

Central Lancashire Online Knowledge (CLoK)

Title	A spectroscopic liquid biopsy for the earlier detection of multiple cancer
	types
Type	Article
URL	https://clok.uclan.ac.uk/id/eprint/48582/
DOI	https://doi.org/10.1038/s41416-023-02423-7
Date	2023
Citation	Cameron, James, Sala, Alexandra, Antoniou, Georgios, Brennan, Paul, Butler, Holly, Conn, Justin, Connal, Siobhan, Curran, Tom, Hegarty, Mark et al (2023) A spectroscopic liquid biopsy for the earlier detection of multiple cancer types. British Journal of Cancer. ISSN 0007-0920
Creators	Cameron, James, Sala, Alexandra, Antoniou, Georgios, Brennan, Paul, Butler, Holly, Conn, Justin, Connal, Siobhan, Curran, Tom, Hegarty, Mark, McHardy, Rose, Orringer, Daniel, Palmer, David, Smith, Benjamin and Baker, Matthew

It is advisable to refer to the publisher's version if you intend to cite from the work. https://doi.org/10.1038/s41416-023-02423-7

For information about Research at UCLan please go to http://www.uclan.ac.uk/research/

All outputs in CLoK are protected by Intellectual Property Rights law, including Copyright law. Copyright, IPR and Moral Rights for the works on this site are retained by the individual authors and/or other copyright owners. Terms and conditions for use of this material are defined in the http://clok.uclan.ac.uk/policies/

Supplementary Materials for

A spectroscopic liquid biopsy for the earlier detection of multiple cancer types

Authors: James M. Cameron^{1†}, Alexandra Sala^{1,2†}, Georgios Antoniou¹, Paul M. Brennan³, Holly J. Butler¹, Justin J.A. Conn¹, Siobhan Connal^{1,2}, Tom Curran⁴, Mark G. Hegarty¹, Rose G. McHardy^{1,2}, Daniel Orringer⁵, David S. Palmer^{1,2}, Benjamin R. Smith¹, Matthew J. Baker^{1,6*}

† These authors contributed equally.

Affiliations:

¹Dxcover Ltd., Royal College Building, 204 George Street, Glasgow, G1 1XW, UK

²Department of Pure and Applied Chemistry, Thomas Graham Building, 295 Cathedral Street, University of Strathclyde, Glasgow G11XL, UK

³Translational Neurosurgery, Centre for Clinical Brain Sciences, University of Edinburgh, Edinburgh, EH4 2XU, UK

⁴Children's Mercy Research Institute at the Children's Mercy Hospital, Kansas City, KS, USA

⁵Department of Neurosurgery, New York University Grossman School of Medicine, New York, NY 10018, USA

⁶School of Medicine, Faculty of Clinical and Biomedical Sciences, University of Central Lancashire, Preston, PR1 2HE, UK

*Corresponding author. Address: Suite RC534, Royal College Building, 204 George Street, Glasgow, G1 1XW. Tel: (+44)0141 548 4700. Email: matthew.baker@dxcover.com

This PDF file includes:

Methods Supplementary Text Supplementary References Fig. S1 Tables S1 to S10

Supplementary Text

Supplementary Methods

Patient Samples

All samples sourced from patients eligible for inclusion in this study were sourced from biobanks. Cancer samples were gathered from the Wellcome Trust Clinical Research Facility at the Western General Hospital, Edinburgh, the Emergency Medicine Research Group (EMERGE) at the Edinburgh Royal Infirmary, The Beatson West of Scotland Cancer Centre in Glasgow, the University of Swansea, Royal Preston Hospital, and Manchester Cancer Research Centre. The study was designed to include approximately 200 serum samples for each individual class – which is sufficient for reliable precision[1] – and the stage distribution was selected to closely match the incidence and prevalence of tumors in the UK population [2]. All cancer samples were collected from patients with a confirmed cancer diagnosis according to the data collection methods of specified biobanks. Samples were collected before surgical resection or the start of other anti-cancer therapies. The non-cancer group was comprised of both asymptomatic controls and patients with suspicious symptomology. The primary analysis was planned to be agnostic as to whether control subjects were symptomatic or asymptomatic. It is intended to expand this research in future to allow investigation of potential differences in test performance between these groups. Blood samples were obtained with venipuncture using serum collection tubes; S-Monovette Z Gel (Sarstedt, Germany) and Vacutainer SST/SST II (BD, USA), and anonymized. Serum was extracted via centrifugation and stored in a -80°C freezer. Non-identifiable clinical and demographic data were obtained in-line with each biobank's data control procedures.

Patient Sample Analysis

In this study, the serum samples sourced were stored at -80 °C until the date of analysis; samples were allowed to thaw for up to 30 minutes at room temperature (18-25 °C) and inverted three times to ensure mixing and thawing before use. Each patient sample was prepared for analysis by pipetting 3 μL of serum onto each of the three sample wells of the Dxcover® Sample Slide (Dxcover® Ltd., UK). Prepared slides were placed in a drying unit incubator (Thermo ScientificTM HerathermTM, USA) at 35 °C for 1 hour, to control the dehydration process of the serum droplets[3]. Each dried sample slide was then inserted into the Dxcover® Autosampler (Dxcover® Ltd., UK) to be prepared for spectral collection. In this study, a PerkinElmer® Spectrum TwoTM FTIR spectrometer (PerkinElmer® Inc., USA) was used to generate the spectral data (16 co-added scans at 4 cm⁻¹ resolution with 1 cm⁻¹ data spacing). A total of three spectra were collected for each sample well, resulting in nine replicates per patient, then submitted to the diagnostic algorithm to generate the disease prediction. Patient samples were reported as cancer positive or negative according to the diagnostic algorithm results.

Supplementary Results

Organ-specific Classifications

The sensitivity-tuned results are described in **Table S6**. The brain cancer detection rate was 100% (8/8) for grade I, 85% (23/27) for grade II, 86% (12/14) for grade III and 99% (191/192) for grade IV, which had the overwhelming majority for the brain cancer set. The breast cancer group had many more early-stage samples, and the detection rate was 96% (24/25) for stage I, 87% (79/91) for stage II, 89% (67/75) for stage III and 100% (9/9) for stage IV. The classifiers for colorectal, kidney and lung cancer all reported extremely high detection rates, between 98-

100% for stage I and II. Despite being the smallest subset in this study, the ovarian cancer predictions were still highly promising: stage I 97% (30/31); II 86% (12/14); III 92% (47/51); IV 100% (29/29). Likewise, due to difficulty sourcing stage I pancreatic cancer samples there were only 8 included in the dataset, yet the sensitivity-tuned model was still capable of successfully predicting 7 of the stage I tumors (88%). Additionally, the detection rates for the other pancreatic cancer stages were 94% (II, 61/65) 99% (III, 71/72) and 95% (IV, 19/20). Lastly, the prostate cancer results further highlighted the potential of earlier detection: stage I 100% (4/4); II 93% (149/160); III 97% (30/31); IV 75% (3/4).

For the models that were tailored for a greater specificity (**Table S7**), more of the cancer samples were not detected. The brain cancer detection rates were low for grade I-III yet reported 52% (100/192) for grade IV (at 99% specificity). The breast cancer group had detection rates of 32% (8/25) for stage I, 47% (43/91) for stage II, 59% (44/75) for stage III, and 78% (7/9) for stage IV. The colorectal classifier reported the highest stage I detection rate for the specificity-tuned models with 18 out of 36 being predicted correctly (50%). Furthermore, the colorectal stage II, III and IV detection rates were 36% (25/70), 51% (34/67) and 44% (12/27), respectively. Kidney cancer reported the highest detection rate for stage II (66%, 19/29), and the remaining stages were: stage I 46% (40/87); III 38% (13/34); IV 29% (15/51). The detection rates for lung cancer were 35% (11/31) for stage I, 40% (24/60) for stage II, 60% (39/65) for stage III and 44% (20/45) for stage IV. The detection rates were similar for ovarian cancer – stage I 39% (12/31), II 36% (5/14), III 53% (27/51), IV 45% (13/29) – and pancreatic cancer – stage I 38% (3/8), II 37% (24/65), III 44% (32/72), IV 55% (11/20). Finally, the prostate cancer performance was respectable for stage II (43%, 68/160) and III (58%, 18/31), but low for stage I and IV which is likely to be attributed to the very small number of samples in those groups (n = 4).

Supplementary Discussion

Combination Scenario

Using the sensitivity-tuned model as a first-line cancer detection test could capture the majority of cancer patients and fast-track patients with a positive result into further testing (e.g., genetic-based analysis) whilst quickly ruling out 60% of the patients without disease. In theory, this could provide a focused, enriched cohort for a second-line test and due to the minute volume used (only 9 μL required), the additional testing could use the same patient blood draw. The impact on patients of a combined testing regime on a single blood draw can be estimated. Based on a 100,000 patient population with a 2% disease prevalence (projected prevalence in the USA in 2040)[4]. 2,000 patients would have cancer and the remaining 98,000 would not. Implementing a high sensitivity first-line cancer detection test (90% sensitivity / 60% specificity) would mean that 41,000 patients will test positive and proceed to a second-line test, such as next generation sequencing (NGS). At this stage, the disease prevalence would be 4.4%, more than twice than the original population. The other 59,000 patients would test negative and not progress to second-line testing, meaning they can be ruled out in a much shorter time (1-2 days compared to 10+ days), with disease prevalence in the negative cohort of 0.3% which is six times less than in the original population.

Supplementary References

- 1. Beleites C, Neugebauer U, Bocklitz T, Krafft C, Popp J. Sample size planning for classification models. Analytica Chimica Acta. 2013;760:25–33.
- 2. Cancer Research UK. Types of Cancer [Internet]. Accessed August 2022. Available from: http://www.cancerresearchuk.org/about-cancer/what-is-cancer/how-cancer-starts/types-of-cancer
- 3. Cameron JM, Butler HJ, Palmer DS, Baker MJ. Biofluid spectroscopic disease diagnostics: A review on the processes and spectral impact of drying. Journal of Biophotonics. 2018;11:e201700299.
- 4. GLOBOCAN. Globocan Cancer Tomorrow [Internet]. Accessed November 2021. Available from: https://gco.iarc.fr/tomorrow/en/dataviz/tables?populations=900 840&age start=10

Supplementary Figures

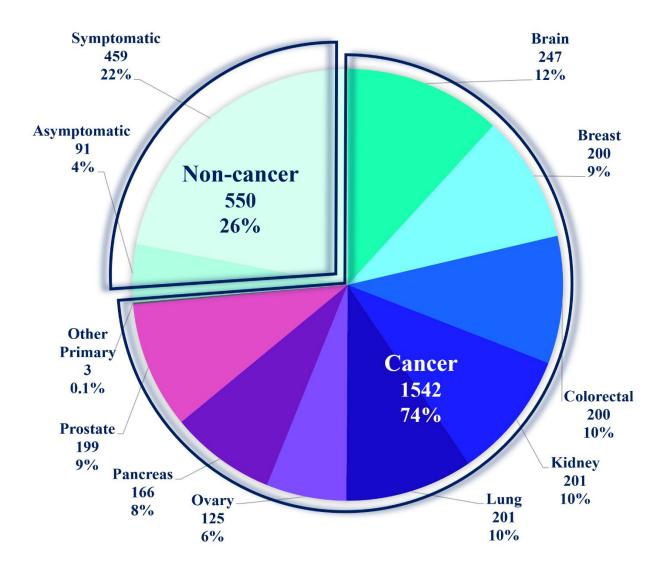


Fig. S1. Schematic breakdown of the full patient cohort. Cancer set is comprised of patients with brain, breast, colorectal, kidney, lung, ovarian, pancreatic, and prostate cancer. Non-cancer group includes asymptomatic participants as well as symptomatic patients with non-malignant disease.

Supplementary Tables

Table S1. Patient demographics for the full patient cohort.

	Cancer (C)	Asymptomatic Non-cancer (NCA)	Non-cancer (NC)	Total [†]
	(n=1542)	(n=91)	(n=550)	(n = 2092)
Age, years				
Mean	63	41	54	61
Min - max	20 - 90	20 - 82	20 - 90	20 - 90
Sex, n (%)				
Female	748 (49)	50 (55)	310 (56)	1058 (51)
Male	794 (51)	41 (45)	240 (44)	1034 (49)
Cancer stage, n (%)				
I	231 (15)	-	-	231 (11)
II	516 (33)	-	-	516 (25)
III	410 (27)	-	-	410 (20)
IV	377 (24)	-	-	377 (18)
Not staged*	6(1)	91 (100)	550 (100)	556 (26)
Unknown**	2 (0)	-	-	2 (0)

[†]Total includes C and NC only; NCA patients are comprised within the NC set. * This group does not require staging (or grading). ** Cancer stage and/or staging information has not been recorded.

Table S2. Patient demographics for the organ specific cancer v non-cancer symptomatic (NCS) classifications.

•144551111•441151151							
	Brain†	Breast	Colorectal	Kidney	Lung	Pancreas	NCS
	(n = 247)	(n = 200)	(n = 200)	(n = 201)	(n = 201)	(n = 166)	(n=459)
Age, years							
Mean	60	62	66	62	66	65	57
Min - max	22 -84	28 - 90	29 - 85	25 - 87	20 - 86	39 - 87	20 - 90
Sex, n (%)							
Female	123 (50)	196 (98)	64 (32)	75 (37)	95 (47)	69 (42)	260 (57)
Male	124 (50)	4(2)	136 (68)	126 (63)	106 (53)	97 (58)	199 (43)
Cancer stage, n (%)							
I	8 (3)	25 (13)	36 (18)	87 (43)	31 (16)	8 (5)	-
II	27 (11)	91 (45)	70 (35)	29 (15)	60 (30)	65 (39)	-
III	14 (6)	75 (37)	67 (33)	34 (17)	65 (32)	72 (43)	-
IV	192 (78)	9 (5)	27 (14)	51 (25)	45 (22)	20 (12)	-
Not staged*	6 (2)	-	-	-	-	-	459 (100)
Unknown**	-	-	-	-	-	1 (1)	-

[†] Brain cancers split by grade, as described in Table S4.

* This group does not require staging (or grading).

** Cancer stage and/or staging information has not been recorded.

Table S3.Patient demographics for the organ specific cancer classifications for ovarian and prostate cancer against the sex-specific non-cancer symptomatic (NCS) groups.

	Ovary	Prostate	NCS Females Only (NCS-F)	NCS Males Only (NCS-M)
	(n=125)	(n = 199)	(n = 260)	(n=199)
Age, years				
Mean	61	62	56	59
Min - max	21 - 88	43 - 87	20 - 90	20 - 90
Sex, n (%)				
Female	125 (100)	-	260 (100)	-
Male	-	199 (100)	-	199 (100)
Cancer stage, n (%)				
I	31 (25)	4 (2)	-	-
II	14 (11)	160 (80)	-	-
III	51 (41)	31 (16)	-	-
IV	29 (23)	4 (2)	-	-
Not staged*	<u>-</u>	-	260 (100)	199 (100)

^{*} This group does not require staging.

Table S4. Summary of brain cancer types split by tumor grade.

WHO Grade	Brain Tumor Type	Number of patients
	Meningioma	6
I	Schwannoma	1
	Subependymoma	1
	Astrocytoma	8
	Ependymoma	1
II	Meningioma	10
	Oligodendroglioma	7
	Pituitary Adenoma	1
	Astrocytoma	7
III	Medulloblastoma	1
	Oligodendroglioma	6
	Glioblastoma	148
IV	Gliosarcoma	2
	Medulloblastoma	2
	Metastatic	40

Table S5.Sensitivity and specificity values for the resampled test sets for each of the organ-specific classifications. The results here are based upon thresholds chosen where either sensitivity or specificity was a minimum value of 90% for the cross-validation. Non-cancer symptomatic (NCS), NCS female-only (NCS-F) and NCS male-only (NCS-M).

Ougan Specific Classifier	Sensitivi	ty-tuned	Specificity-tuned		
Organ Specific Classifier	Sensitivity (%)	Specificity (%)	Sensitivity (%)	Specificity (%)	
Brain v NCS	91	64	74	91	
Breast v NCS	93	31	38	93	
Colorectal v NCS	91	76	77	90	
Kidney v NCS	92	79	71	91	
Lung v NCS	93	78	69	90	
Ovary v NCS-F	92	54	58	90	
Pancreas v NCS	92	52	57	90	
Prostate v NCS-M	92	51	66	93	

Table S6. Detection rates from the organ-specific classifications for the sensitivity-tuned models, broken down by stage, based upon the models with a lower limit of 45% specificity for the cross validation.

Cancer Type	Stage	Actual	Identified Correctly	Detection Rate (%)
	I	8	8	100
	II	27	23	85
Brain [†]	III	14	12	86
	IV	192	191	99
	Not Graded *	6	5	83
	I	25	24	96
D 4	II	91	79	87
Breast	III	75	67	89
	IV	9	9	100
	I	36	36	100
0.1	II	70	70	100
Colorectal	III	67	65	97
	IV	27	24	89
	I	87	86	99
17' 1	II	29	29	100
Kidney	III	34	34	100
	IV	51	50	98
	I	31	31	100
T	II	60	59	98
Lung	III	65	65	100
	IV	45	45	100
	I	31	30	97
	II	14	12	86
Ovary	III	51	47	92
	IV	29	29	100
	I	8	7	88
	II	65	61	94
Pancreas	III	72	71	99
	IV	20	19	95
	Unknown**	1	1	100
	I	4	4	100
D 4.4	II	160	149	93
Prostate	III	31	30	97
	IV	4	3	75

[†] Brain cancers split by grade, as described in Table S4. * This group of cancers do not require grading.

^{**} Cancer stage and/or staging information has not been recorded.

Table S7.Detection rates from the organ-specific classifications, for the specificity-tuned models split by stage. The results here are based upon a lower limit of 45% sensitivity for the cross-validation.

Cancer Type	Stage	Actual	Identified Correctly	Detection Rate (%)
	I	8	2	25
	II	27	4	15
Brain [†]	III	14	3	21
	IV	192	100	52
	Not Graded *	6	2	33
	I	25	8	32
Breast	II	91	43	47
Dreast	III	75	44	59
	IV	9	7	78
	I	36	18	50
0.1	II	70	25	36
Colorectal	III	67	34	51
	IV	27	12	44
	I	87	40	46
IZ' 1	II	29	19	66
Kidney	III	34	13	38
	IV	51	15	29
	I	31	11	35
T	II	60	24	40
Lung	III	65	39	60
	IV	45	20	44
	I	31	12	39
	II	14	5	36
Ovary	III	51	27	53
	IV	29	13	45
	I	8	3	38
	II	65	24	37
Pancreas	III	72	32	44
	IV	20	11	55
	Unknown**	1	1	100
	I	4	1	25
D	II	160	68	43
Prostate	III	31	18	58
	IV	4	1	25

[†] Brain cancers split by grade, as described in Table S4.

^{*} This group of cancers do not require grading.

^{**} Cancer stage and/or staging information has not been recorded.

Table S8. Positive predictive values (PPV) split by cancer type, for sensitivity-tuned and specificity-tuned models. PPVs have been calculated by assuming 2% disease prevalence for a screening population and 7% prevalence for a symptomatic setting.

	Screening scenario	(est. 2% prevalence)	Symptomatic scenario	io (est. 7% prevalence)
Cancer Type	PPV (Sensitivity-tuned)	PPV (Specificity-tuned)	PPV (Sensitivity-tuned)	PPV (Specificity-tuned)
Brain	3.3%	46.5%	11.2%	75.1%
Breast	3.1%	7.6%	10.5%	23.2%
Colorectal	3.4%	23.7%	11.6%	53.3%
Kidney	3.3%	19.6%	11.2%	47.0%
Lung	3.3%	15.6%	11.0%	40.2%
Ovary	3.1%	12.5%	10.7%	34.4%
Pancreas	3.2%	15.8%	10.9%	40.8%
Prostate	3.3%	19.2%	11.2%	46.5%

Table S9. 95% confidence intervals (CI) for a one-sample Student's t-test, carried out for each selected threshold on the presented receiver operating characteristic curves for every classification.

Model	Tuning	CI Sens (%)	CI Spec (%)
G NG	98% Sens	(97.6, 98.0)	(56.9, 61.2)
C v NCA	98% Spec	(55.7, 57.8)	(98.1, 99.7)
G 376	90% Sens	(89.8, 90.6)	(58.8, 60.5)
C v NC	95% Spec	(39.3, 40.9)	(94.3, 95.3)
	45% Sens	(44.6, 47.6)	(98.8, 99.3)
D ' MGG	45% Spec	(94.7, 96.3)	(40.8, 43.9)
Brain v NCS	90% Sens	(89.0, 90.8)	(61.0, 63.8)
	90% Spec	(72.4, 75.2)	(89.6, 90.7)
	45% Sens	(44.9, 48.7)	(85.9, 87.8)
D. A. NGC	45% Spec	(86.8, 88.9)	(42.5, 44.5)
Breast v NCS	90% Sens	(91.7, 93.6)	(30.5, 32.6)
	90% Spec	(34.8, 37.5)	(90.8, 92.4)
	45% Sens	(43.3, 46.5)	(96.3, 97.2)
C.1. A.1. NGC	45% Spec	(96.6, 97.8)	(42.4, 45.5)
Colorectal v NCS	90% Sens	(89.5, 91.4)	(73.7, 75.8)
	90% Spec	(73.4, 76.7)	(88.9, 90.5)
	45% Sens	(42.9, 46.0)	(95.0, 96.1)
V: 4 NCC	45% Spec	(98.6, 99.3)	(40.3, 42.5)
Kidney v NCS	90% Sens	(90.4, 92.3)	(77.7, 79.4)
	90% Spec	(68.7, 71.4)	(89.5, 90.9)
	45% Sens	(45.1, 48.6)	(94.1, 95.3)
I NCC	45% Spec	(99.3, 99.8)	(38.2, 40.9)
Lung v NCS	90% Sens	(90.7, 92.8)	(76.7, 78.6)
	90% Spec	(65.7, 69.2)	(88.6, 90.1)
	45% Sens	(44.3, 48.4)	(94.7, 96.1)
Ovary v NCS-F	45% Spec	(93.4, 95.6)	(41.0, 43.9)
Ovary v NCS-r	90% Sens	(90.8, 93.2)	(52.0, 55.3)
	90% Spec	(58.6, 62.3)	(88.5, 90.4)
	45% Sens	(41.6, 45.7)	(92.2, 93.7)
Pancreas v NCS	45% Spec	(94.3, 96.0)	(40.9, 43.6)
Fancicas V INCS	90% Sens	(91.0, 93.0)	(50.6, 53.3)
	90% Spec	(51.9, 55.9)	(89.0, 90.7)
	45% Sens	(43.5, 46.7)	(95.8, 97.2)
Prostate v NCC M	45% Spec	(92.3, 94.2)	(42.1, 45.6)
Prostate v NCS-M	90% Sens	(90.3, 92.5)	(49.1, 53.0)
	90% Spec	(63.2, 66.6)	(89.8, 91.8)

Table S10.Detection rates for the cancer (C) v non-cancer (NC) sensitivity and specificity-tuned models split by age and sex.

		Correctly identif				ntified <i>n</i> (%)	
		No. of	Patients	Sensitivity-tuned Model (90% Sensitivity)		Specificity-tuned Model (95% Specificity)	
		С	NC	C	NC	C	NC
Corr	Male	794	240	732 (92)	153 (64)	327 (41)	229 (95)
Sex	Female	748	310	660 (88)	184 (59)	291 (40)	296 (95)
	20	15	52	13 (87)	42 (81)	6 (40)	52 (100)
	30	37	65	28 (76)	48 (74)	14 (38)	65 (100)
	40	140	66	118 (84)	44 (67)	47 (34)	62 (94)
A (50	331	84	294 (89)	49 (58)	118 (36)	81 (96)
Age (deciles)	60	539	105	490 (91)	52 (50)	219 (41)	98 (93)
	70	373	116	347 (93)	63 (54)	170 (46)	107 (92)
	80	104	54	100 (96)	34 (63)	43 (41)	53 (98)
	90	3	4	2 (67)	2 (50)	1 (33)	3 (75)

Table S11.The top 5 wavenumber regions which were found to be the most discriminatory for each of the binary classifications, with their corresponding tentative biological assignments and vibrational modes.

	Classification	Approximate wavenumber (cm ⁻¹)	Importance	Biological assignments	Vibrational Modes
		1530	100	Amide II of Proteins	N-H bending, C-N stretching
		1260	58	Amide III of Proteins, Phosphodiesters	N-H in-plane bend, C-N stretching, asymmetric PO_2^- stretching
	C v NC	1025	58	Glycogen, Carbohydrates	C-O and C-C stretching, C-OH deformation
		1061	57	Nucleic acids, Phosphodiesters	Symmetric PO ₂ ⁻ stretching, C-O stretching
		3345	56	Amide A of Proteins	OH, C-H, N-H stretching
		1523	100	Amide II of Proteins	N-H bending, C-N stretching
		1607	75	Amide I of Proteins	C=O and C-N stretching, N-H bending
	Brain v NCS	3278	58	Proteins (Amide A), Nucleic acids	Symmetric O-H stretching, N-H stretching
		2861	52	Lipids	C-H, CH ₂ stretching
		1256	49	Amide III of Proteins, Phosphodiesters	N-H in-plane bend, C-N stretching, asymmetric PO_2^- stretching
		2872	100	Glycogen, Carbohydrates	C-O and C-C stretching, C-OH deformation
		1261	80	Amide III of Proteins, Phosphodiesters	N-H in-plane bend, C-N stretching, asymmetric PO ₂ ⁻ stretching
	Breast v NCS	1549	76	Amide II of Proteins	N-H bending, C-N stretching
		3351	74	Amide A of Proteins	OH, C-H, N-H stretching
		1025	65	Glycogen, Carbohydrates	C-O and C-C stretching, C-OH deformation
		1530	100	Amide II of Proteins	N-H bending, C-N stretching
		3351	69	Amide A of Proteins	O-H, C-H, N-H stretching
	Colorectal v NCS	3234	65	Proteins (Amide A), Nucleic acids	Symmetric O-H stretching, N-H stretching
		1246	59	Amide III of Proteins, Phosphodiesters	N-H in-plane bend, C-N stretching, asymmetric PO ₂ stretching
		1666	54	Amide I of Proteins	C=O and C-N stretching, N-H bending

Kidney v NCS	1258	100	Amide III of Proteins, Phosphodiesters	N-H in-plane bend, C-N stretching, asymmetric PO_2^- stretching
	3345	86	Amide A of Proteins	O-H, C-H, N-H stretching
	1531	83	Amide II of Proteins	N-H bending, C-N stretching
	1348	65	Lipids, Proteins	C-O stretching, C-H and N-H deformation
	1664	65	Amide I of Proteins	C=O and C-N stretching, N-H bending
Lung v NCS	1167	100	Nucleic Acids	Asymmetric PO ₂ stretching
	1074	98	Nucleic acids, Phosphodiesters	Symmetric PO ₂ ⁻ stretching, C-O stretching
	1532	74	Amide II of Proteins	N-H bending, C-N stretching
	2750	66	Lipids	C-H, CH ₂ stretching
	1124	63	Carbohydrates	C-O and C-C stretching
Ovary v NCS-F	1529	100	Amide II of Proteins	N-H bending, C-N stretching
	3327	82	Amide A of Proteins	O-H, C-H, N-H stretching
	3244	79	Proteins (Amide A), Nucleic acids	Symmetric O-H stretching, N-H stretching
	1263	74	Amide III of Proteins, Phosphodiesters	N-H in-plane bend, C-N stretching, asymmetric PO ₂ stretching
	1084	70	Nucleic acids, Phosphodiesters	Symmetric PO ₂ ⁻ stretching, C-O stretching
Pancreas v NCS	3277	100	Proteins (Amide A), Nucleic acids	Symmetric O-H stretching, N-H stretching
	1529	84	Amide II of Proteins	N-H bending, C-N stretching
	1636	71	Amide I of Proteins	C=O and C-N stretching, N-H bending
	1288	67	Amide III of Proteins, Phosphodiesters	N-H in-plane bend, C-N stretching, asymmetric PO_2^- stretching
	2784	64	Lipids	C-H, CH ₂ stretching