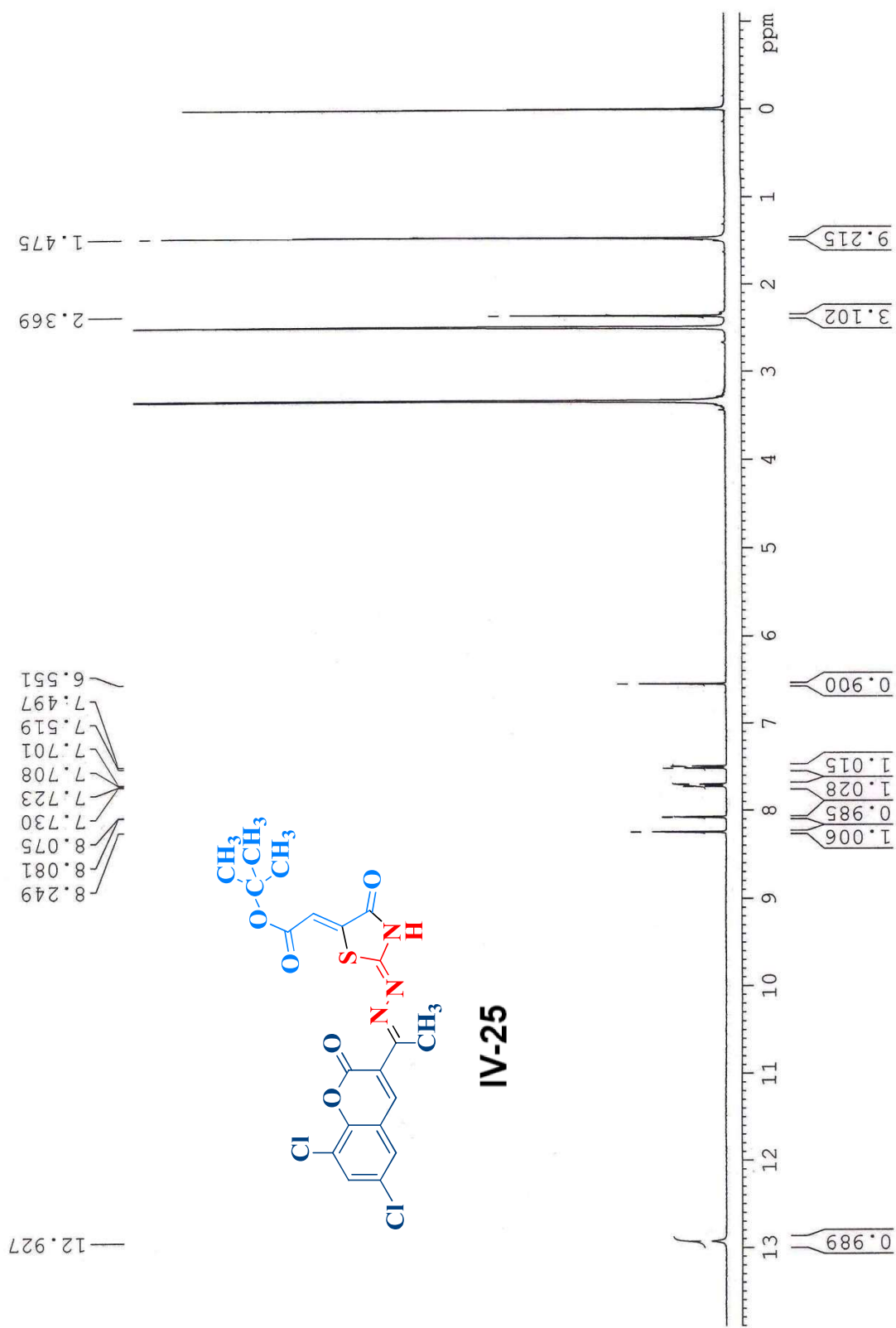
DTB
¹H NMR IN DMSO-D₆



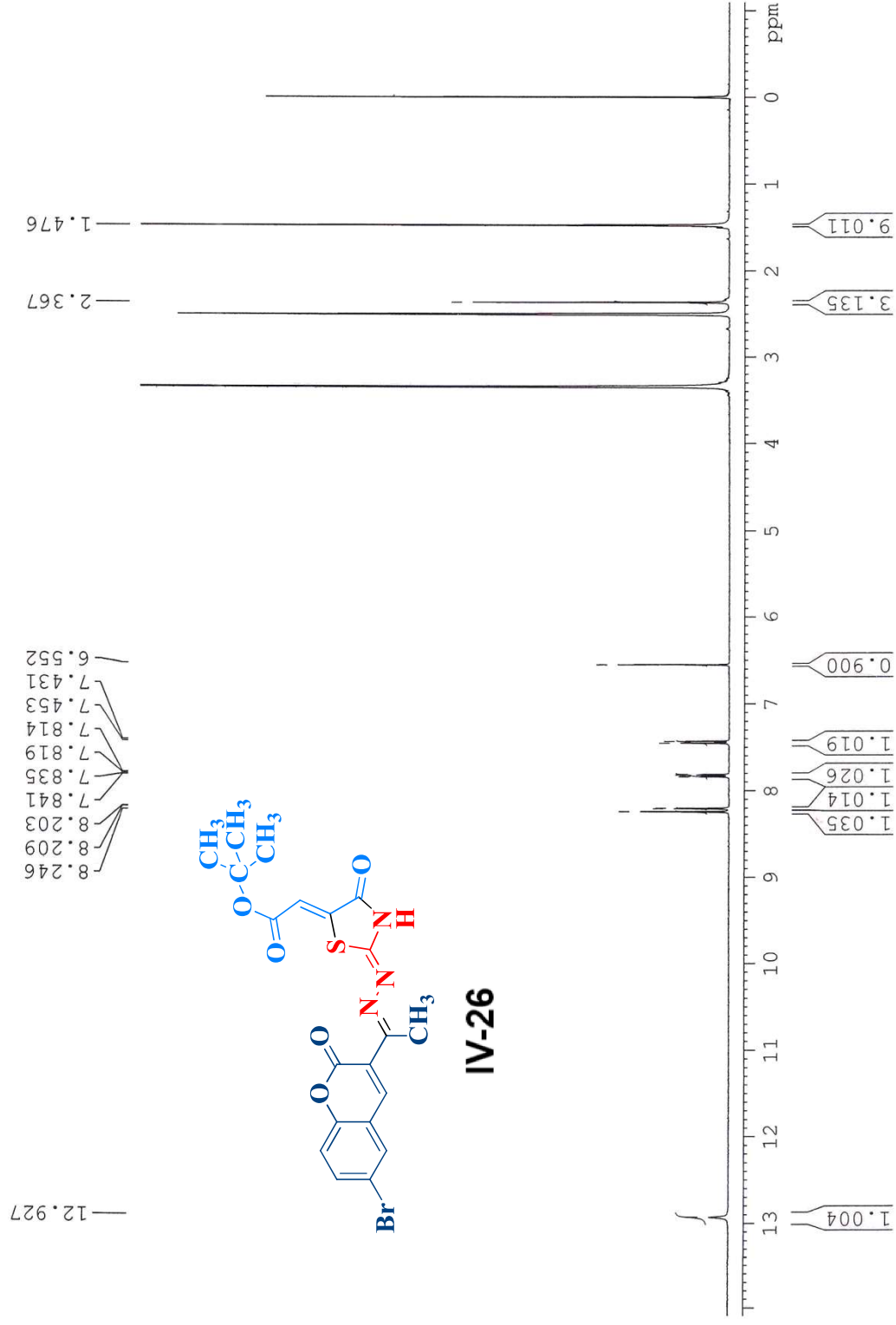
MC
¹H NMR IN DMSO-D₆



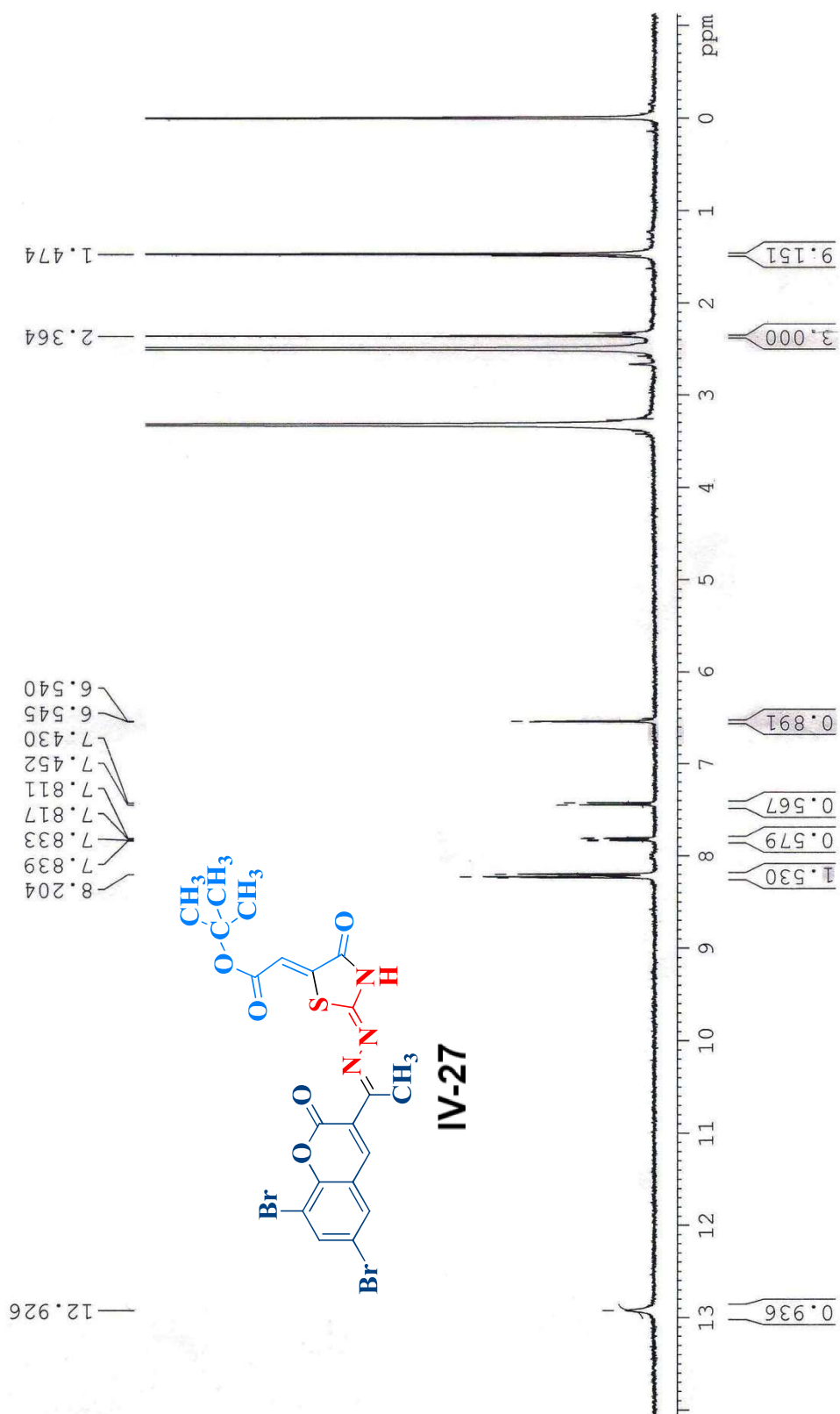
DC
¹H NMR IN DMSO-D6



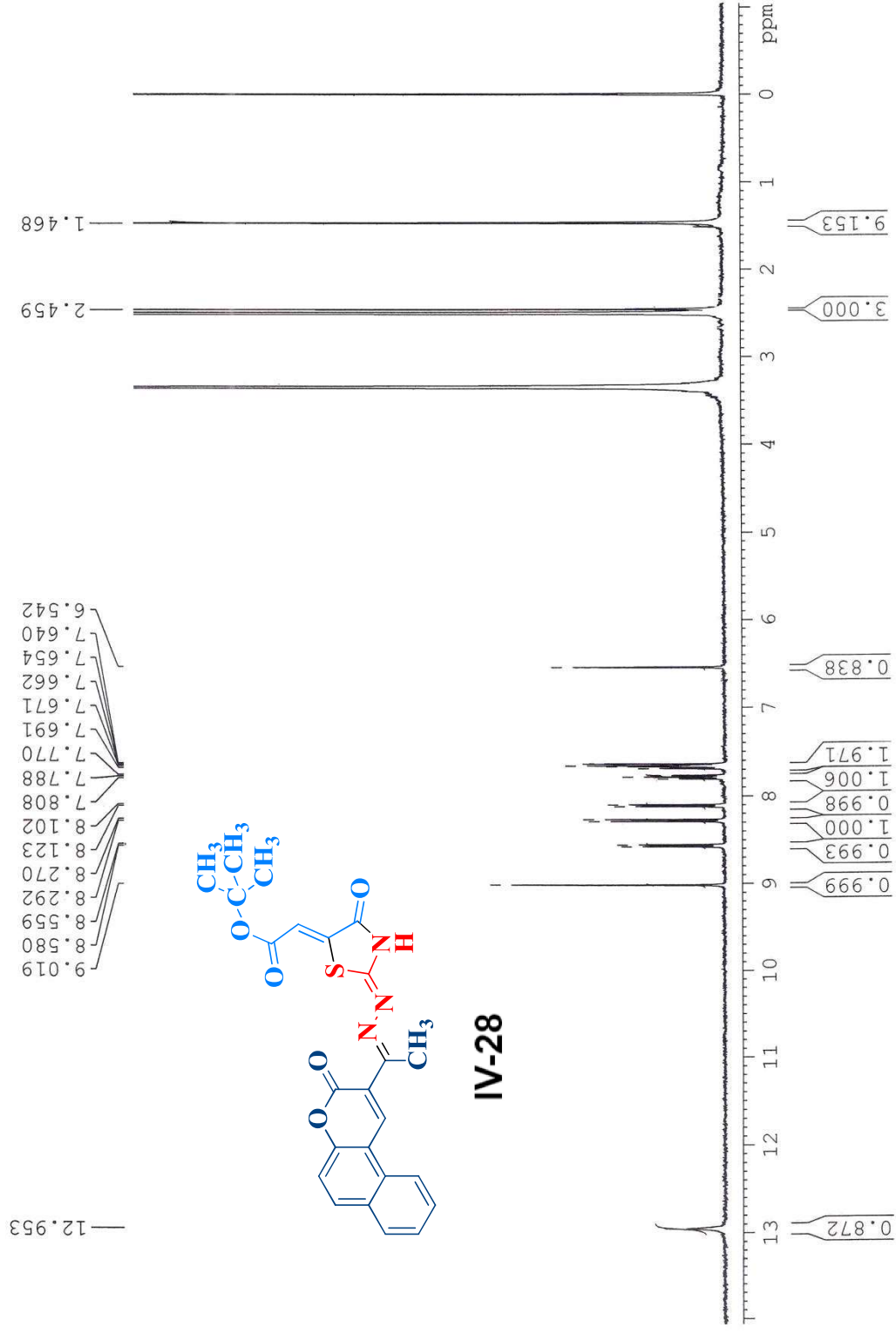
MB
1H NMR IN DMSO-D6



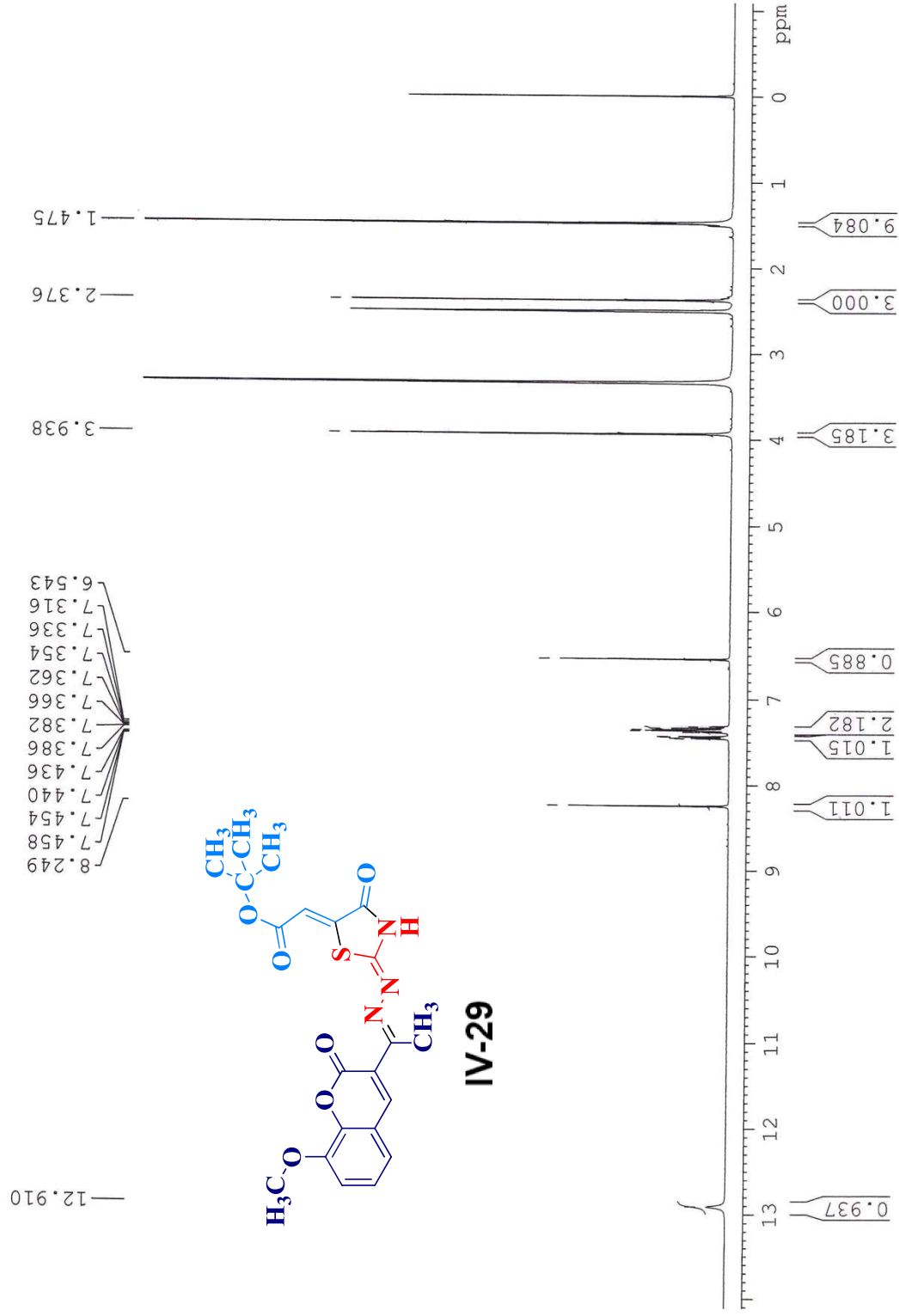
DB+DTBAD
1H NMR IN DMSO-D6



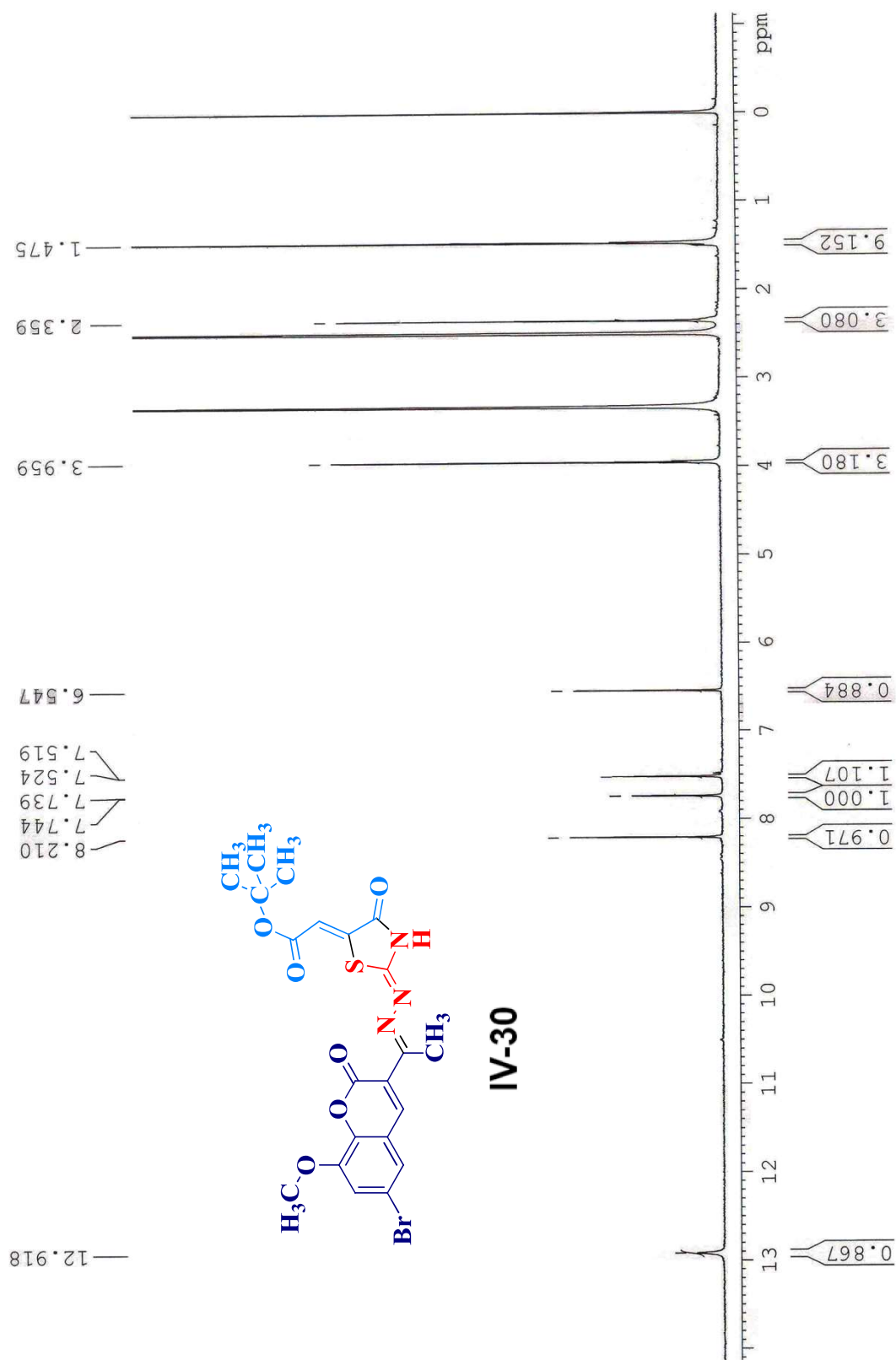
NAP
¹H NMR IN DMSO-D6



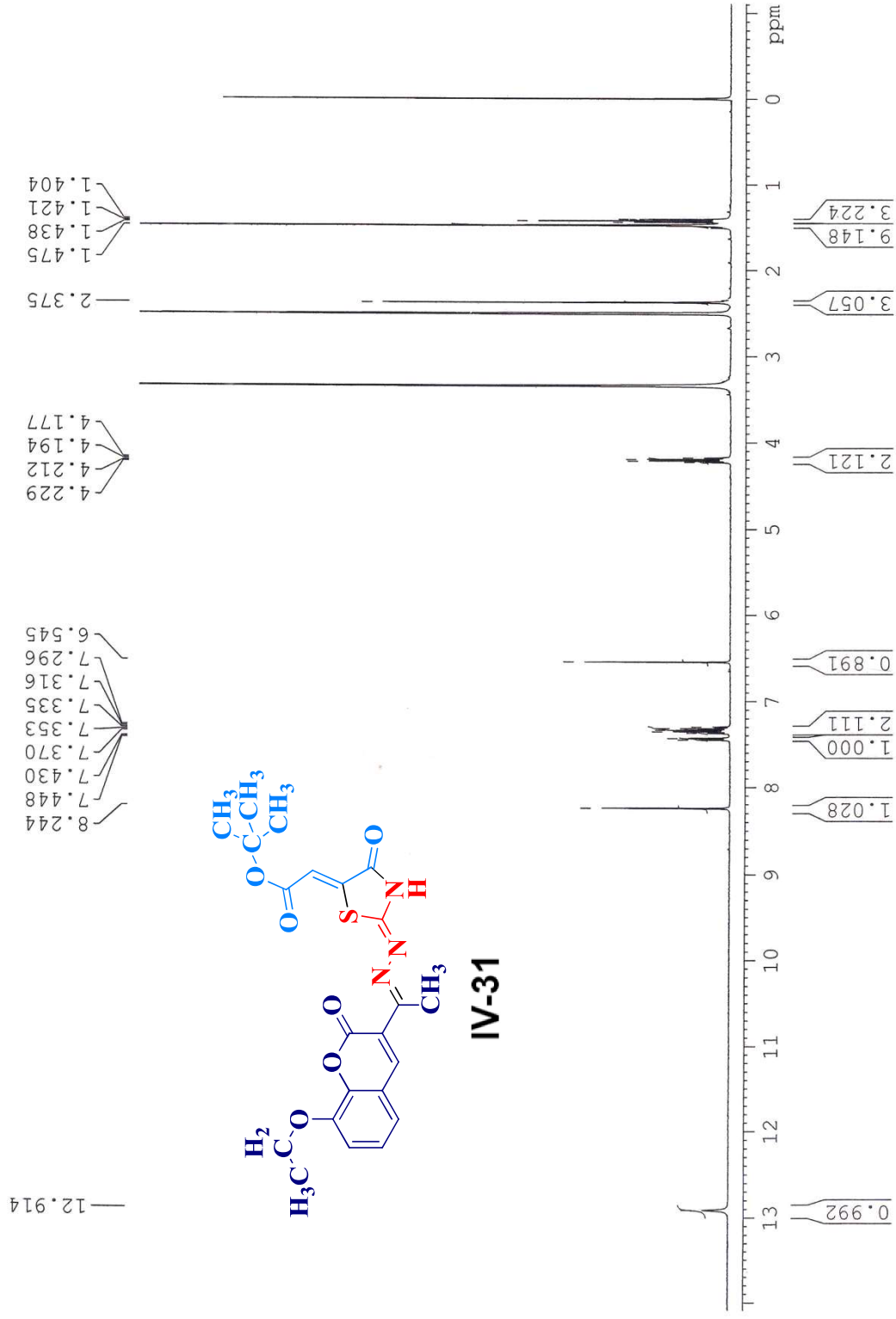
OV
¹H NMR IN DMSO-D₆



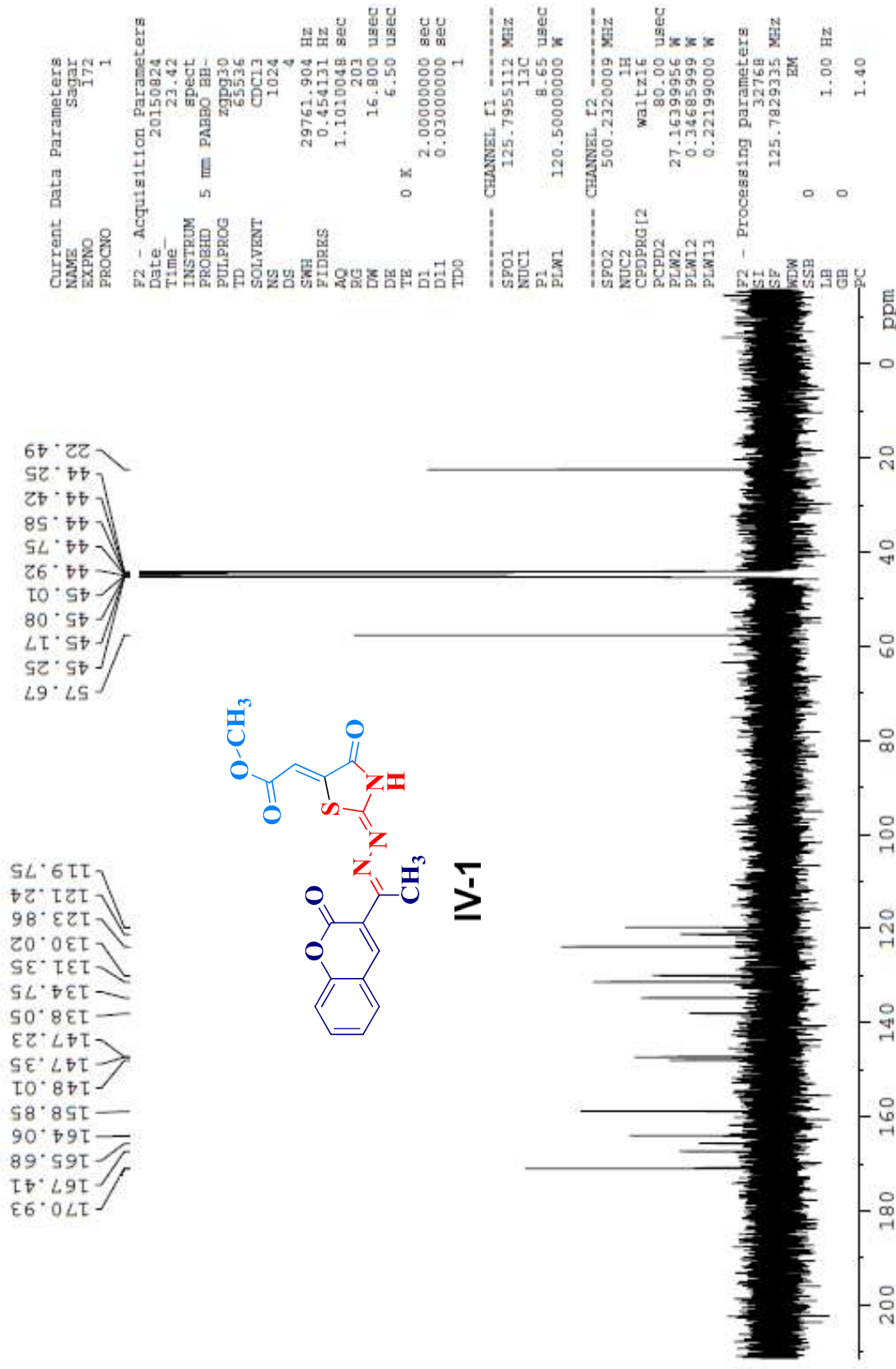
Br+OV+DTB
¹H NMR IN DMSO-D₆



DET
¹H NMR IN DMSO-D6



KMS-D-SBM
 Cl3CPD CDCl3 {C:\Bruker\TopSpin3.2} nmr 40

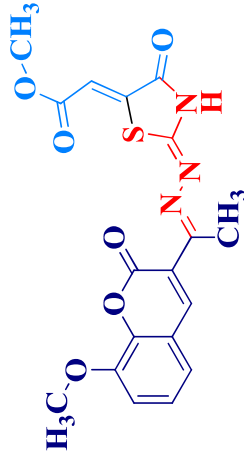


OV-DMAD
13C NMR IN DMSO-D6

165.797
158.488
146.283
142.896
142.188
126.119
124.752
122.430
120.438
119.171
114.985
114.501

56.147
52.416

17.134

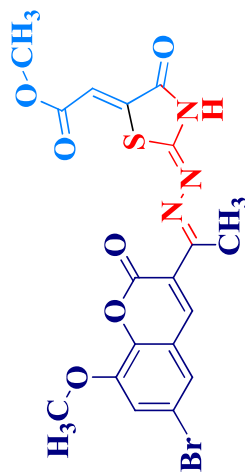


IV-7



BroVDM
 C13CPD DMSO {C:\Bruker\TopSpin3.2} nmr 42

164.16
 158.65
 147.69
 142.51
 141.46
 141.32
 127.40
 122.94
 122.53
 121.25
 120.98
 117.60
 116.67
 116.52



IV-8

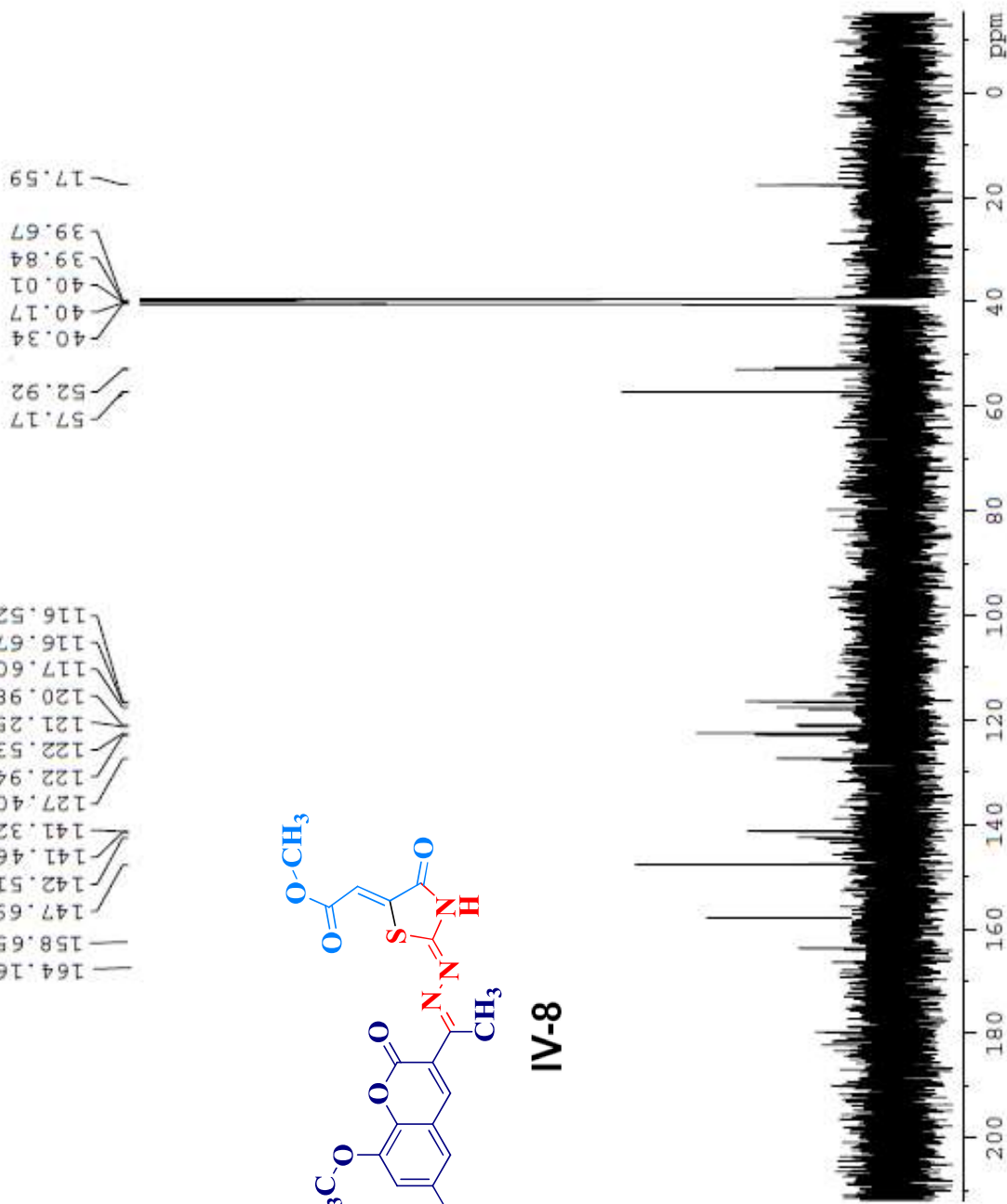
Current Data Parameters
 NAME Sagar
 EXPNO 176
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150825
 Time_ 2.33
 INSTRUM spect
 PROBHD 5 mm PABBO BB
 PULPROG zgpg30
 TD 65536
 SOLVENT DMSO
 NS 1024
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010048 sec
 RG 203
 DW 16.800 usec
 DE 6.50 usec
 TE 0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

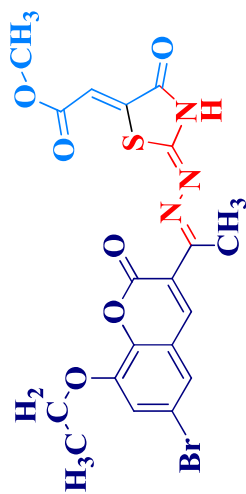
----- CHANNEL f1 -----
 SFO1 125.7955112 MHz
 NUC1 13C
 P1 8.65 usec
 PLW1 120.50000000 W

----- CHANNEL f2 -----
 SFO2 500.2320009 MHz
 NUC2 1H
 CPDPRG2 waitz16
 PCPD2 80.00 usec
 PLW2 27.16399956 W
 PLW12 0.34685999 W
 PLW13 0.22199000 W

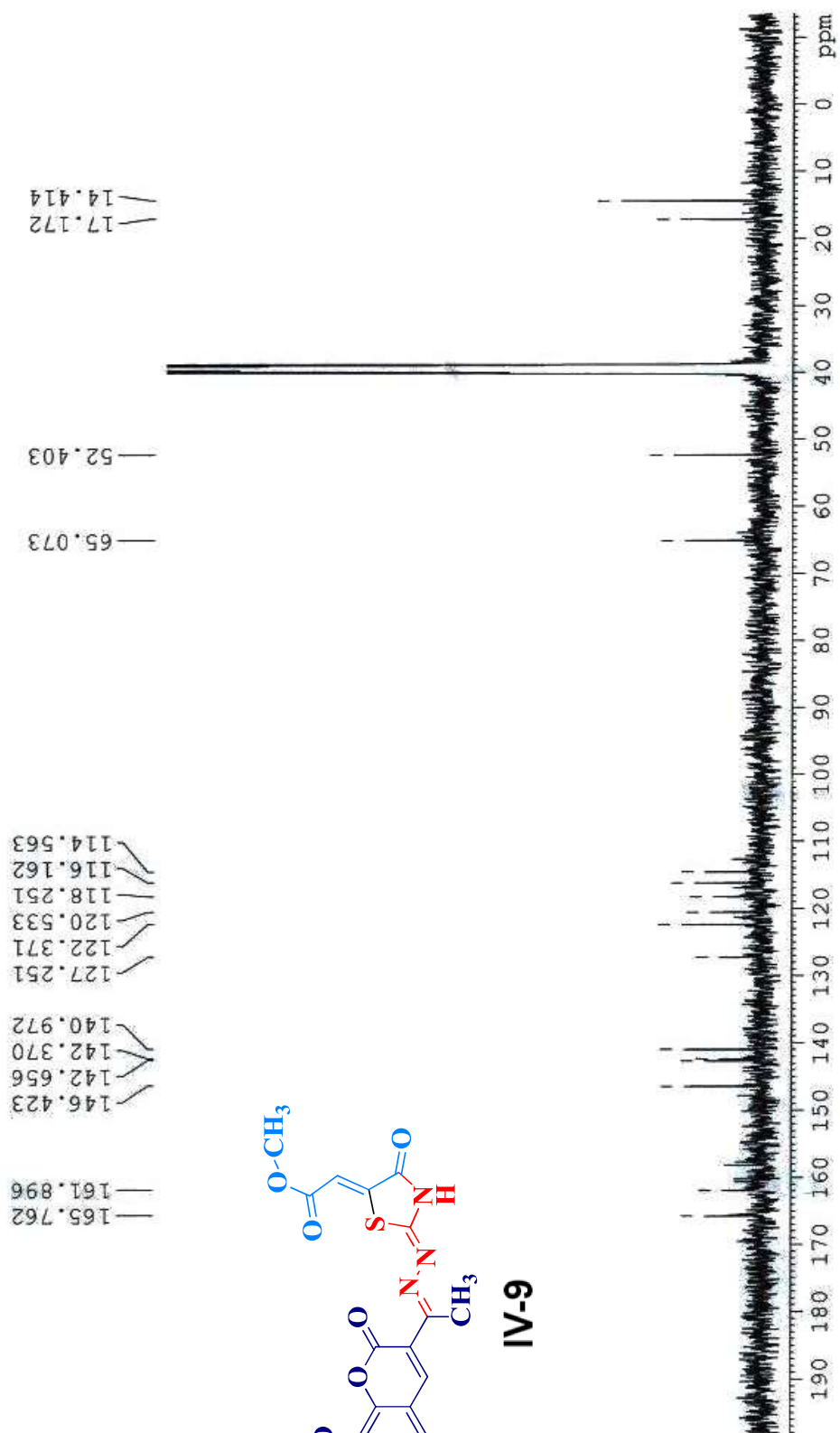
F2 - Processing parameters
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 SF 125.7829335 MHz
 WDW 0
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



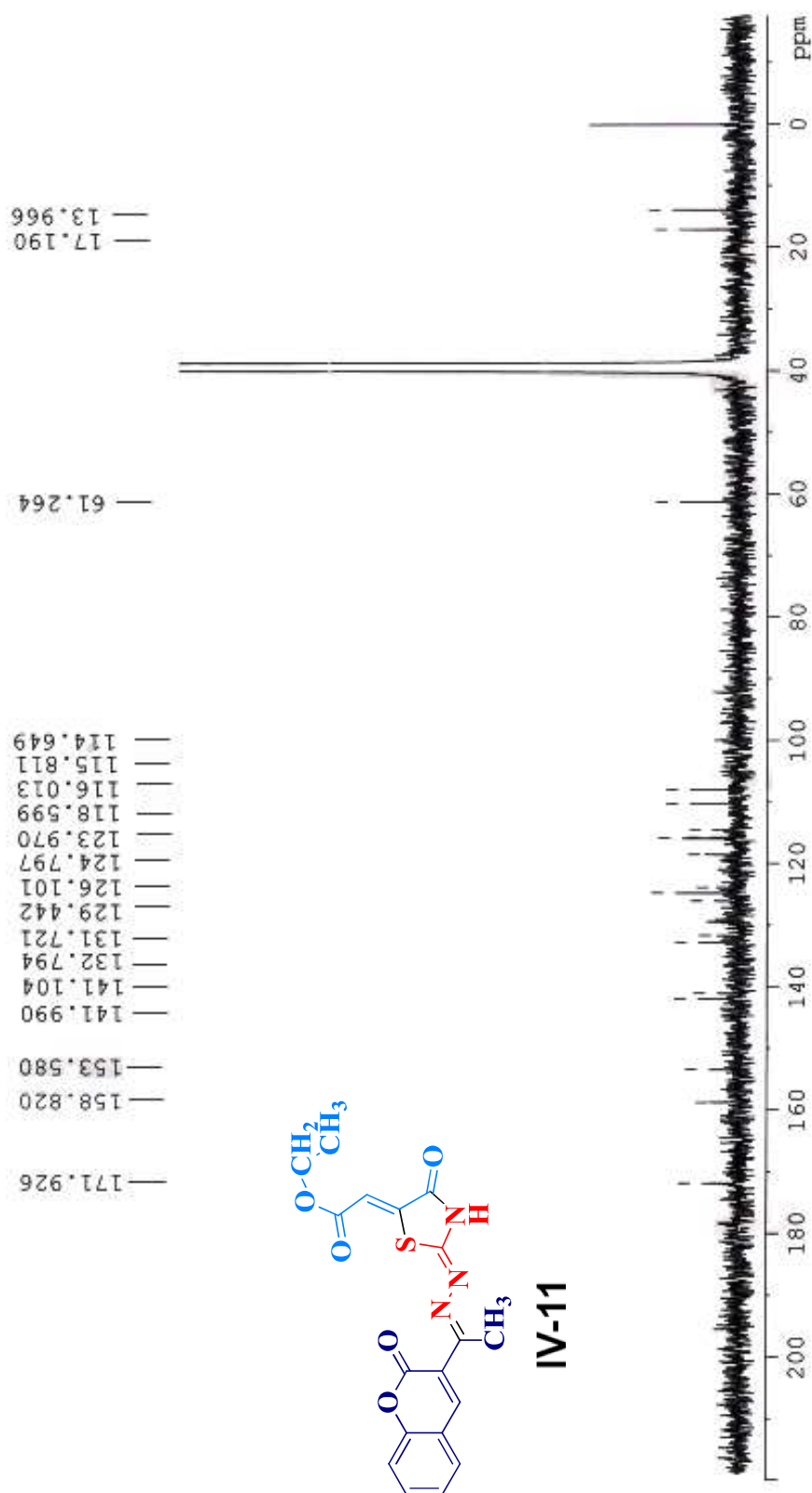
BROET-DM
¹³C NMR IN DMSO-D₆



IV-9



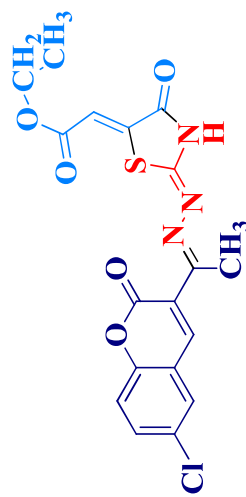
DEAD
¹³C NMR IN DMSO-D6



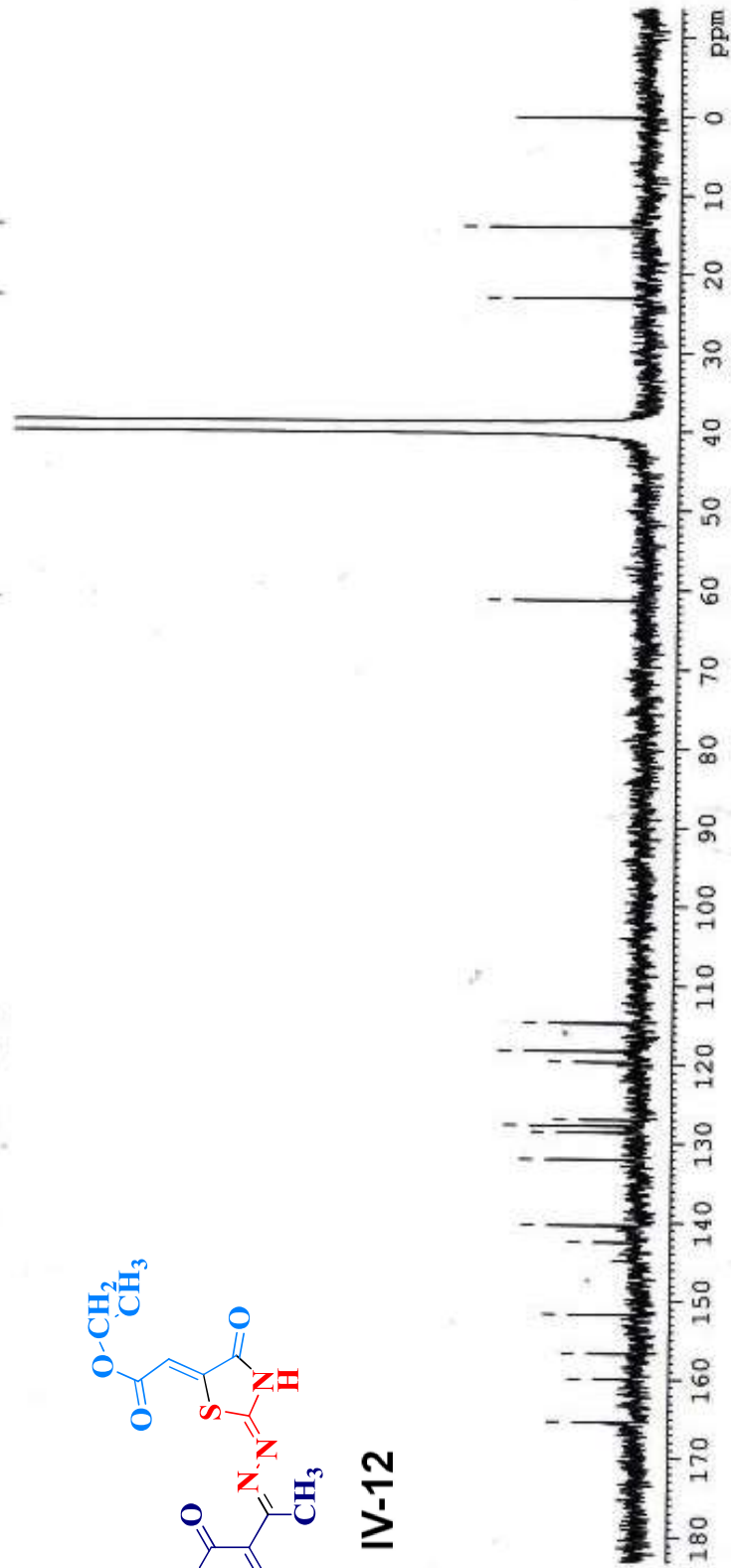
DE-MC
 13C NMR IN DMSO-D6

165.375
 159.915
 156.748
 151.724
 142.498
 140.363
 132.082
 128.609
 127.772
 127.012
 119.620
 118.265
 114.730

61.331
 23.046
 14.010



IV-12



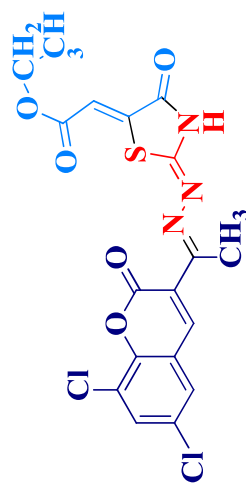
DE-DC
¹³C NMR IN DMSO-D₆

— 165.334
 — 156.710
 — 151.697
 — 142.475
 — 140.333
 — 132.043
 — 128.574
 — 128.305
 — 127.740
 — 126.974
 — 119.592
 — 118.230
 — 114.678

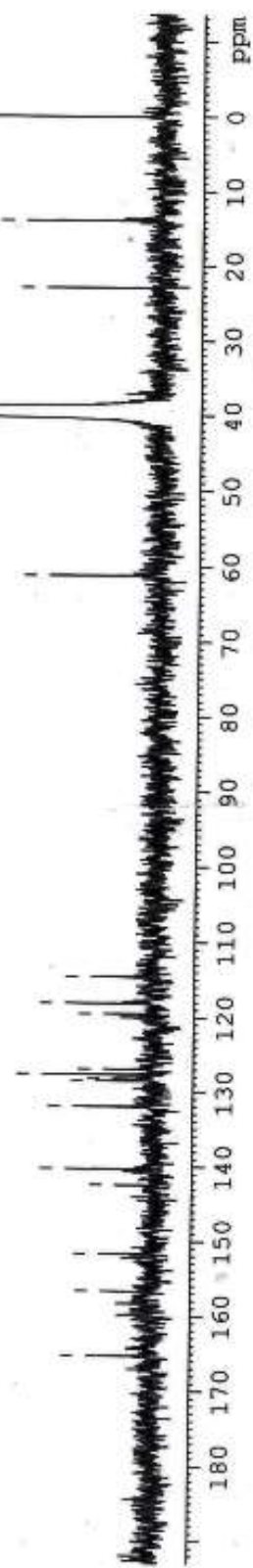
— 61.287

— 23.016

— 13.986

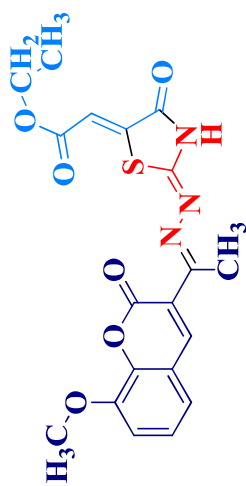


IV-13

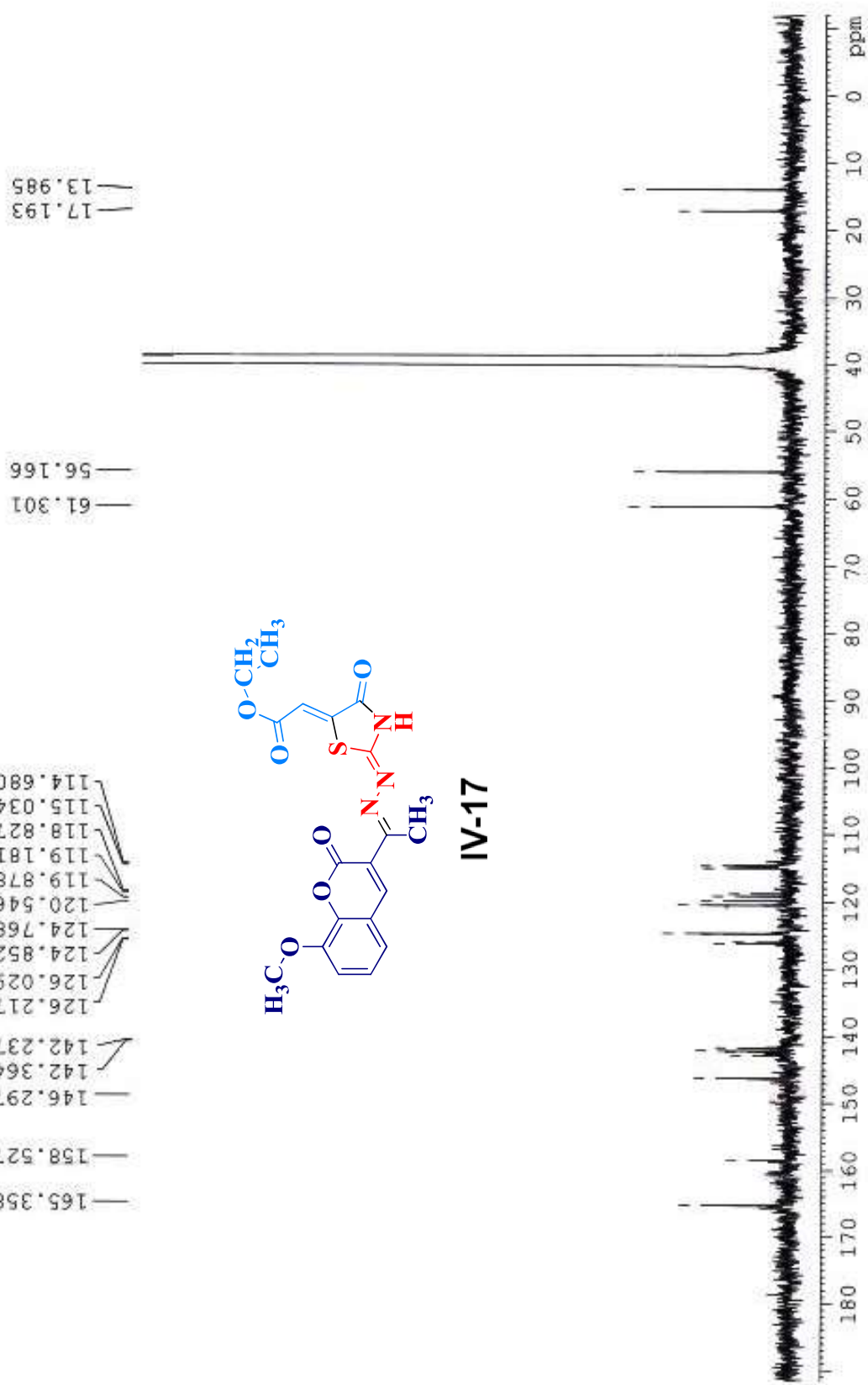


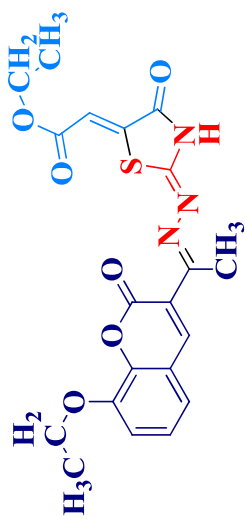
OV-DE
¹³C NMR IN DMSO-D6

165.358
 158.527
 146.297
 142.364
 142.237
 126.217
 126.029
 124.852
 124.768
 120.546
 119.878
 119.181
 118.827
 115.034
 114.680



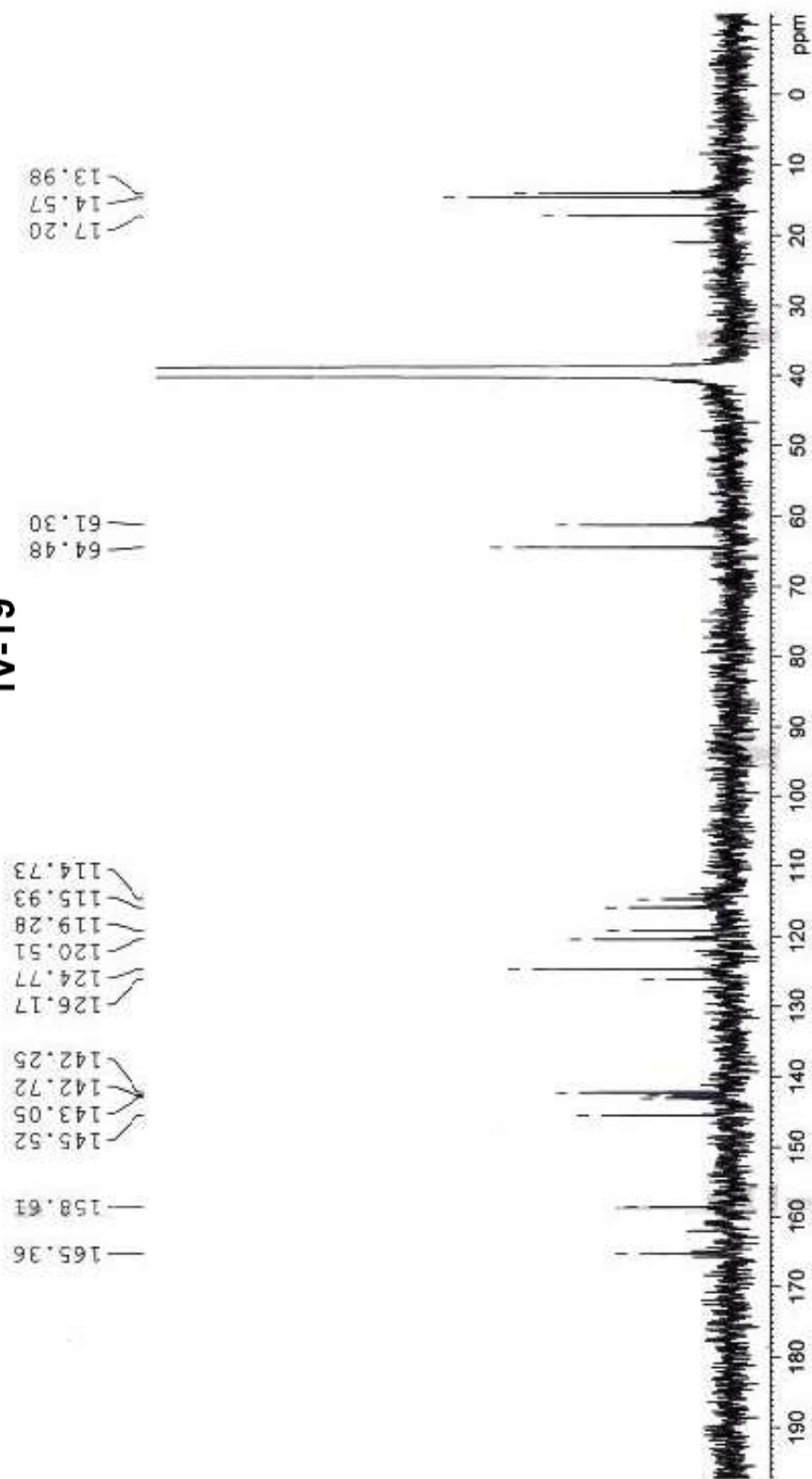
IV-17

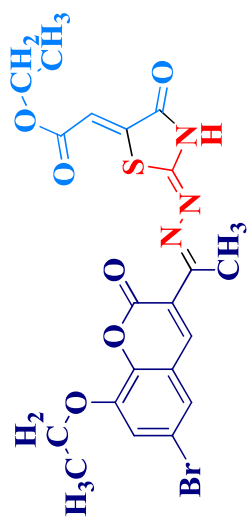




IV-19

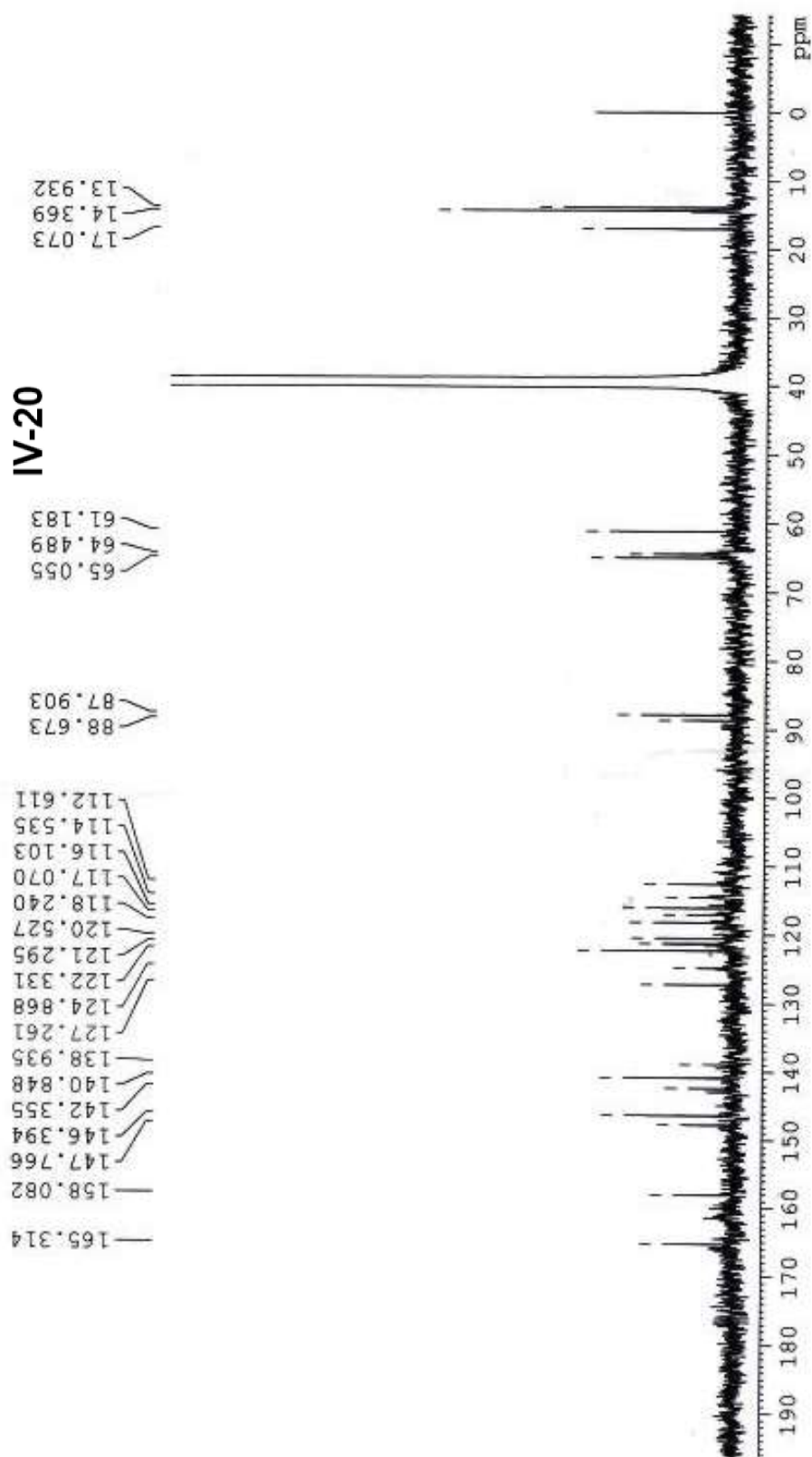
OET-DE
¹³C NMR IN DMSO-D₆





DE-BrOEt
¹³C NMR IN DMSO-D6

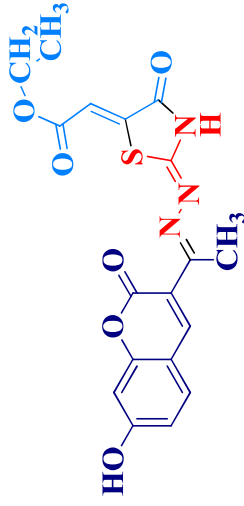
IV-20



C13CPD DMSO {C:\Bruker\TopSpin3.2} nmr 26

166.15
165.84
162.83
160.68
160.18
159.87
154.18
143.47
143.19
128.25
120.96
115.08
113.13
111.57
110.99

61.79
40.36
40.28
40.19
40.11
40.03
39.86
39.69
39.52
39.36
17.68
14.44



IV-21

Current Data Parameters
NAME Asha
EXPNO 6
PROCNO 1

F2 - Acquisition Parameters
Date_ 20151104
Time 9.59
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 411
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010048 sec
RG 203
DW 16.800 usec
DE 6.50 usec
TE 301.5 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

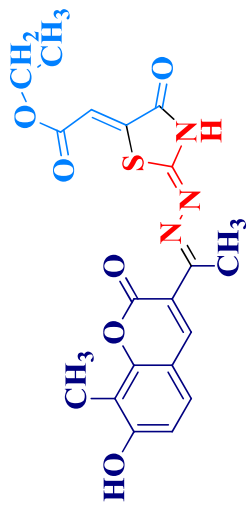
----- CHANNEL f1 -----
SFO1 125.7955112 MHz
NUC1 13C
P1 8.65 usec
PLW1 120.50000000 W

----- CHANNEL f2 -----
SFO2 500.2320009 MHz
NUC2 1H
PCPD2 80.00 usec
PLW2 27.16399956 W
PLW12 0.34685999 W
PLW13 0.22199000 W

F2 - Processing parameters
SI 32768
SF 125.7829335 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

ppm

-DE-MERE
¹³C NMR IN DMSO-D6

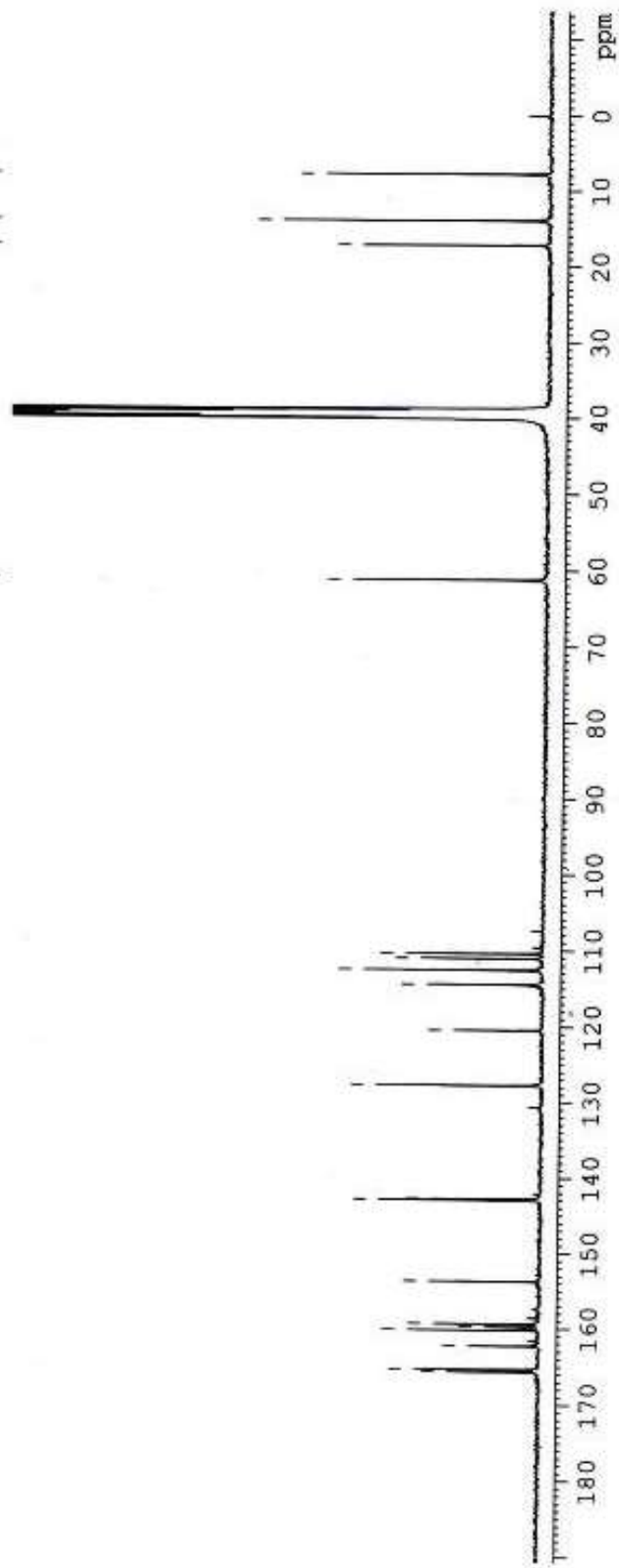


IV-22

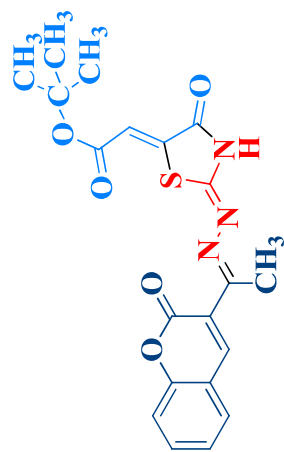
165.664
 165.353
 162.323
 160.199
 159.761
 159.359
 153.721
 142.969
 142.808
 127.788
 120.523
 114.518
 112.632
 111.096
 110.462

61.260

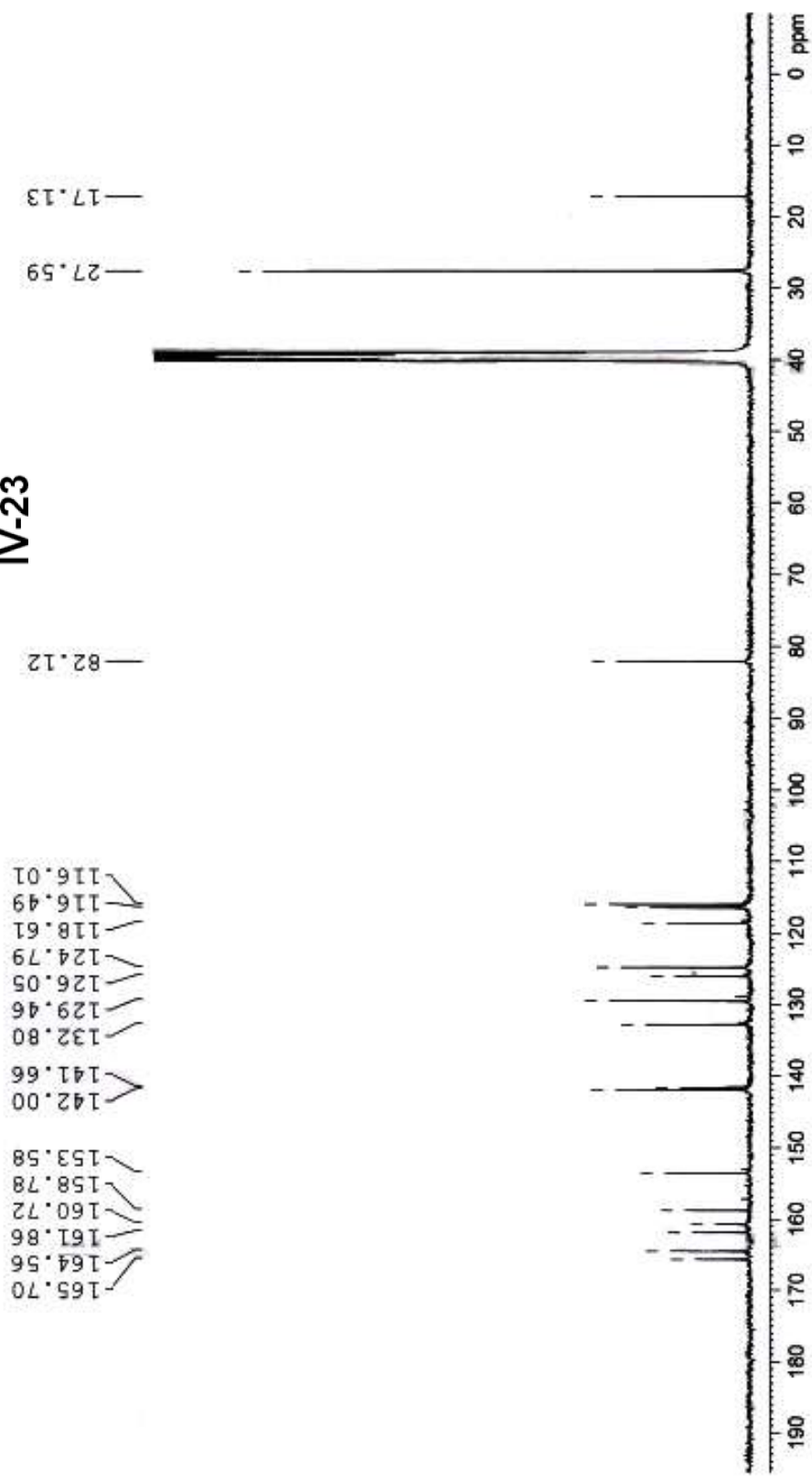
17.224
 13.978
 7.806



SDTB
¹³C NMR IN DMSO-D6



IV-23

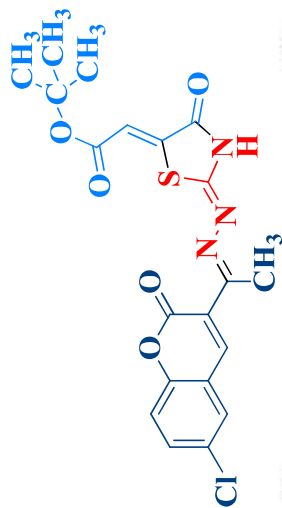


MC-DTB
13C NMR IN DMSO-D6

164.613
161.589
158.411
152.215
140.723
132.218
128.473
128.412
127.202
120.073
118.013
116.458

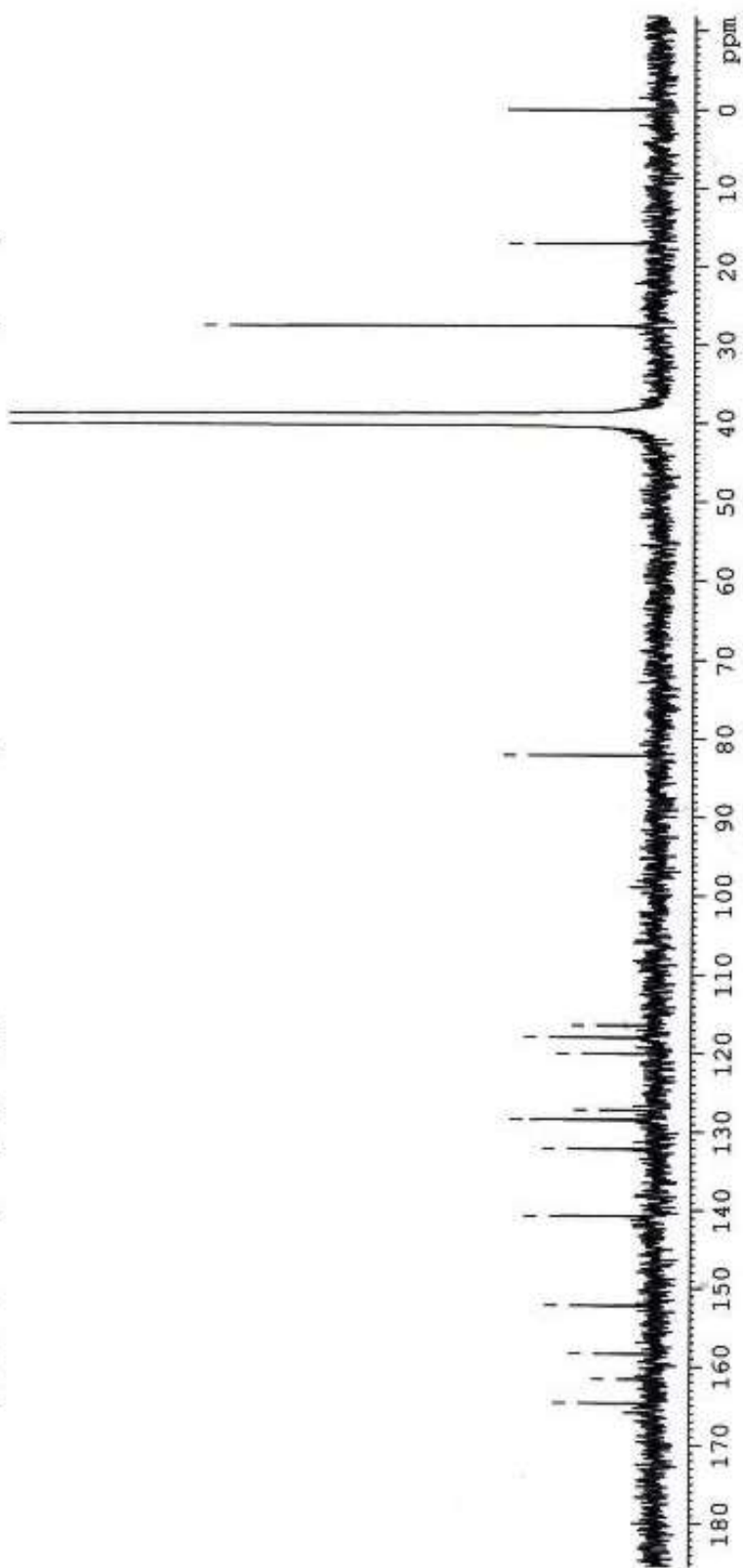
82.109

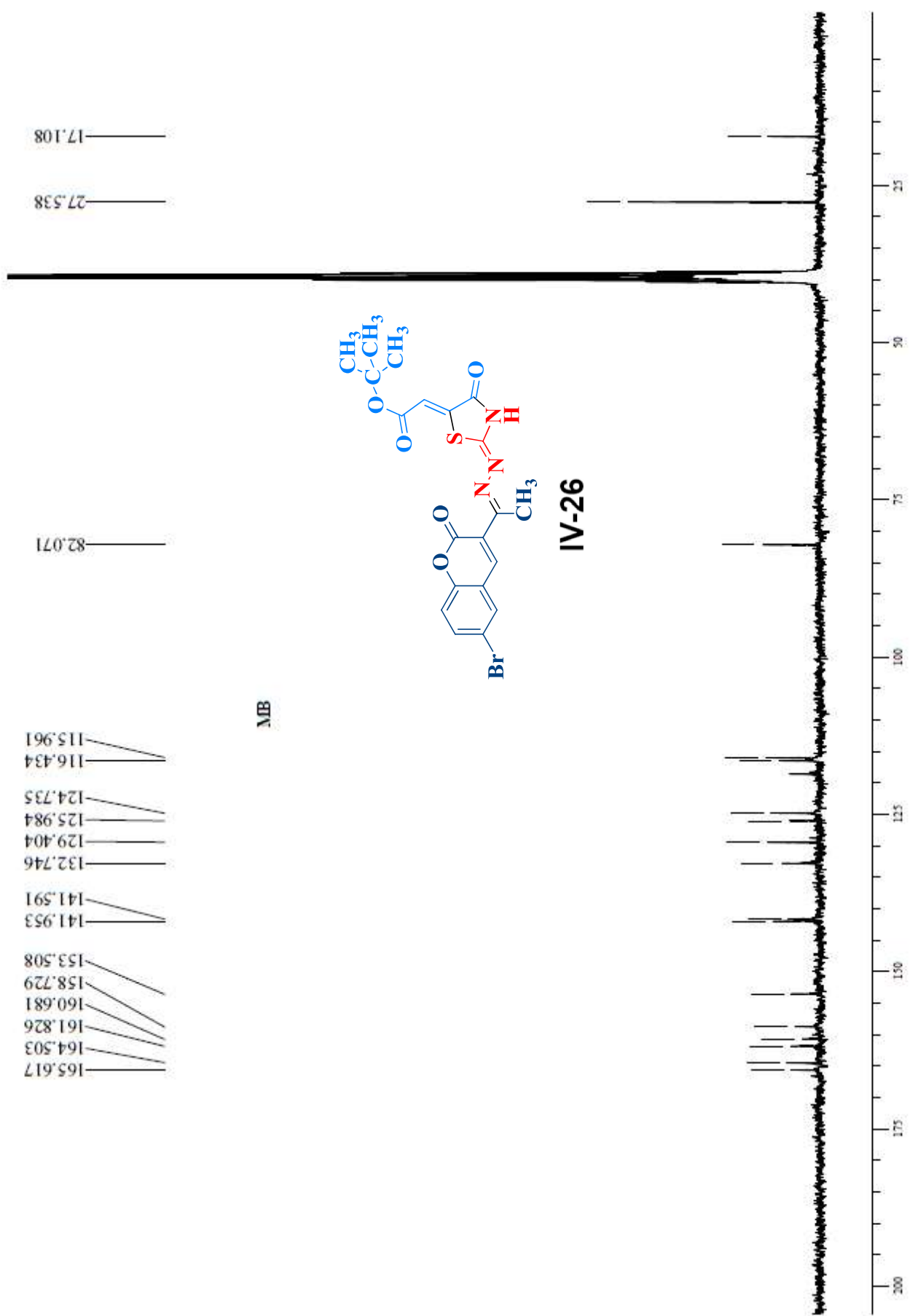
IV-24

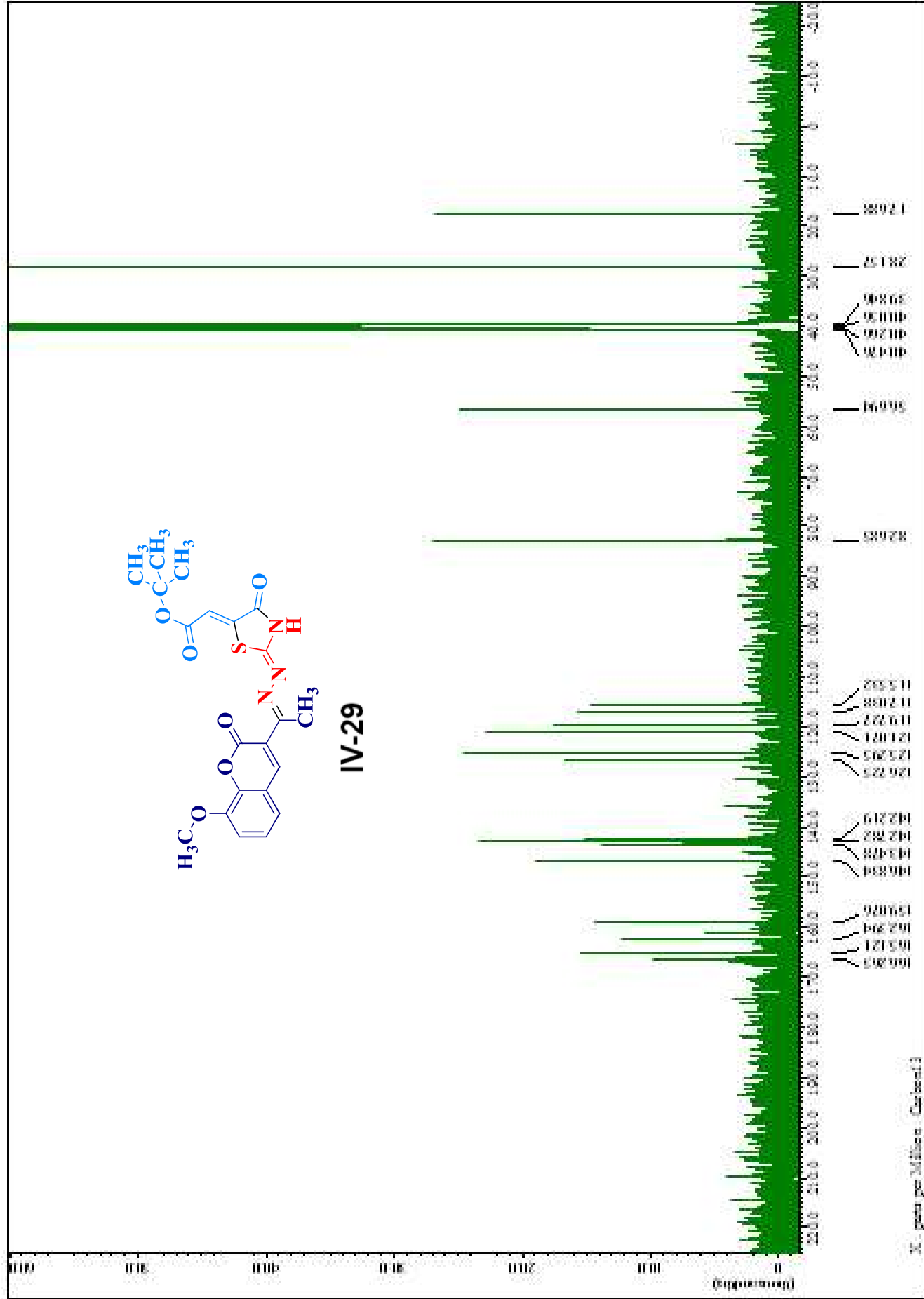


27.593

17.096







BrOV-DTB
¹³C NMR IN DMSO-D6

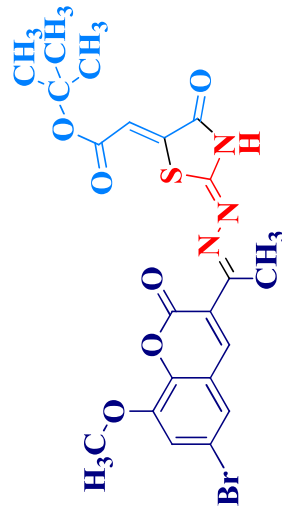
165.711
 164.566
 161.546
 161.059
 158.023
 147.156
 142.275
 141.671
 140.917
 127.251
 122.395
 120.459
 117.515
 116.479
 116.157

82.105

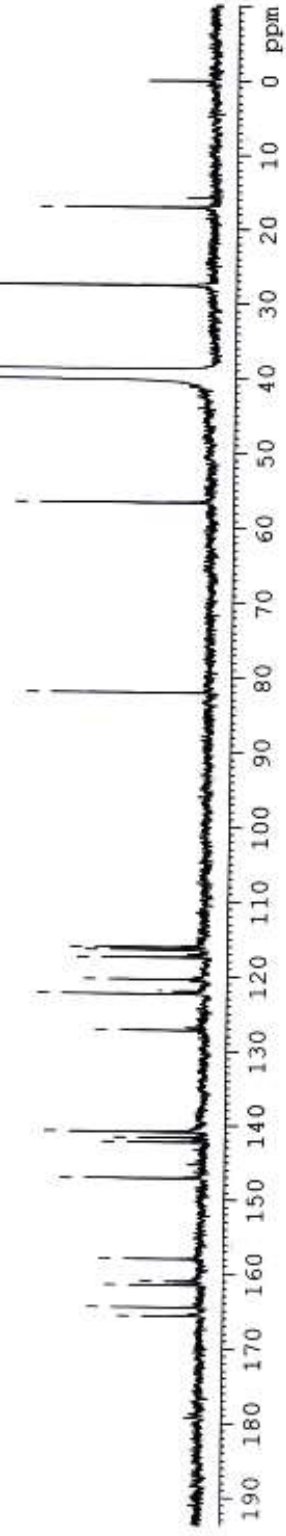
56.676

27.590

17.069



IV-30

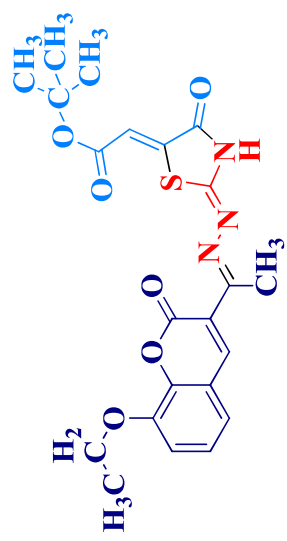


OV-OET+DTB
¹³C NMR IN DMSO-D6

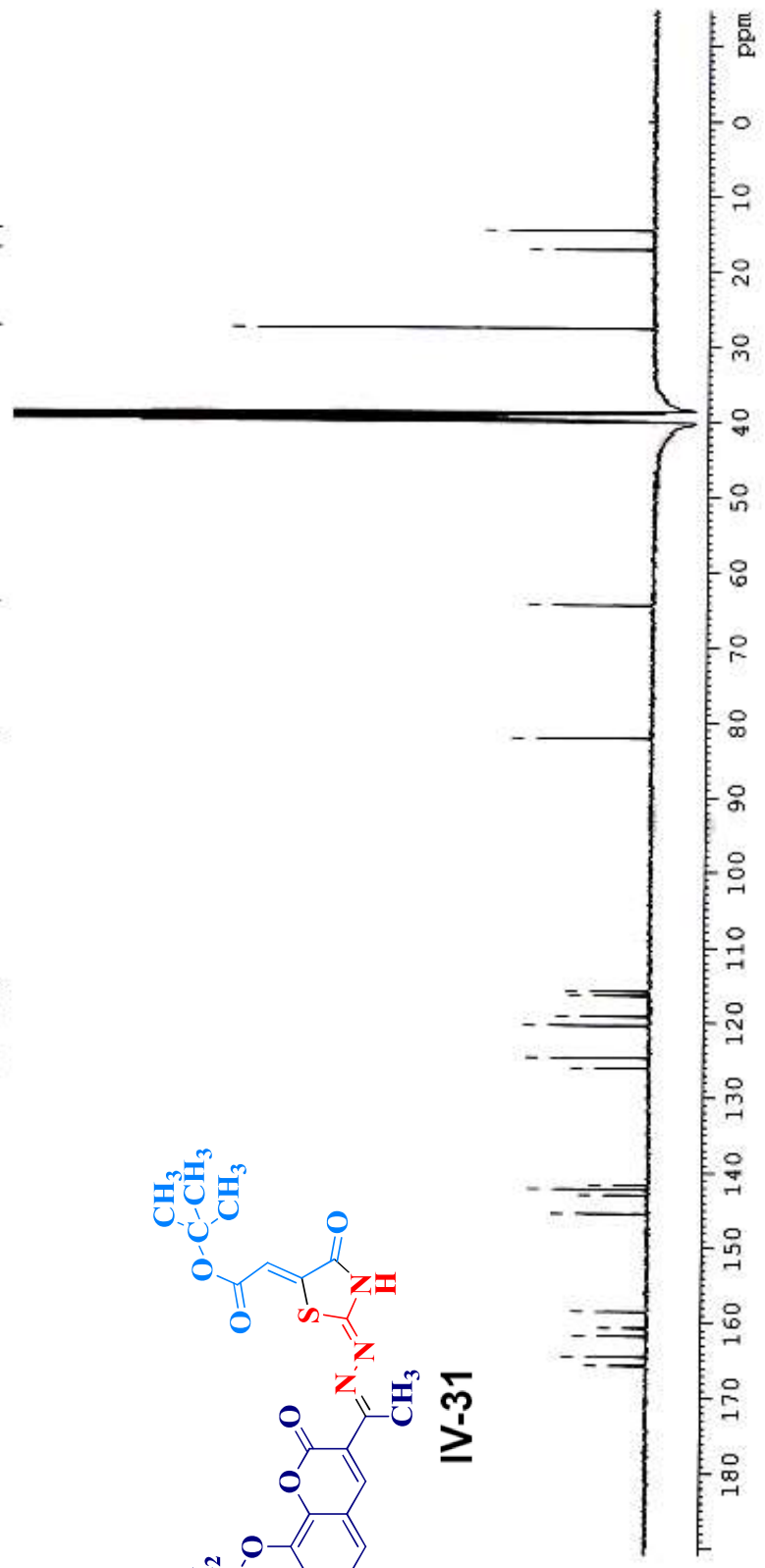
165.766
 164.592
 161.867
 160.814
 158.623
 145.532
 143.038
 142.269
 141.704
 126.146
 124.770
 120.498
 119.285
 116.510
 115.894

82.162
 64.473

27.618
 17.157
 14.584



IV-31



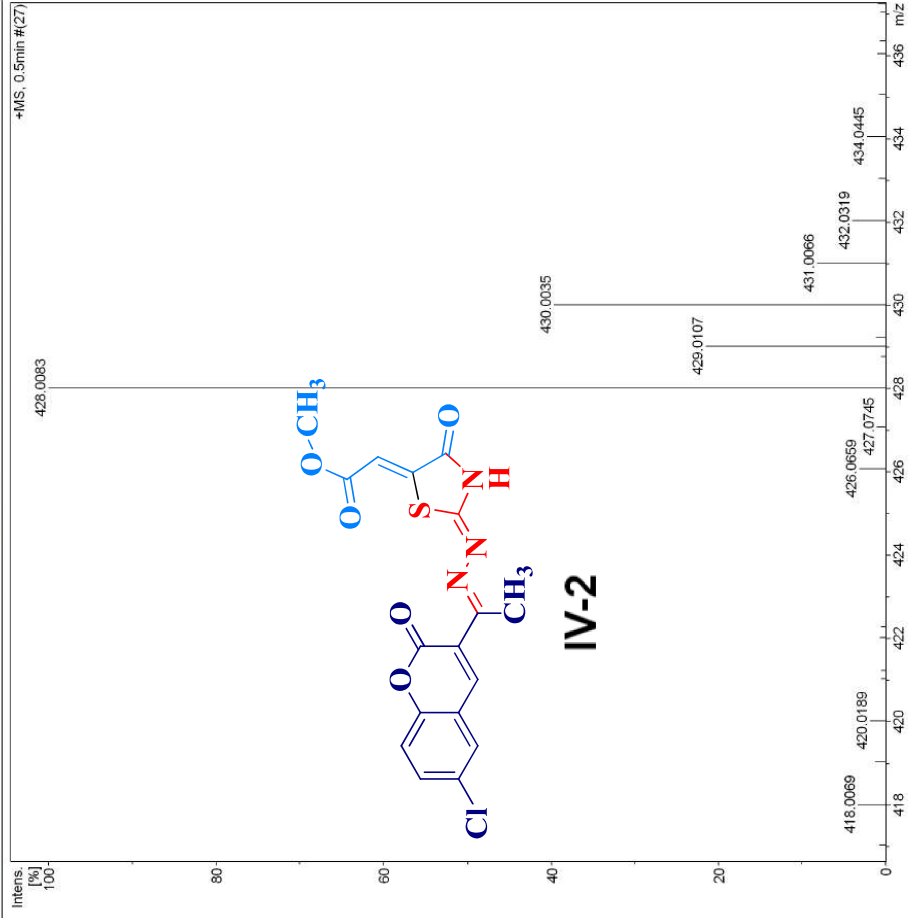
Display Report

Analysis Info

Analysis Name D:\Data\prof v k gupta\NA-242+Zn5.d Acquisition Date 3/19/2015 10:06:45 AM
Method tune_low.m Operator IIT ROORKEE
Sample Name NA-242+Zn5 Instrument microTOF-Q II 10328
Comment

Acquisition Parameter

Source Type ESI Ion Polarity Positive Set Nebulizer 0.4 Bar
Focus Not active Set Capillary 4500 V Set Dry Heater 180 °C
Scan Begin 250 m/z Set End Plate Offset -500 V Set Dry Gas 4.0 l/min
Scan End 1100 m/z Set Collision Cell RF 150.0 Vpp Set Divert Valve Source





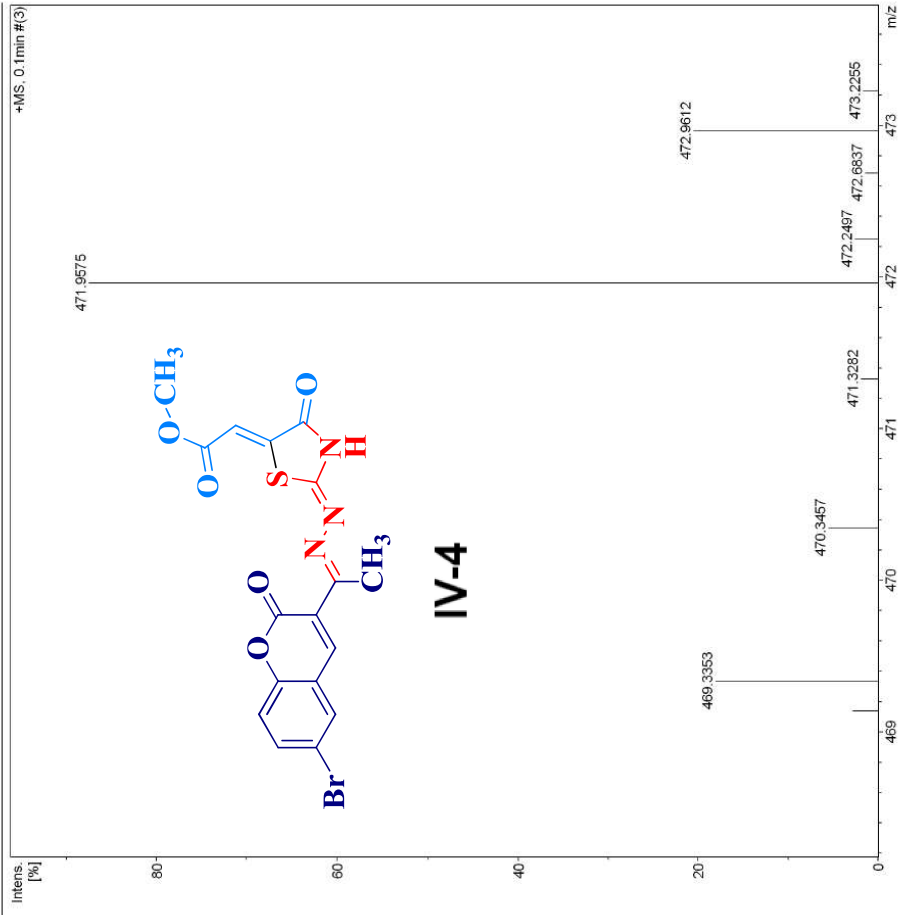
Display Report

Analysis Info

Analysis Name D:\Data\prof v k gupta\NA-242+Cu6.1.d Acquisition Date 3/19/2015 11:00:41 AM
Method tune_low.m Operator IIT ROORKEE
Sample Name NA-242+Cu6.1 Instrument micrOTOF-Q II 10328
Comment

Acquisition Parameter

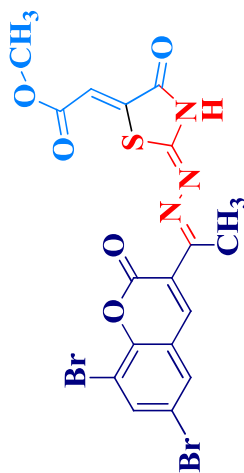
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	250 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	1100 m/z	Set Collision Cell RF	150.0 Vpp	Set Divert Valve	Source



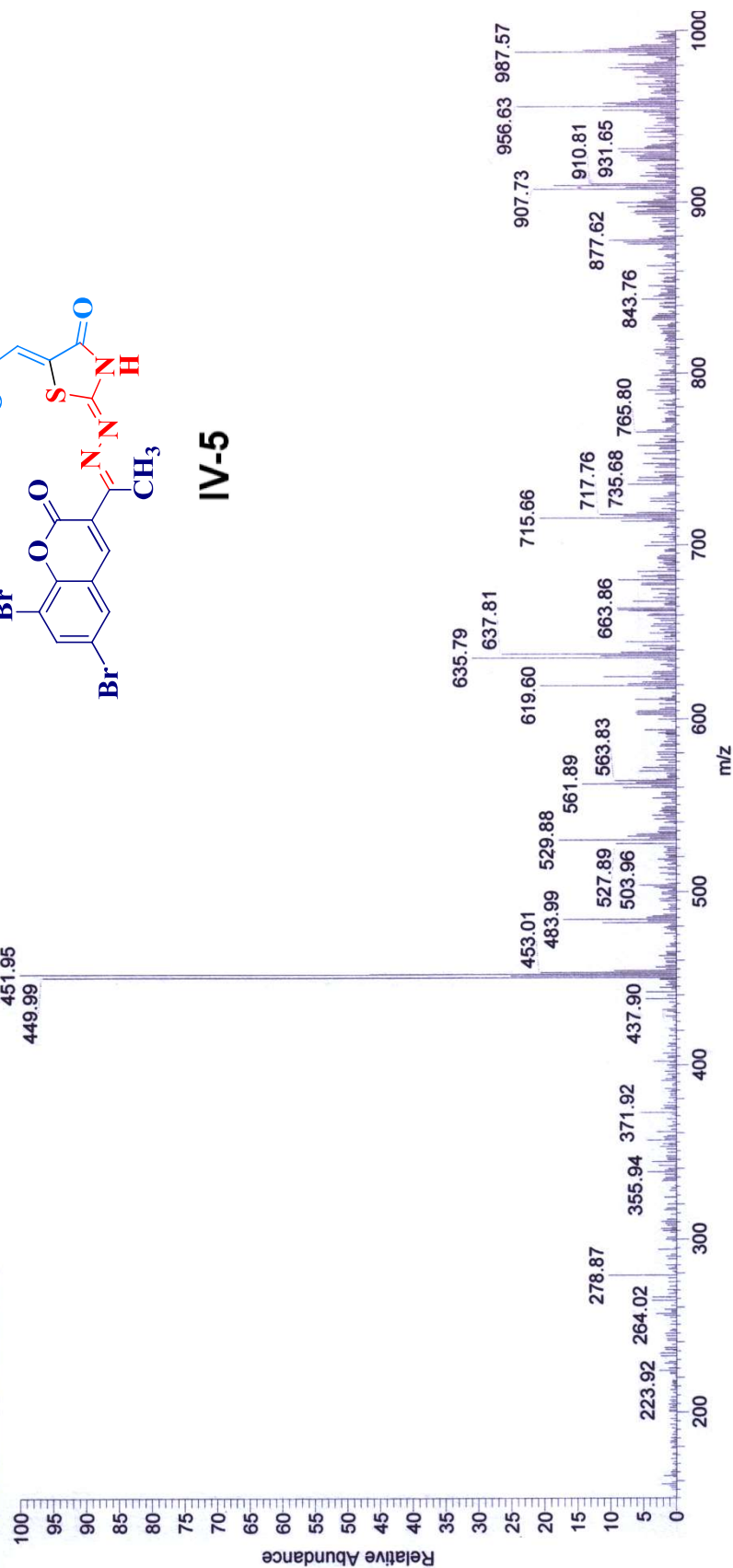
SAIF, CSIR-CDRI, Lucknow

Data File: 15109APR46
Original Data Path: 15109APR46 RAW
Current Data Path: C:\XCALIBUR\DATA\APR2015\08APR2015\1
Sample ID: RM-DBDM
Acquisition Date: 04/09/15 13:43:46

15109APR46 #19-46 RT: 0.29-0.71 AV: 28 SB: 2 0.00 , 0.00 NL: 1.87E6
T: + c ESI Full ms [150.00-1000.00]



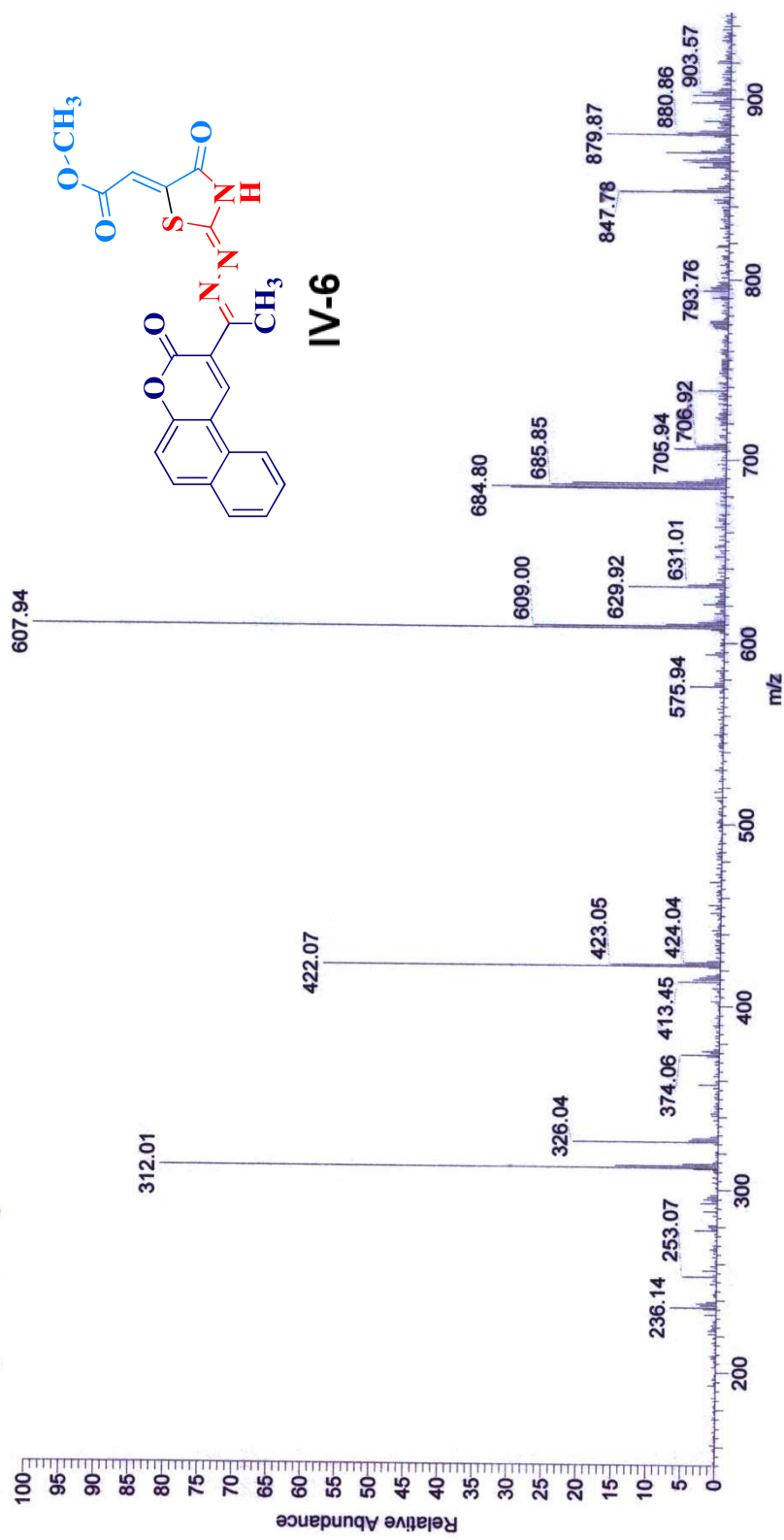
IV-5



ION TRAP LCQ ADVANTAGE MAX
THERMO ELECTRON CORPORATION

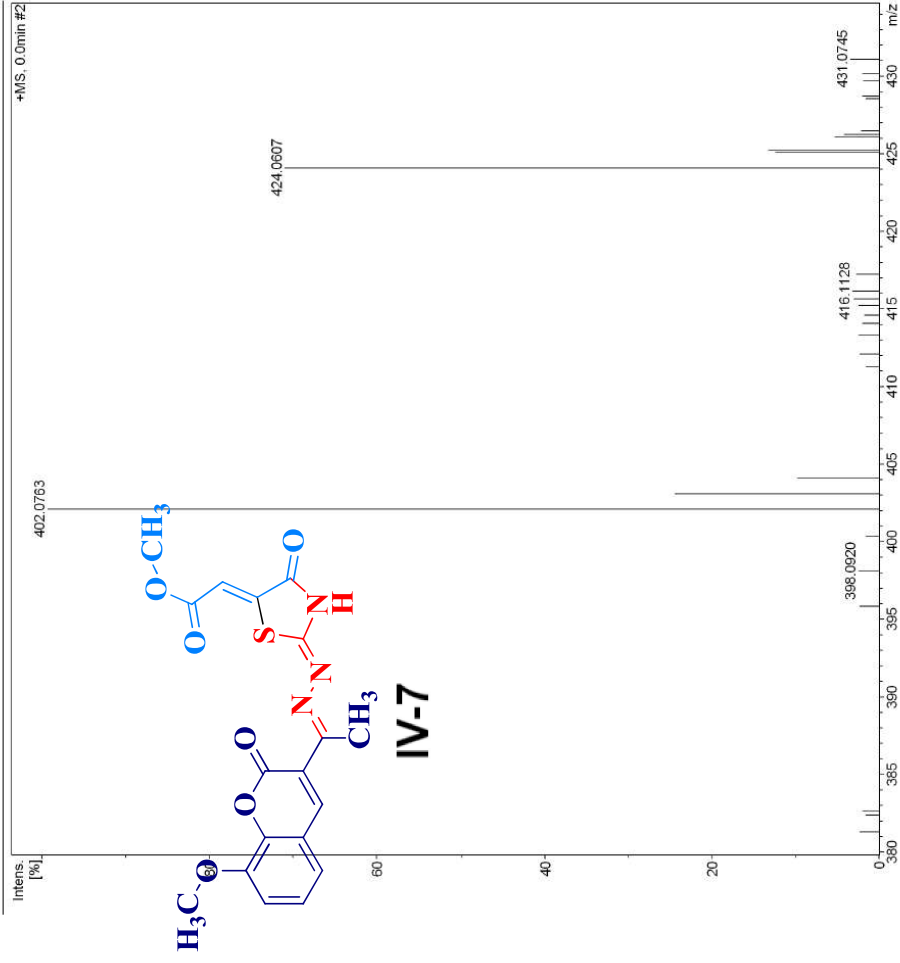
Data File: 15109APR47
Original Data Path: 15109APR47.RAW
Current Data Path: C:\XCALIBUR\DATA\PR201508APR2015\1
Sample ID: RM-NAPDM
Acquisition Date: 04/09/15 13:45:46

15109APR47 #19-46 RT: 0.29-0.69 AV: 28 SB: 2 0.00, 0.00 NL: 5.68E6
T: + c ESI Full ms [150.00-1000.00]



Display Report

Analysis Info	
Analysis Name	D:\Data\prof.v.k.gupta\NA-242+Fe5.d
Method	tune_low.m
Sample Name	NA-242+Fe5
Comment	
Acquisition Parameter	
Source Type	ESI
Focus	Not active
Scan Begin	250 m/z
Scan End	1100 m/z
Ion Optics	
Ion Polarity	Positive
Set Capillary	4500 V
Set End Plate Offset	-500 V
Set Collision Cell RF	150.0 Vpp
Source	
Set Nebulizer	0.4 Bar
Set Dry Heater	180 °C
Set Dry Gas	4.0 l/min
Set Divert Valve	Source



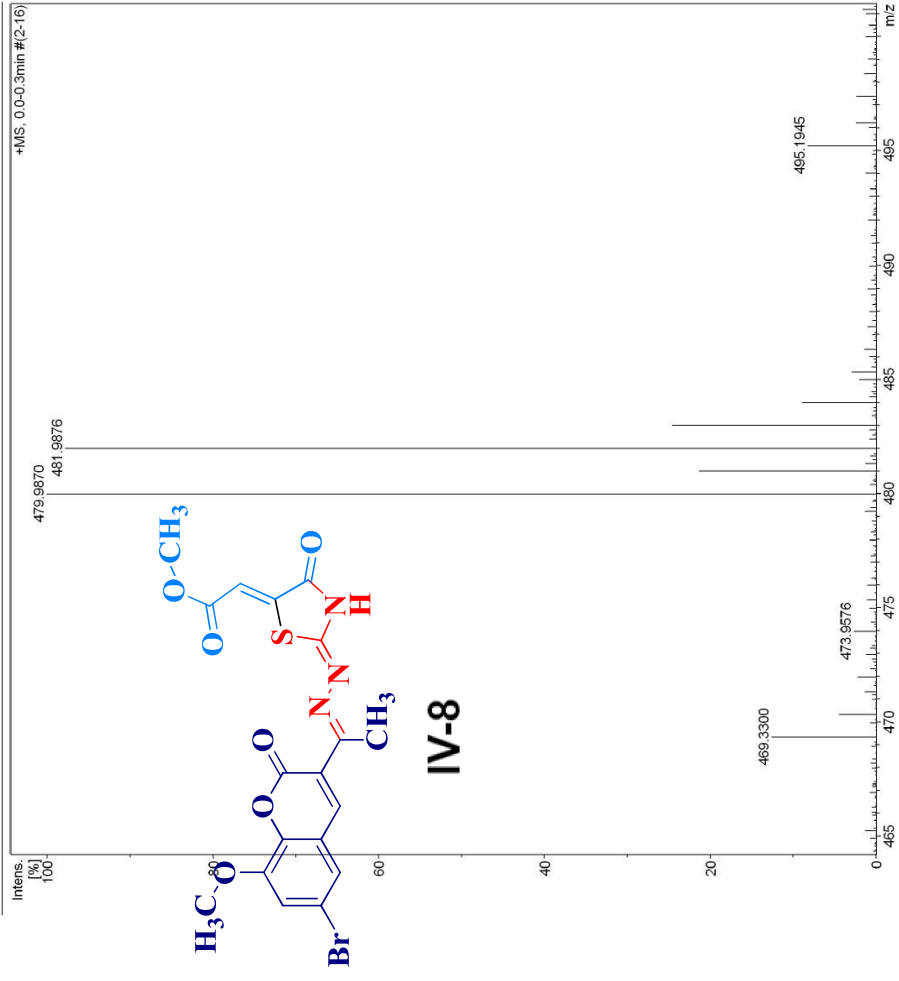
Display Report

Analysis Info

Analysis Name D:\Data\prof.v.k.gupta\NA-242+Fe6.2.d
Method tune_low.m
Sample Name NA-242+Fe6.2
Comment
Acquisition Date 3/19/2015 11:47:52 AM
Operator IIT ROORKEE
Instrument microTOF-Q II 10328

Acquisition Parameter

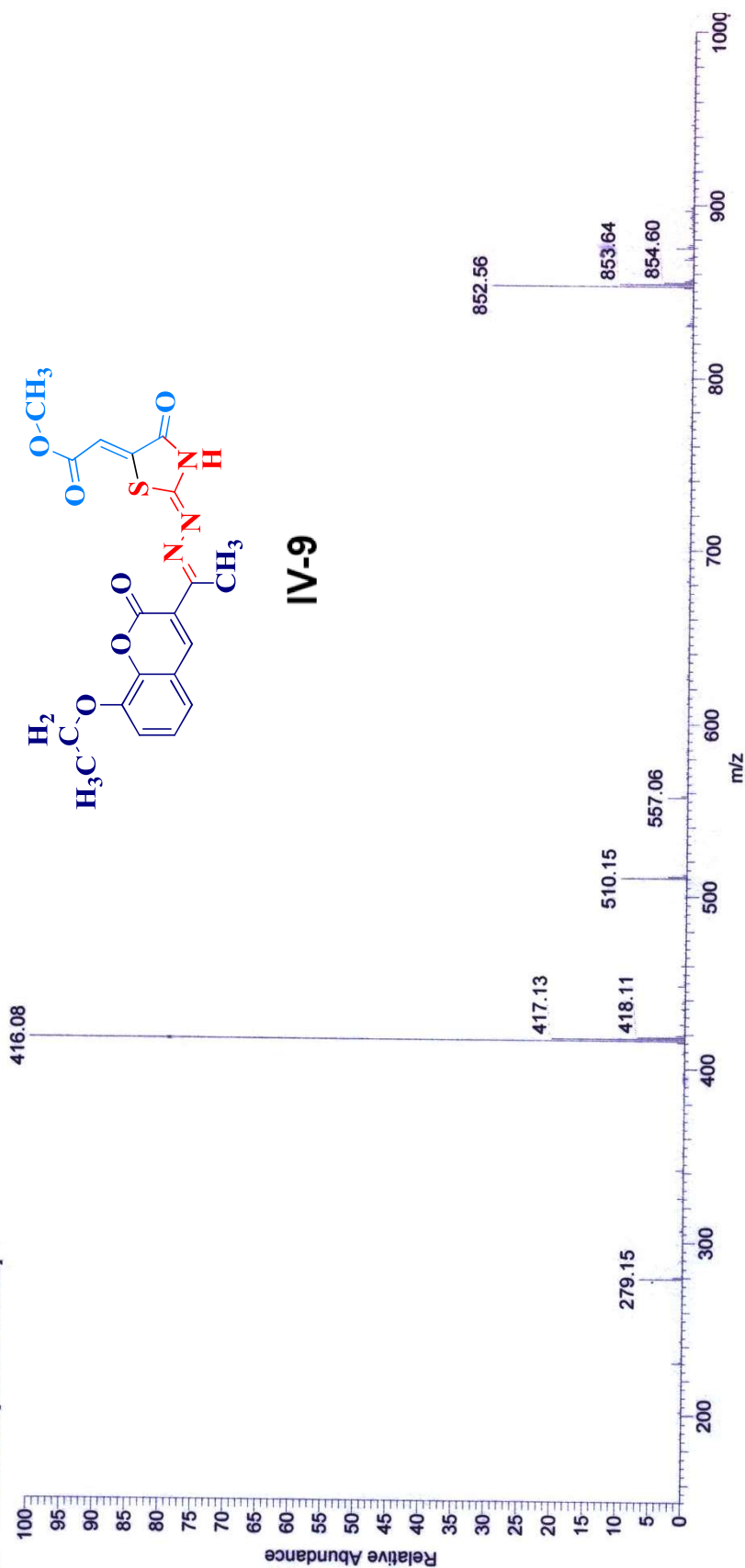
Source Type ESI
Focus Not active
Scan Begin 250 m/z
Scan End 1100 m/z
Ion Polarity Positive
Set Capillary 4500 V
Set End Plate Offset -500 V
Set Collision Cell RF 150.0 Vpp
Set Nebulizer 0.4 Bar
Set Dry Heater 180 °C
Set Dry Gas 4.0 l/min
Set Divert Valve Source



SAIF, CSIR-CDRI, Lucknow

Data File: 15115APR41
Original Data Path: 15115APR41_RAW
Current Data Path: C:\Xcalibur\data\APR2015\151APR2015\1
Sample ID: RM4-OETDM
Acquisition Date: 04/15/15 13:33:52

15115APR41 #19-47 RT: 0.30-0.70 AV: 29 SB: 2 0.00, 0.00 NL: 3.42E7
T: + c ESI Full ms [150.00-1000.00]

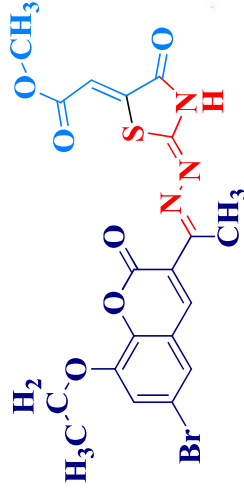
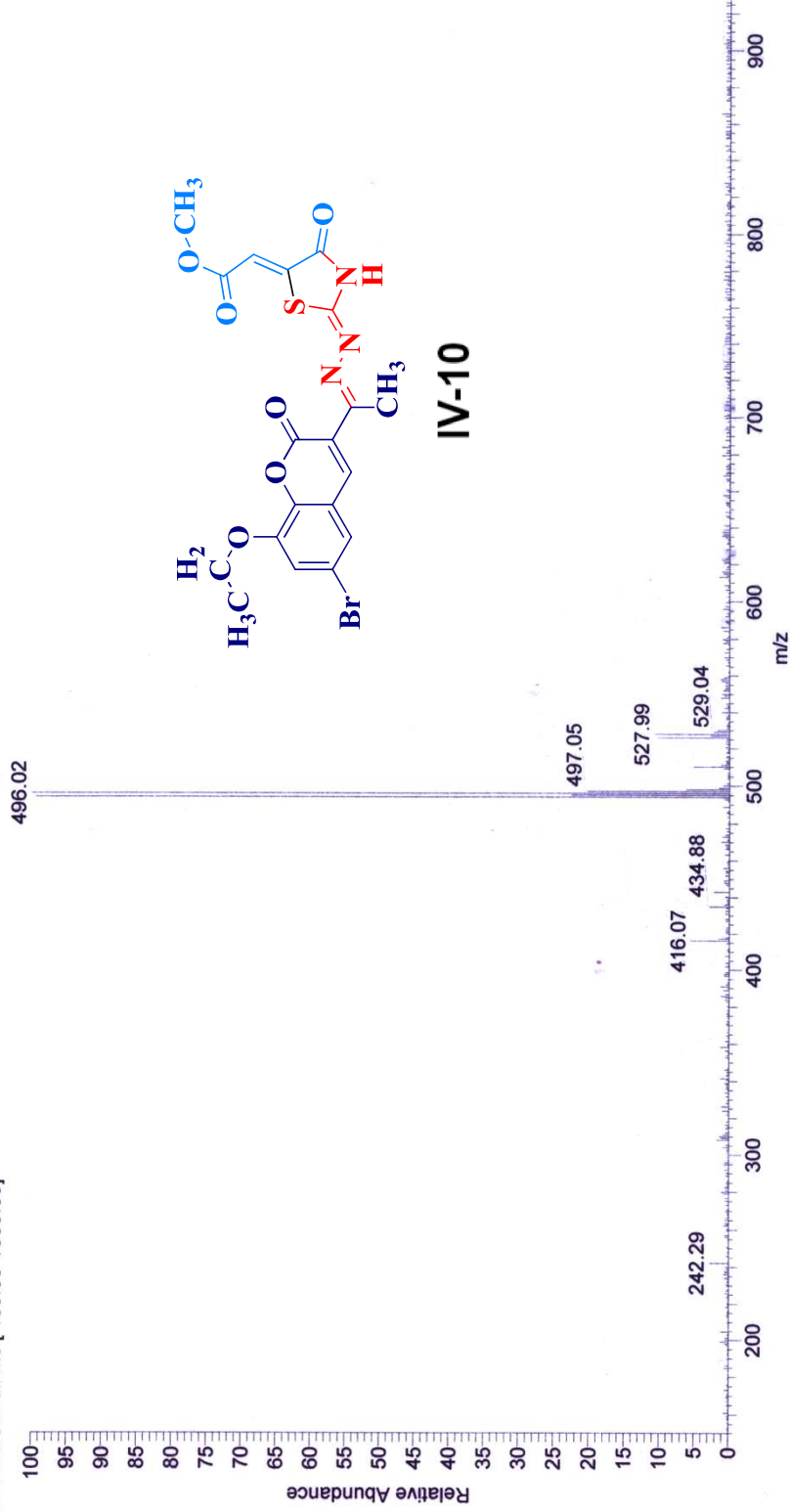


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SAIF, CSIR-CDRI, Lucknow

Data File: 15I15APR40
Original Data Path: 15I15APR40.RAW
Current Data Path: C:\Xcalibur\data\APR2015\15APR2015\1
Sample ID: RM-BROETDM
Acquisition Date: 04/15/15 13:31:49

15I15APR40 #19-46 RT: 0.30-0.71 AV: 28 SB: 2 0.00, 0.00 NL: 6.12E6
T: + c ESI Full ms [150.00-1000.00]



IV-10

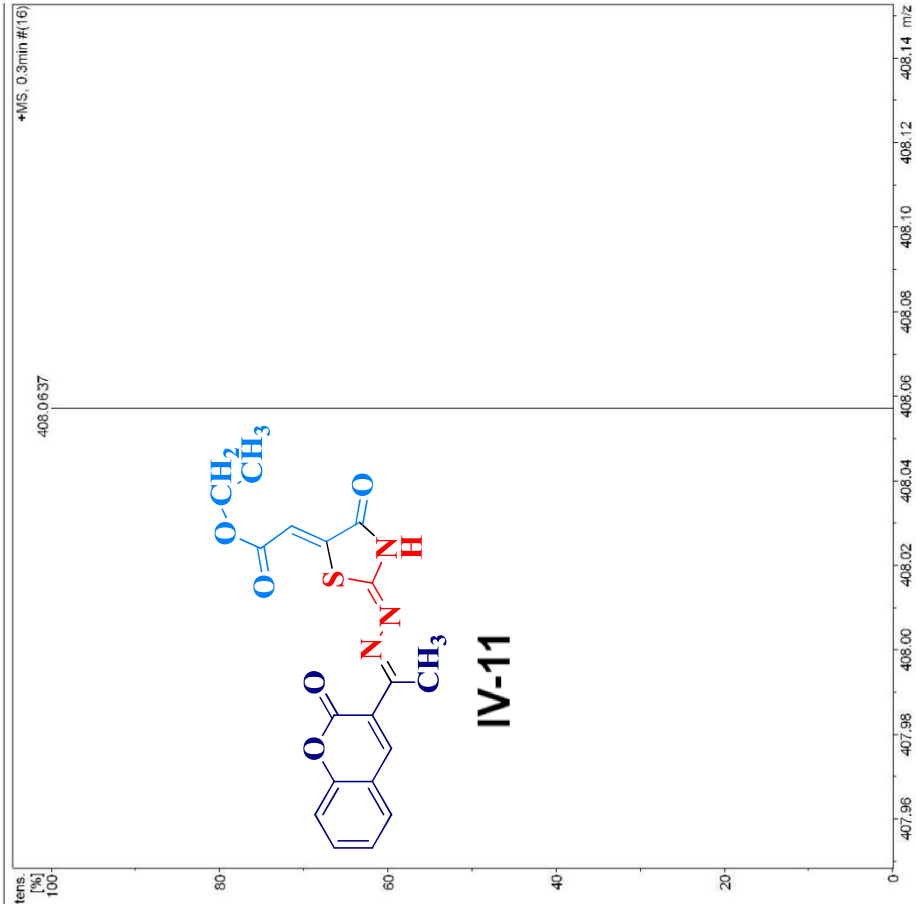
Display Report

Analysis Info

Analysis Name D:\Data\prof.v.k gupta\NA-DEAD.d Acquisition Date 6/22/2014 3:58:03 PM
Method tune_low.m Operator IIT ROORKEE
Sample Name NA-DEAD Instrument micrOTOF-Q II 10328
Comment

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	150.0 Vpp	Set Divert Valve	Source



021405A7434

Medicinal Chemistry Laboratory-Analytical Research

Sample ID:GVK-DU-1928-E11884-138-2

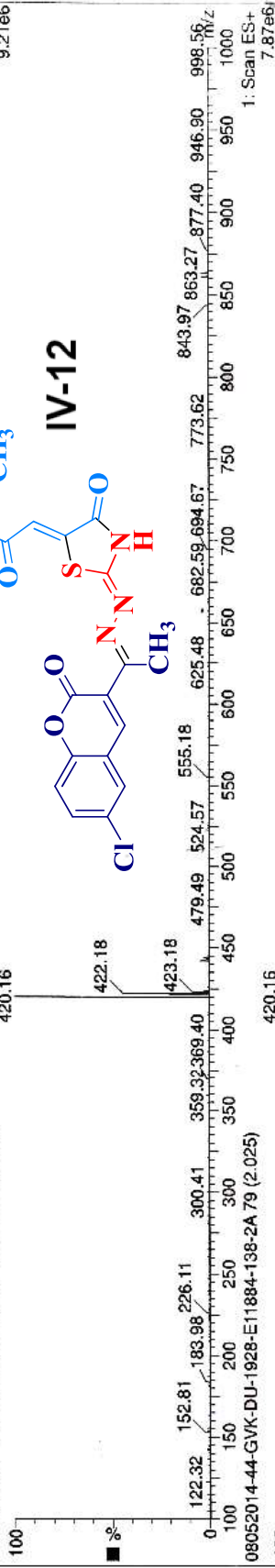
Acq.Method:RND-FA-4.5 MIN

08052014-44-GVK-DU-1928-E11884-138-2A 82 (2.103)

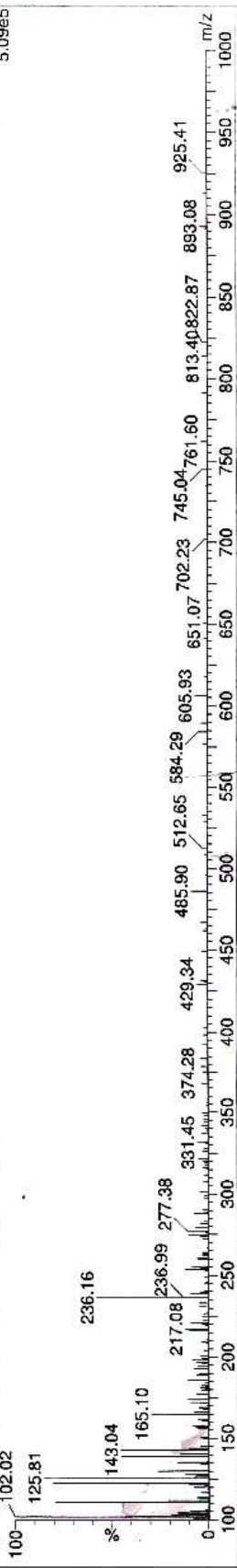
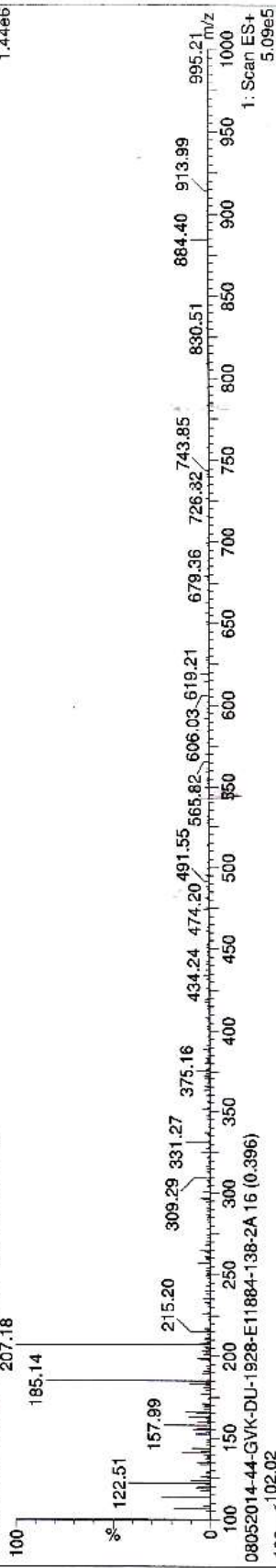
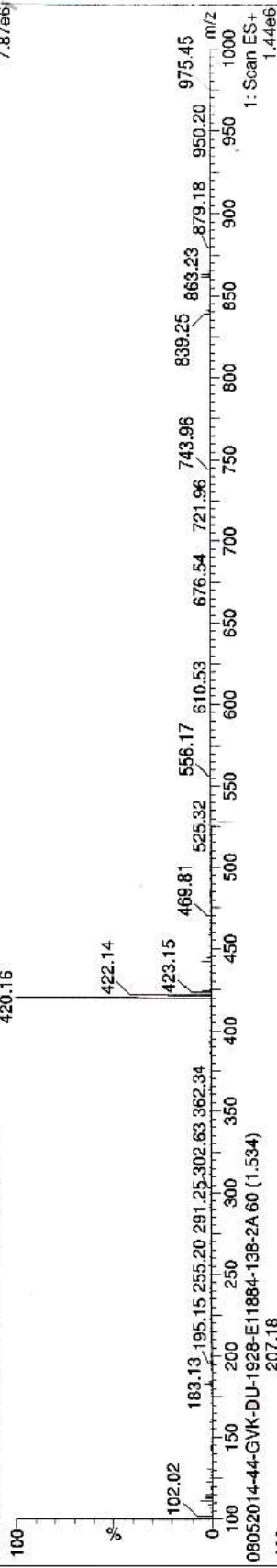
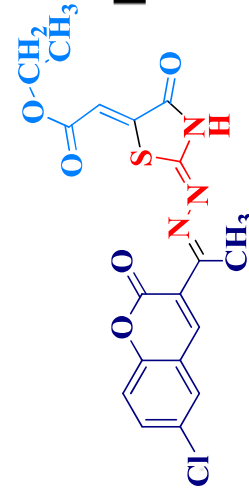
Date of Analysis: 08-May-2014 16:15:45

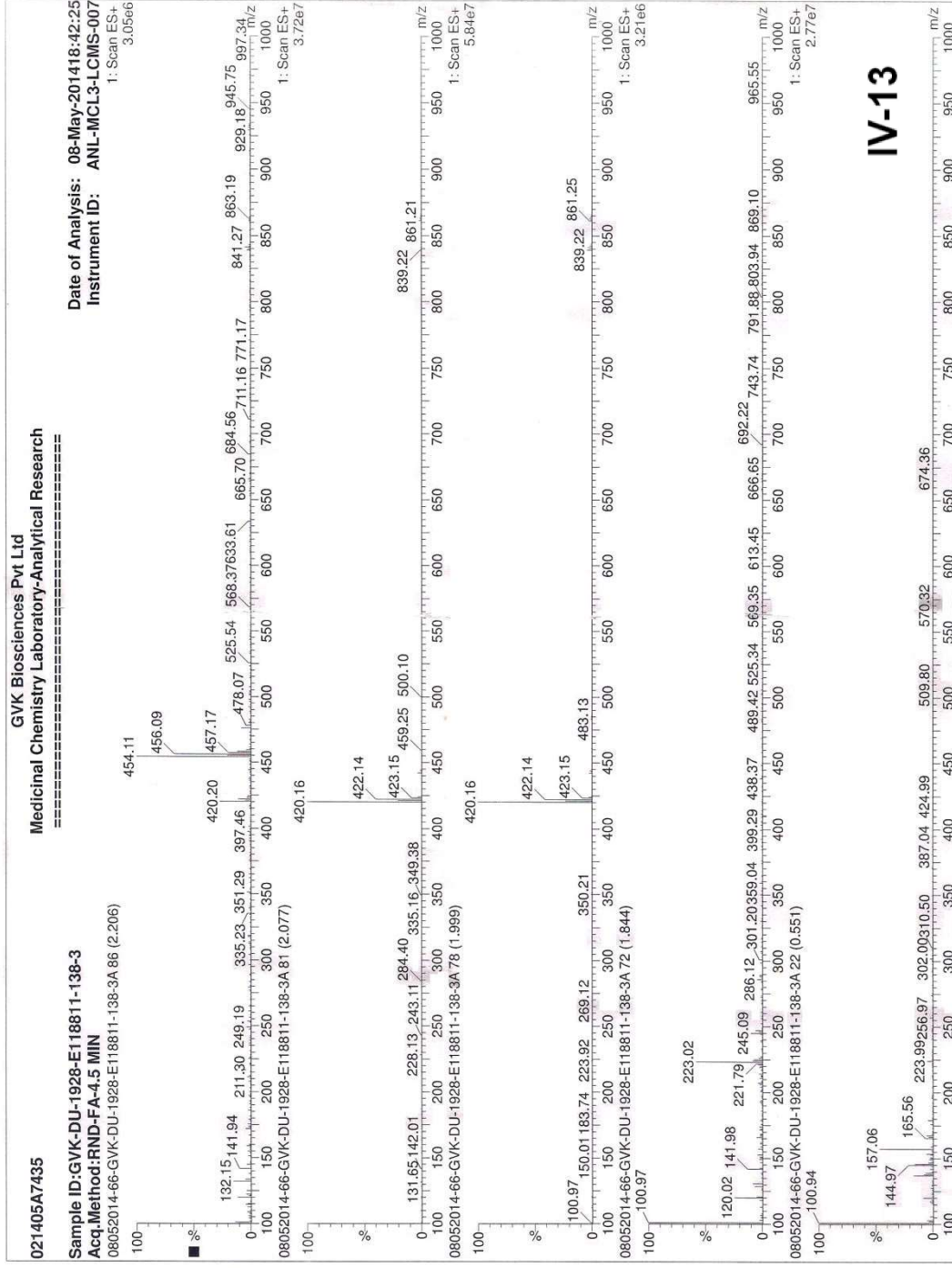
Instrument ID: ANL-MCL3-LCMS-007

1: Scan ES+
9.21e6



IV-12





IV-13

GVK Biosciences Pvt Ltd
Medicinal Chemistry Laboratory-Analytical Research

021405A7433

Sample ID: GVK-DU-1928-E118811-138

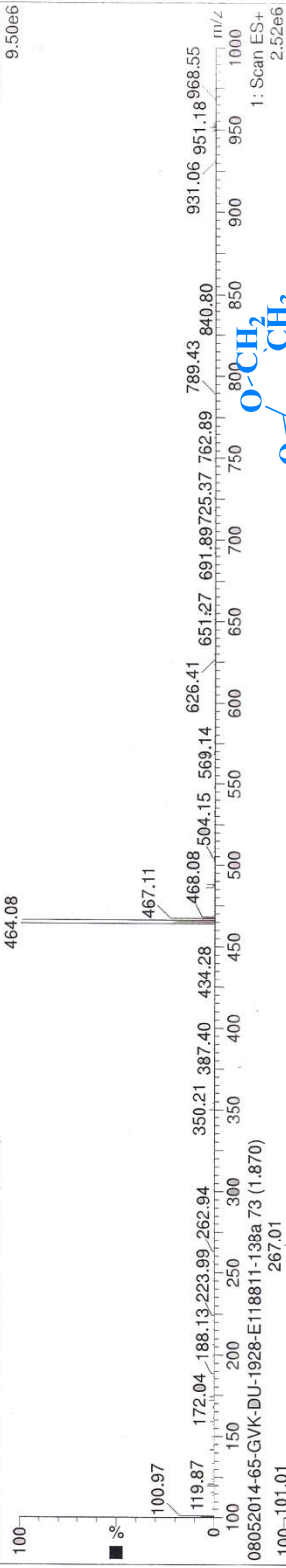
Acq. Method: RND-FA-4.5 MIN

08052014-65-GVK-DU-1928-E118811-138a 82 (2.103)

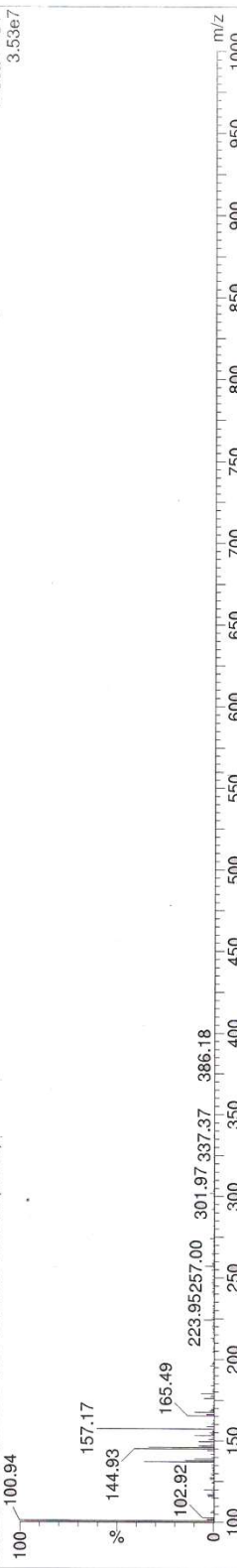
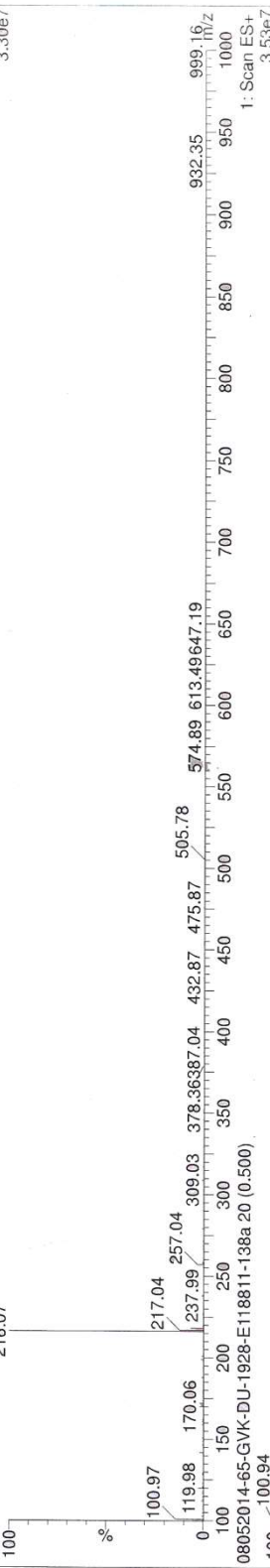
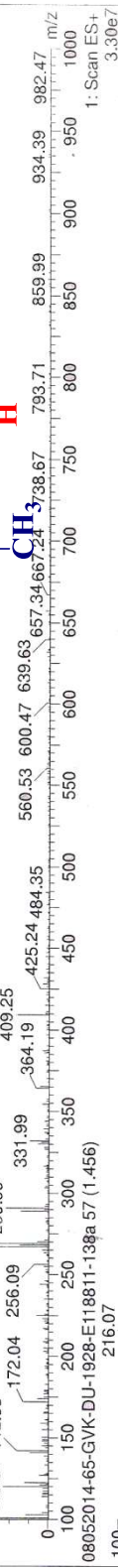
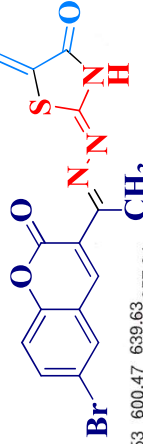
Date of Analysis: 08-May-2014 18:36:30

Instrument ID: ANL-MCL3-LCMS-007

1: Scan ES+
9.50e6



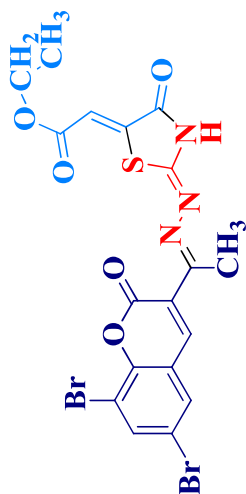
IV-14



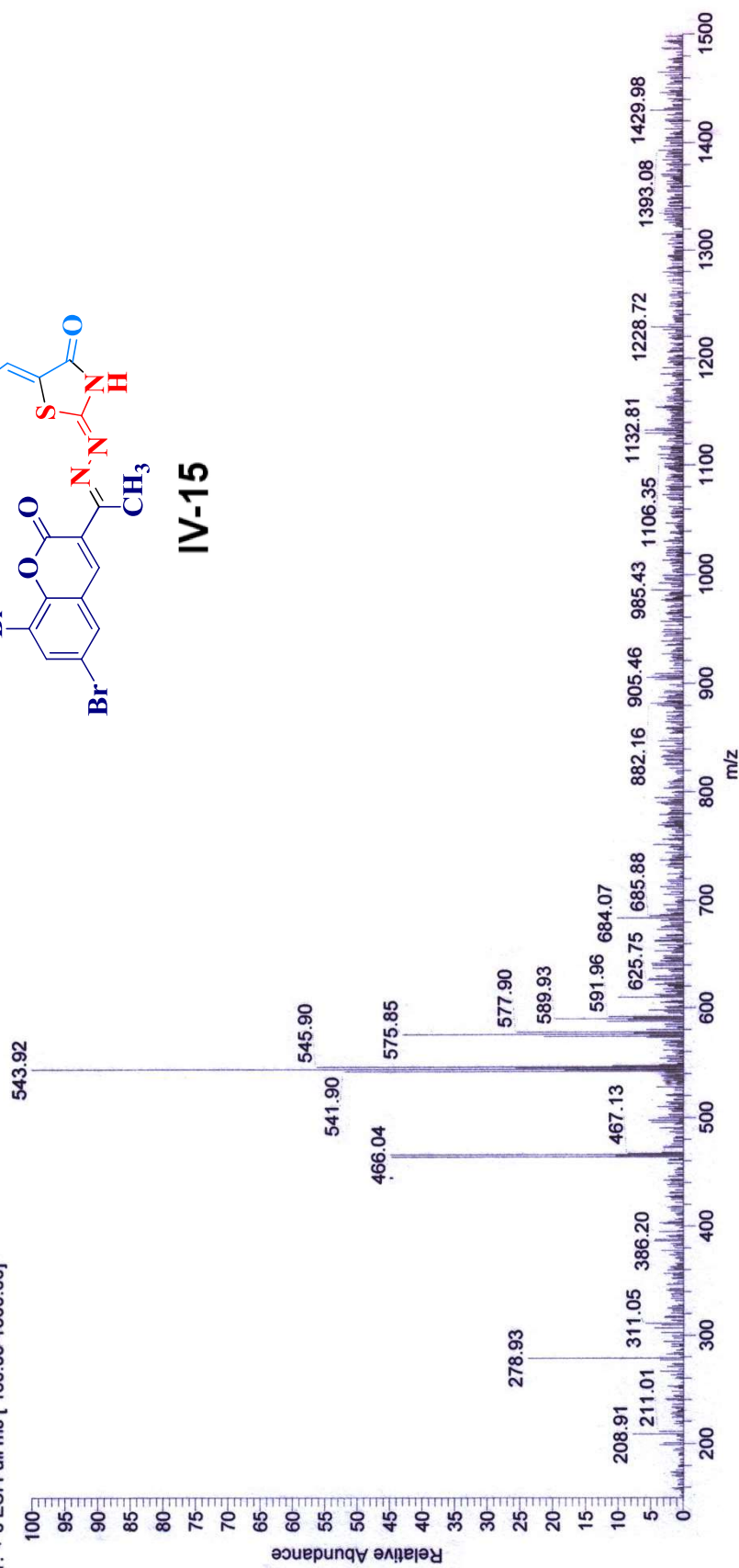
SAIF, CSIR-CDRI, Lucknow

Data File: 15116APR49
Original Data Path: 15116APR49_RAW
Current Data Path: C:\Xcalibur\data\15116APR2015\
Sample ID: RM-DBDE
Acquisition Date: 04/16/15 13:36:13

15116APR49 #15-35 RT: 0.30-0.71 AV: 21 SB: 2 0.00, 0.00 NL: 1.39E6
T: + c ESI Full ms [150.00-1500.00]



IV-15

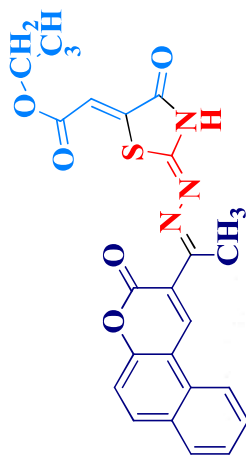
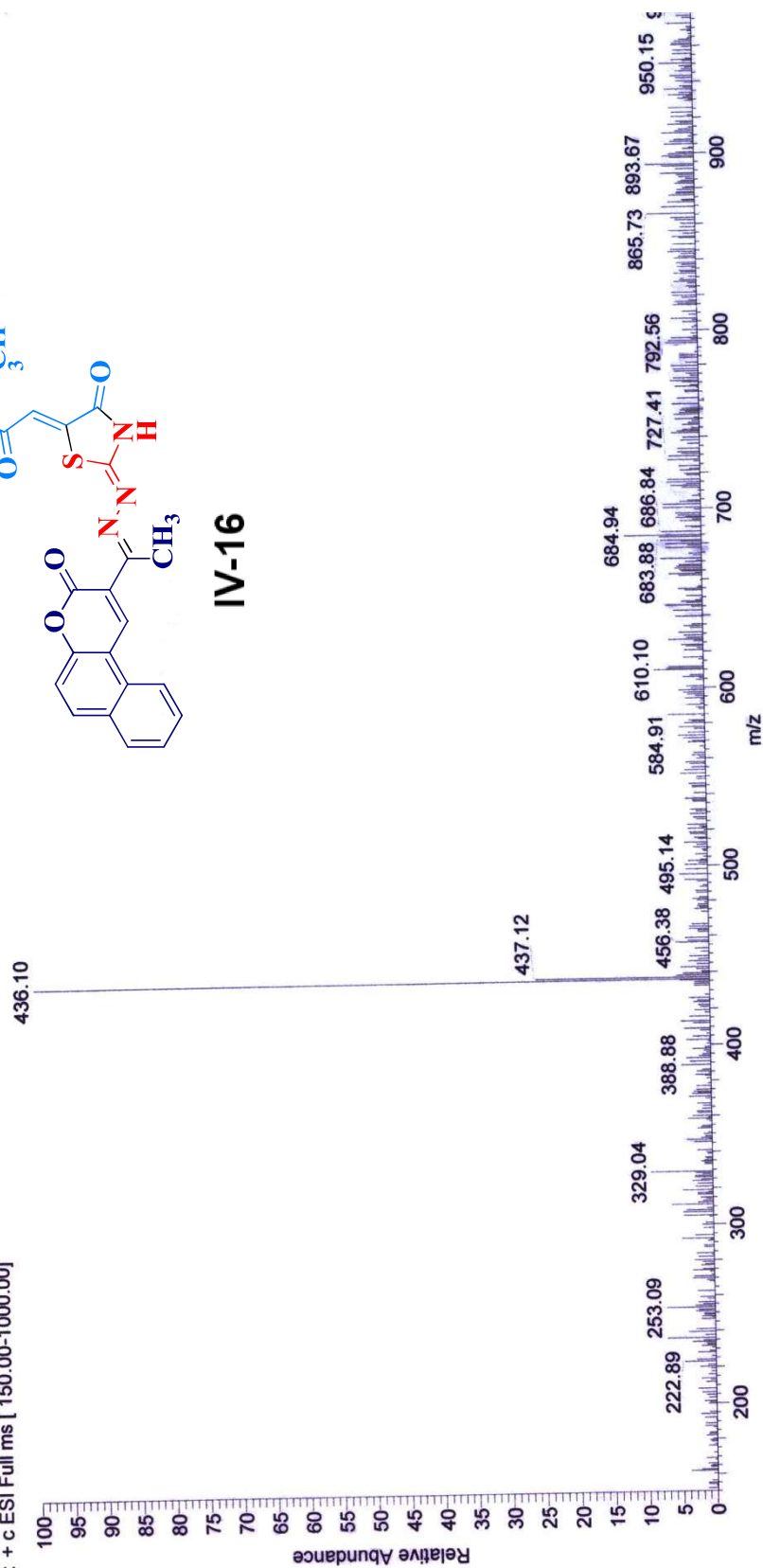


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SAIF, CSIR-CDRI, Lucknow

Data File: 15116APR48
Original Data Path: 15116APR48.RAW
Current Data Path: C:\Xcalibur\data\APR2015\16APR2015\1
Sample ID: RM-NADE
Acquisition Date: 04/16/15 13:34:17

15116APR48 #19-45 RT: 0.30-0.70 AV: 27 SB: 2 0.01 , 0.01 NL: 1.07E6
T: + c ESI Full ms [150.00-1000.00]



IV-16

GVK Biosciences Pvt Ltd
Medicinal Chemistry Laboratory-Analytical Research

021405B4575

Sample ID: DU-1928-E118811-129-NI

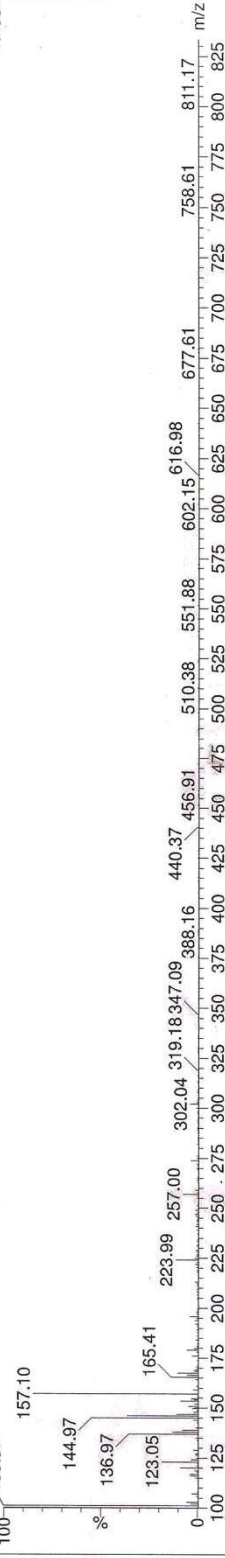
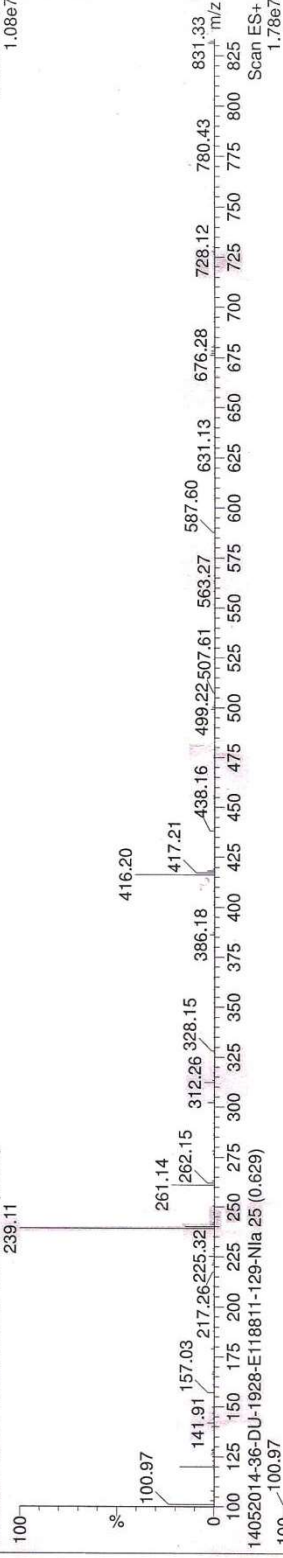
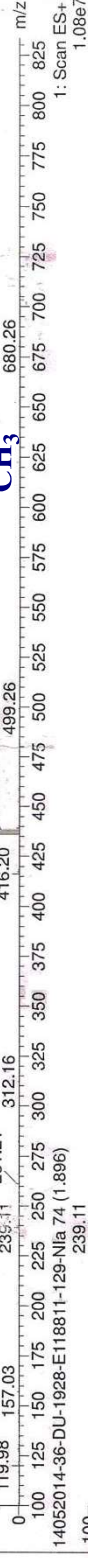
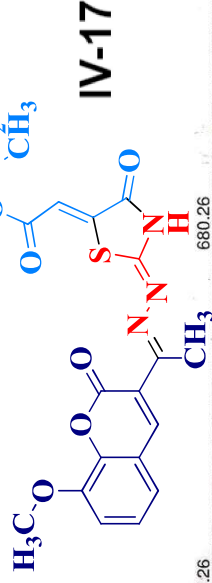
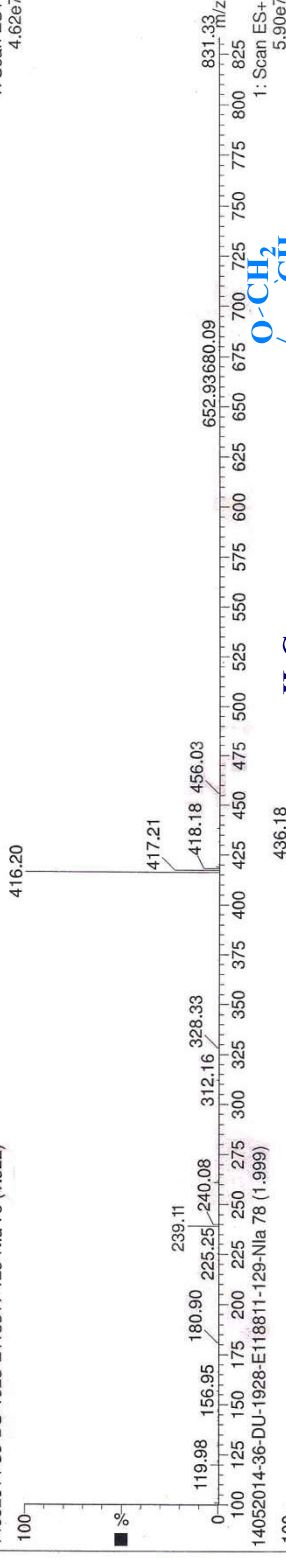
Acq. Method: RND-FA-4.5 MIN

14052014-36-DU-1928-E118811-129-NIa 75 (1.922)

Date of Analysis: 14-May-2014 14:45:11

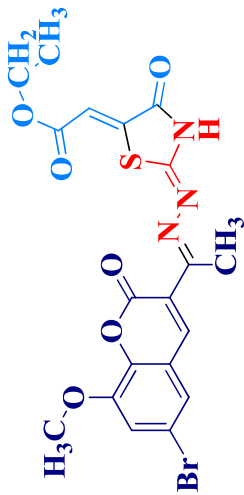
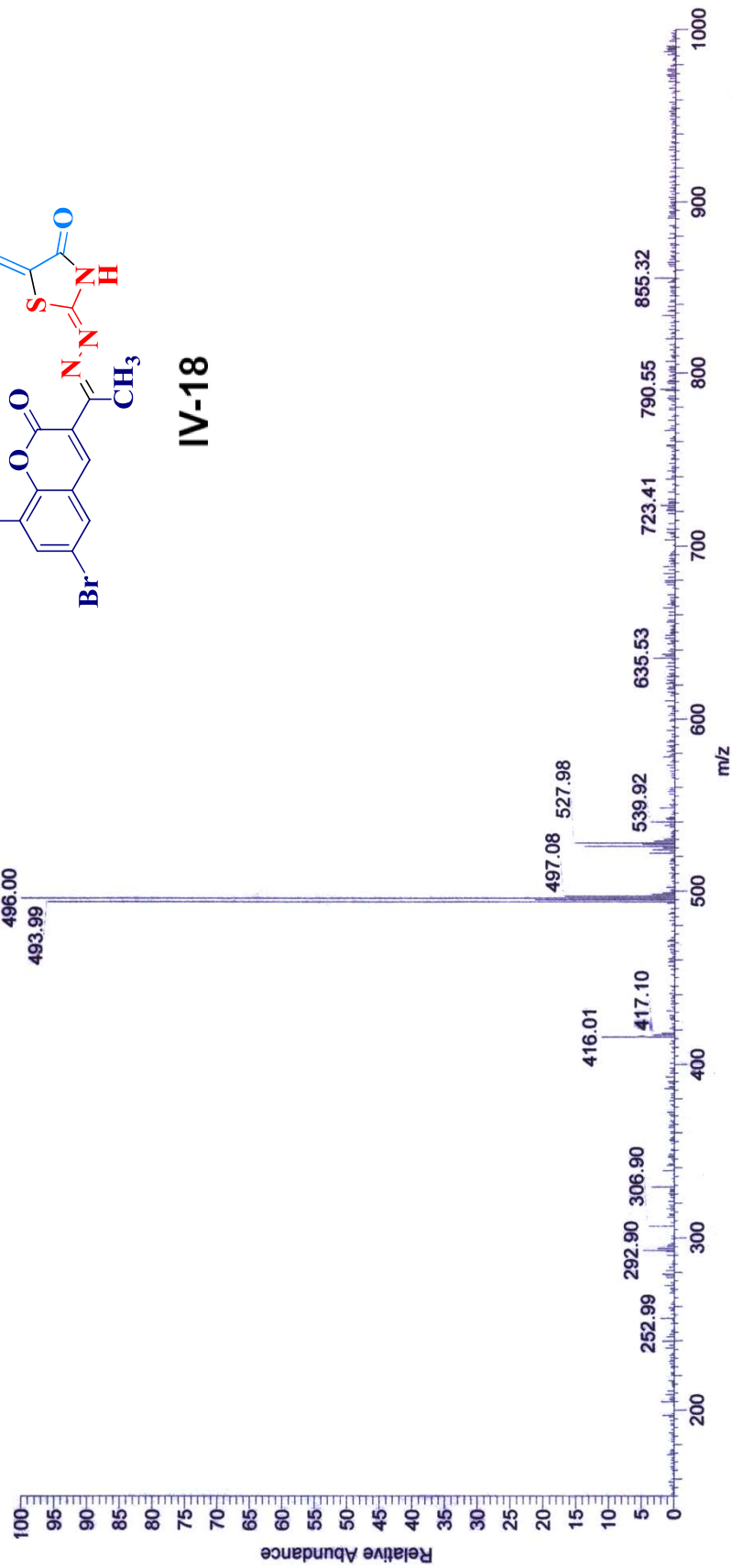
Instrument ID: ANL-MCL3-LCMS-007

1: Scan ES+
4.62e7

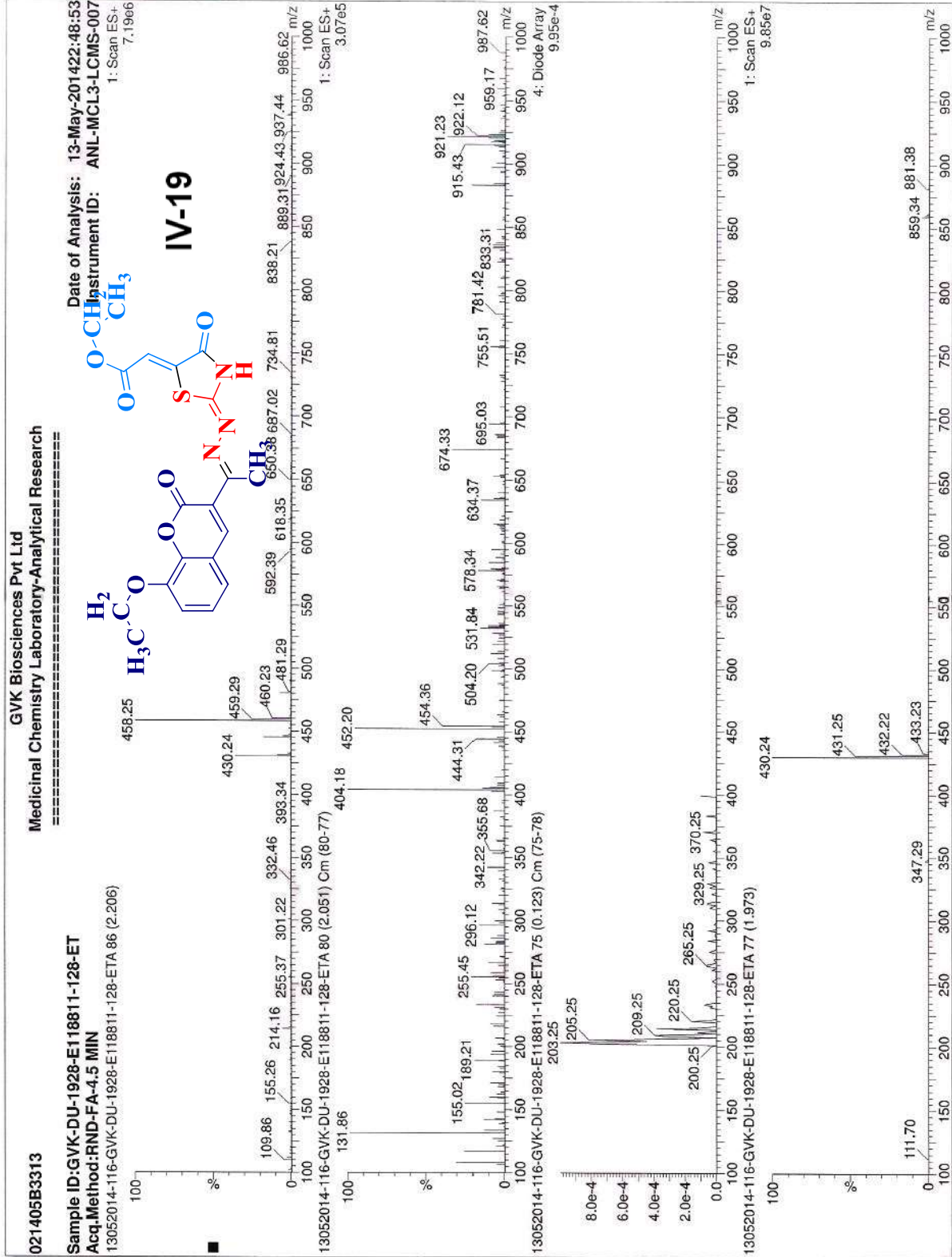


Data File: 15116APR47
Original Data Path: 15116APR47_RAW
Current Data Path: C:\Xcalibur\data\APR2015\1516APR2015\RM-BROVDE
Sample ID:
Acquisition Date: 04/16/15 13:32:17

15I16APR47 #20-46 RT: 0.30-0.70 AV: 27 SB: 2 0.00 , 0.00 NL: 4.03E6
T: + c ESI Full ms [150.00-1000.00]



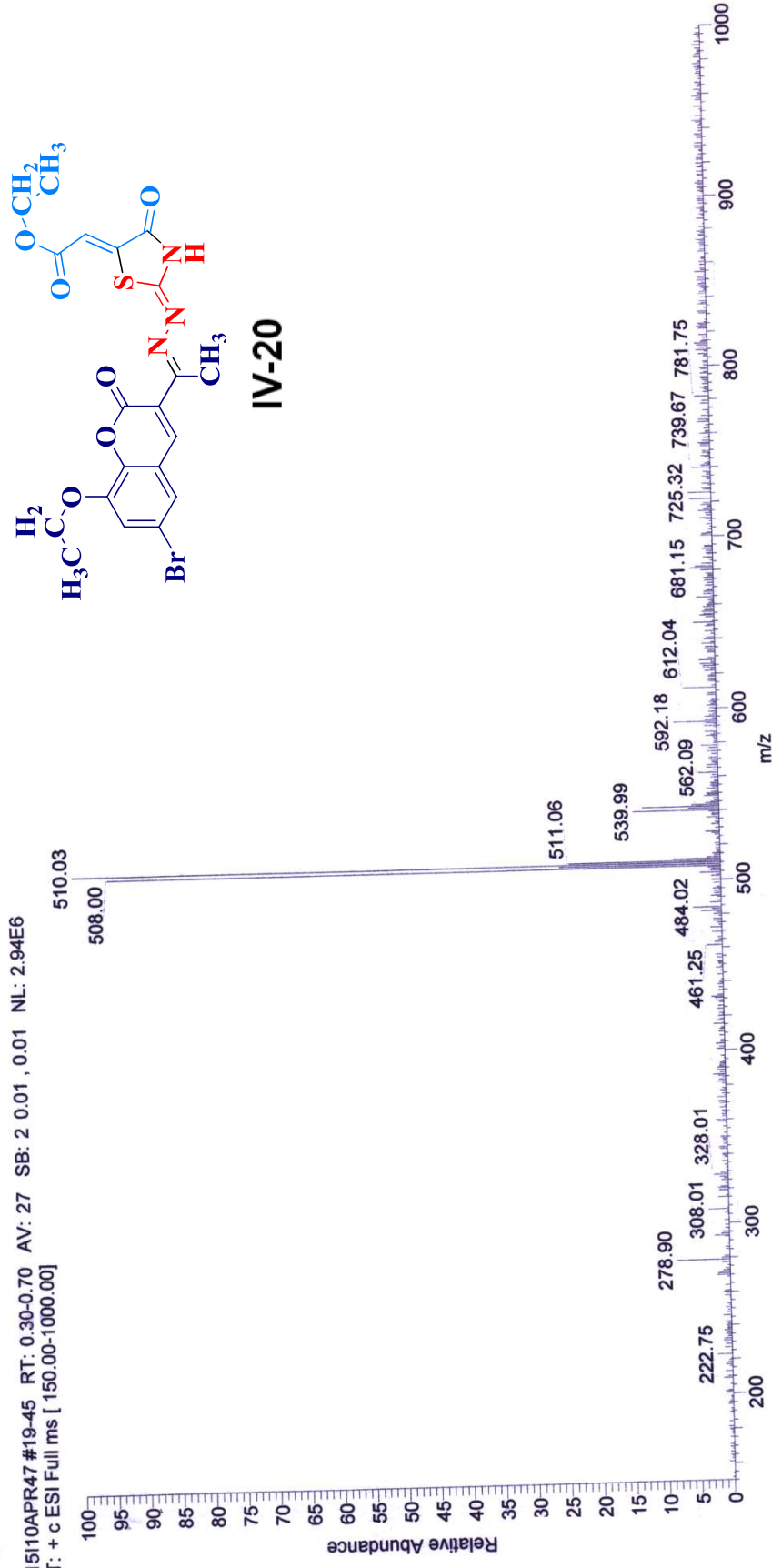
IV-18



SAIF, CSIR-CDRI, Lucknow

Data File: 15110APR47
Original Data Path: 15110APR47.RAW
Current Data Path: C:\XCALIBUR\DATA\15110APR2015\08APR2015\1
Sample ID: RM-OETBR
Acquisition Date: 04/10/15 14:19:46

15110APR47 #19-45 RT: 0.30-0.70 AV: 27 SB: 2 0.01, 0.01 NL: 2.94E6
T: + c ESI Full ms [150.00-1000.00]



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Openlynx Report SAIF, CDRI-LUCKNOW

File:14108DEC385

Vial:1:C,2

Printed: Mon Dec 08 12:58:45 2014

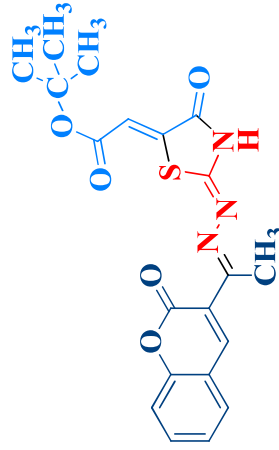
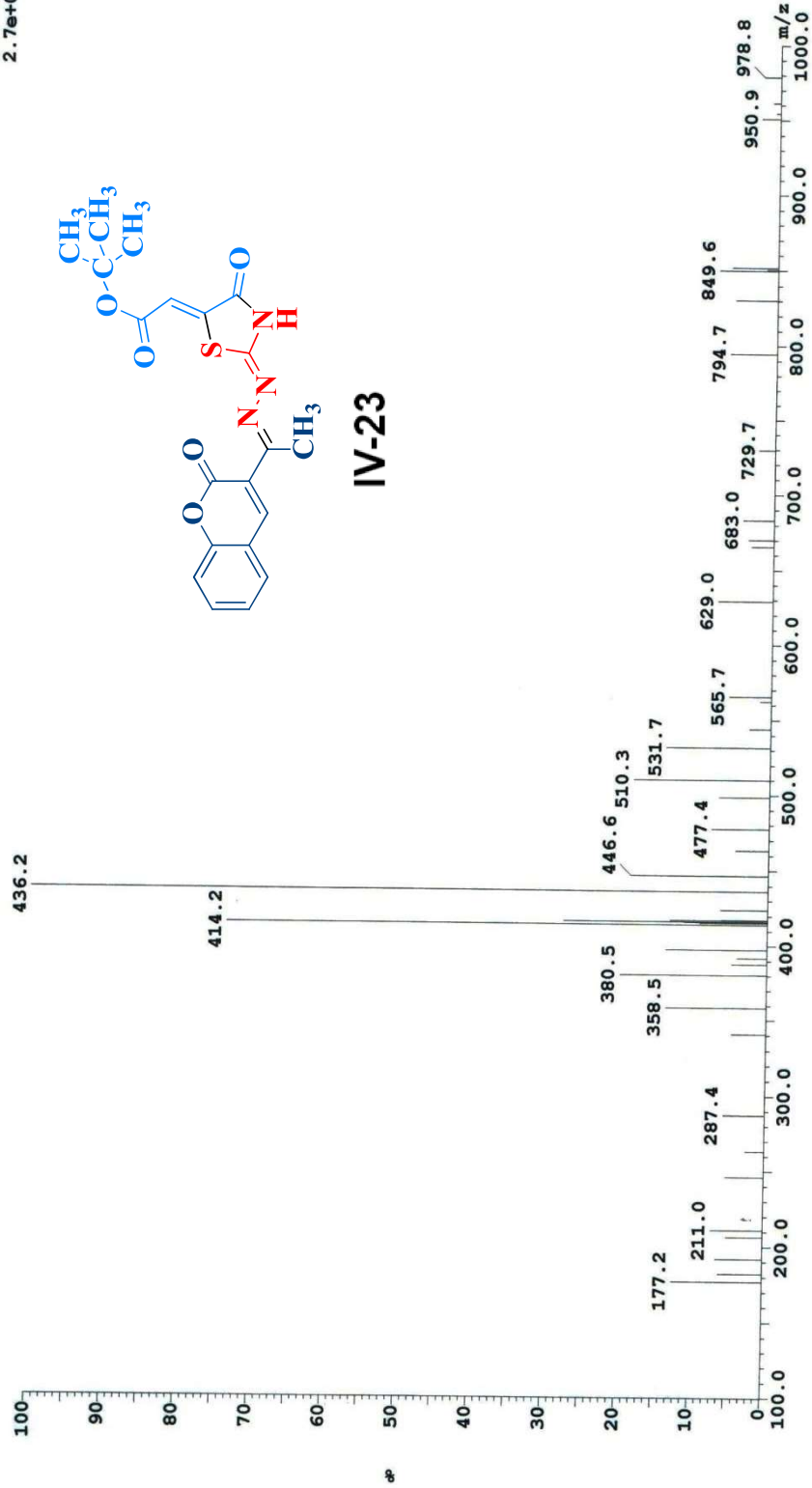
Date:08-Dec-2014
ID:MSR-NTP-9

Time:12:55:29

Page 1

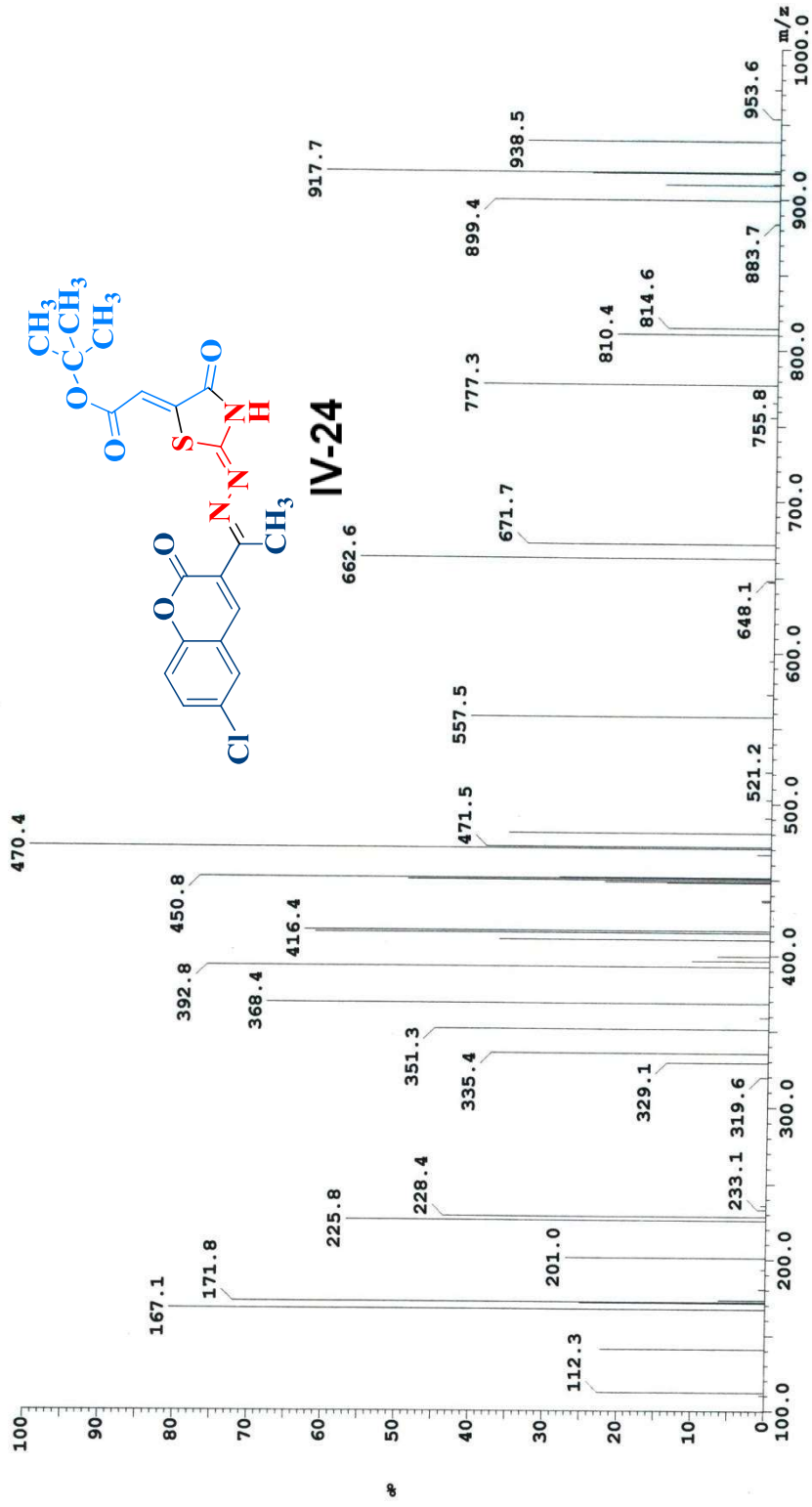
1: (Time: 0.32) Center (Top, 4, Ar) ; Smooth (Mn, 2x0.75) ; Subtract (1, 40.00, 0.010) ; Combine (13:17-(6.7+23:24))

1:MS ES+
2.7e+004



IV-23

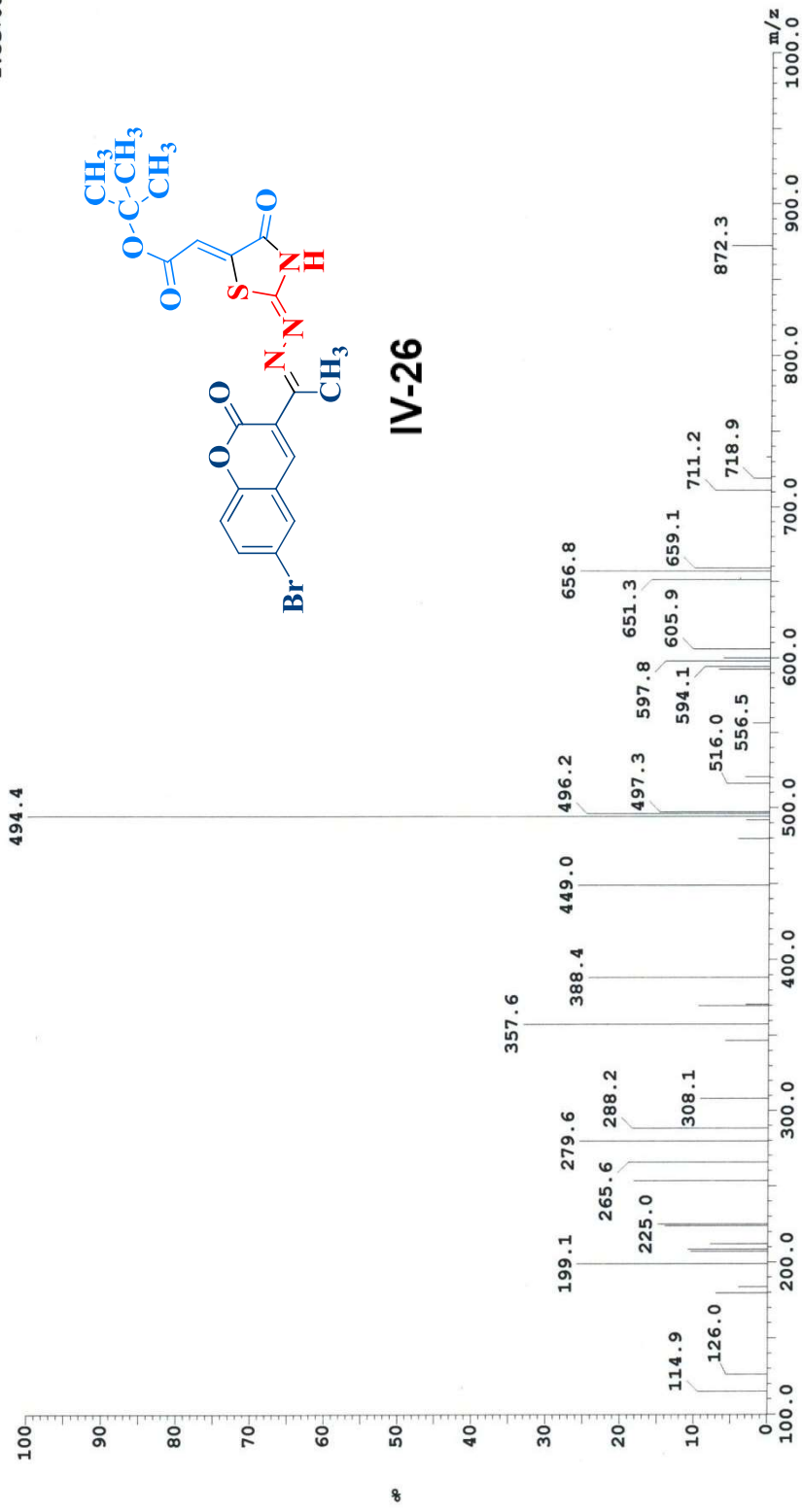
1: (Time: 0.29) Center (Top, 4, Ar); Smooth (Mn, 2x0.75); Subtract (1, 40.00, 0.010); Combine (12:16-(5:6+22:23)) 1:MS ES+



Printed: Mon Dec 08 13:11:55 2014

1: (Time: 0.38) Center (Top, 4, Ar); Smooth (Mn, 2x0.75); Subtract (1, 40.00, 0.010); Combine (16:20- (9:10+26:27))

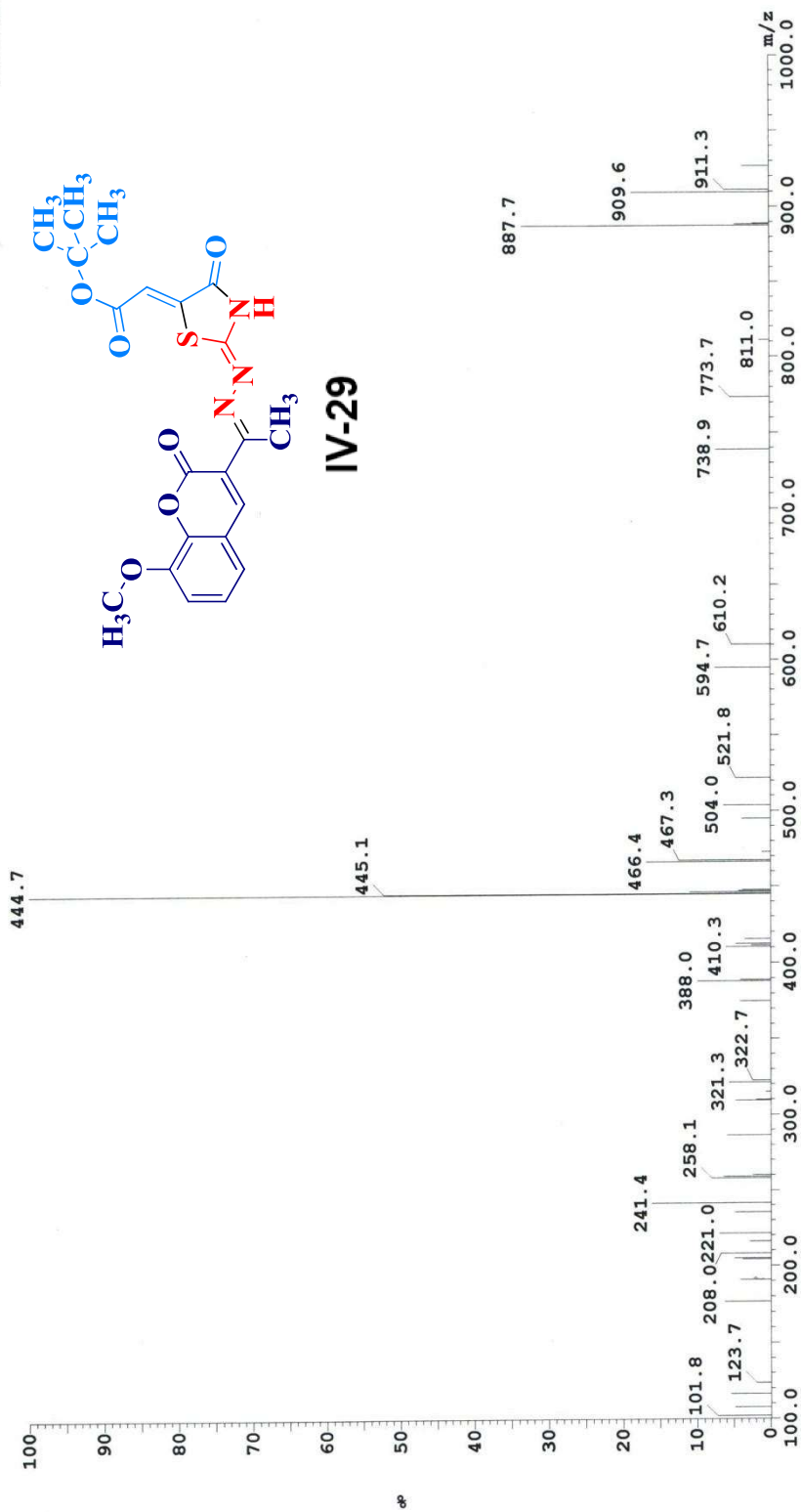
1:MS ES+
1.5e+004



Printed: Mon Dec 08 13:25:06 2014

1: (Time: 0.34) Center (Top, 4, Ar); Smooth (Mn, 2x0.75); Subtract (1, 40.00, 0.010); Combine (14.18-(7:8+24:25))

1:MS ES+
4.9e+004



Display Report

Analysis Info

Analysis Name D:\Data\prof.v.k.gupta\NA-257-1.d Acquisition Date 12/29/2014 6:38:53 PM
 Method tune_low.m Operator IIT ROORKEE
 Sample Name NA-257-1 Instrument microTOF-Q II 10328
 Comment

Acquisition Parameter

Source Type ESI Ion Polarity Positive
 Focus Not active Set Capillary 4500 V
 Scan Begin 50 m/z Set End Plate Offset -500 V
 Scan End 3000 m/z Set Collision Cell RF 150.0 Vpp
 Set Nebulizer 0.4 Bar
 Set Dry Heater 180 °C
 Set Dry Gas 4.0 l/min
 Set Divert Valve Source

